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FMS 2+

Fluorescence Monitoring System Manual

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Fluorescence Monitoring System Manual

by Hansatech Instruments Ltd

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Introduction

1 Introduction

The FMS 2+ field-fluorescence monitoring system is a versatile pulse-modulated instrument, designed to measure chlorophyll fluorescence emission from samples engaged in photosynthesis under ambient light conditions in the field.

It consists of a control unit housing all of the electronics, optics and light sources. A custom fibre optic interface module links the control unit to the sample which is oriented in a leaf-clip.

The system may be operated in several different modes: serial connection to a Windows PC enables real-time instrument control and data presentation. Captured data is simultaneously presented as a real-time chart recorder emulation and parameters- only format for easy identification of key experimental events. This “PC” mode of operation is suitable for development of complex protocols which may be programmed into the instrument using the simple “drag and drop” editor to generate user-defined scripts. These scripts automate the execution of experiments, allowing complex protocols involving many control events to be operated with the same ease as single control event measurement such as Fv/Fm.

Once programmed, the FMS 2+ can be used as a stand-alone fluorometer in either laboratory or field situations, running off internal batteries with all measurement data and calculated parameters saved to integral protected memory. The unit can store up to six experimental

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protocols, any one of which may be accessed and executed using the built-in menu system. When data collection is complete the FMS can be downloaded to the Windows software for full analysis in the laboratory. This document aims to briefly cover the measurement principles used by FMS 2+, optimum hardware configuration, system operation and data handling with the “Modfluor” and “Parview” software packages. The document is separated into three sections:

- Section 1: Measurement Principles and Hardware.
- Section 2: Software installation, instrument operation and data handling.
- Section 3: Stand-alone instrument operation and data upload.

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Section 1: Measurement Principles and Hardware

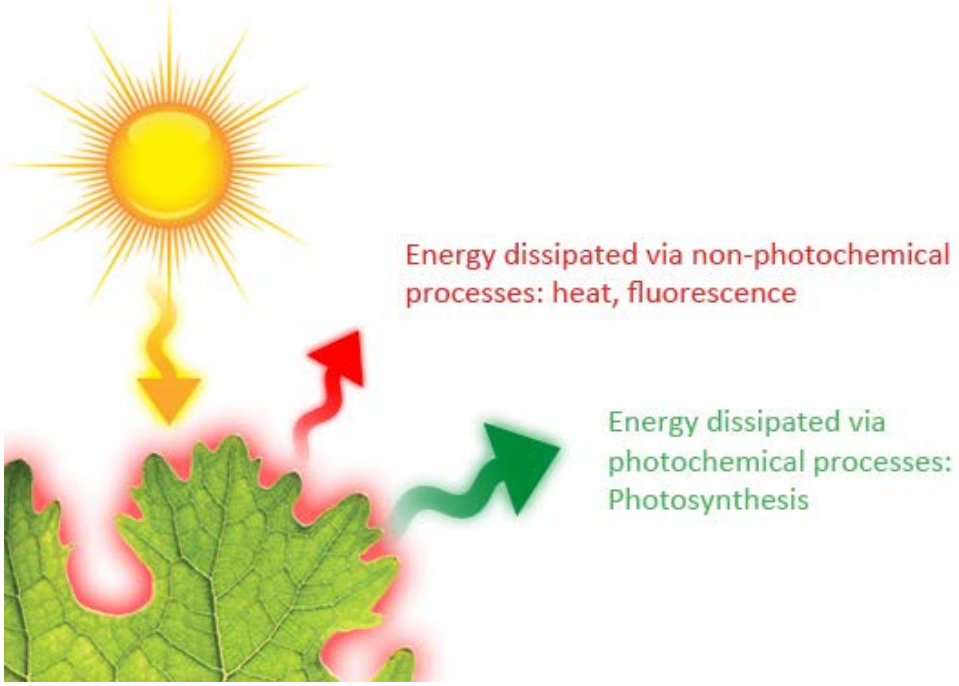
2 Section 1: Measurement Principles and Hardware

2.1 Introduction

2.1.1 What is chlorophyll fluorescence?

These processes utilise absorbed energy for photochemistry, during which electron donation from pigment to an acceptor molecule occurs. Such processes direct energy for the chemical work involved in photosynthesis.

Non-Photochemical Processes:
These processes dissipate energy from the photosynthetic apparatus in a manner which does not drive photosynthesis. Energy is usually re-emitted from the sample in the form of infra-red radiation (heat) and red / far-red radiation which is known as chlorophyll fluorescence.



The competition between these processes for absorbed energy ensures that a reduction in the rate of one process will be associated with a corresponding increase in the rates of competing processes e.g. a reduction in the dissipation of energy by
photochemistry will be reflected in an increase in energy dissipation by nonphotochemical processes such as heat production and chlorophyll fluorescence.

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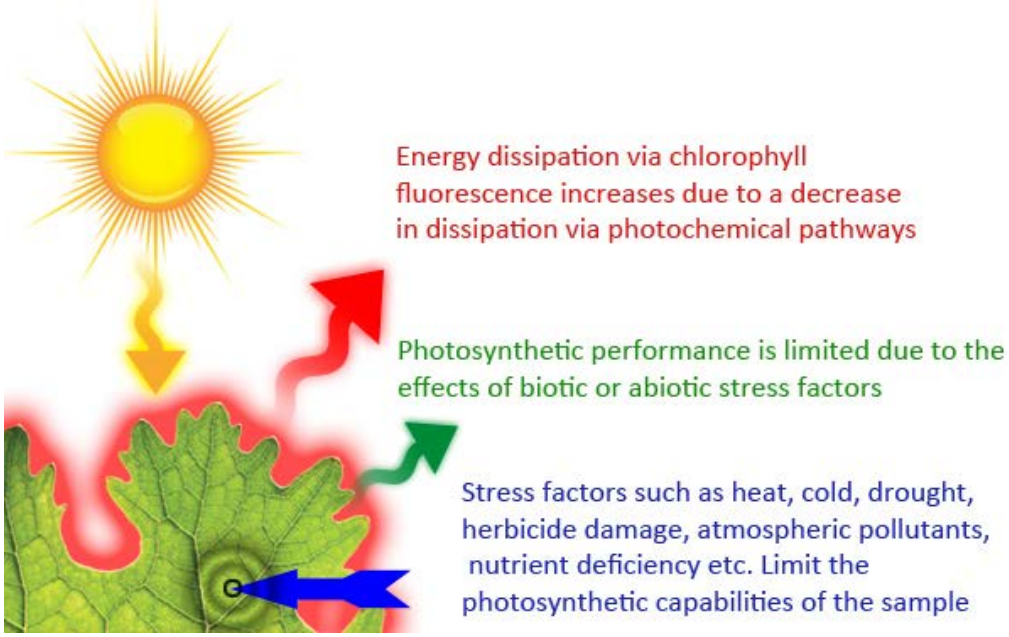

Consequently measurement of changes in the extent of fluorescence emission with FMS 2+ can be used to infer information about changes in the efficiency of light use for photochemistry (Reviewed in Krause and Weis 1991 and Govindjee 1995)
Since the work of Emerson et al. (1932) it has been widely accepted that there are two separate photochemical steps during photosynthesis which are associated with different groups of pigments; these groups have become known as photosystem I and photosystem II. In practice differences in the functionality of photosystem I and II mean that at least 95% of the chlorophyll fluorescence signal observed under physiological temperatures is derived from chlorophyll molecules associated with
photosystem II (PSII). Hence the fluorescence signal detected by FMS reflects changes in the efficiency with which absorbed light is used for PSII photochemistry.

2.1.2 How does the FMS measure chlorophyll fluorescence?

At physiological temperatures peak fluorescence emission occurs in the red region of the spectrum (685 nm) and extends into the infra-red region to around 800 nm (Krause and Weiss 1984) (Figure 1-1).

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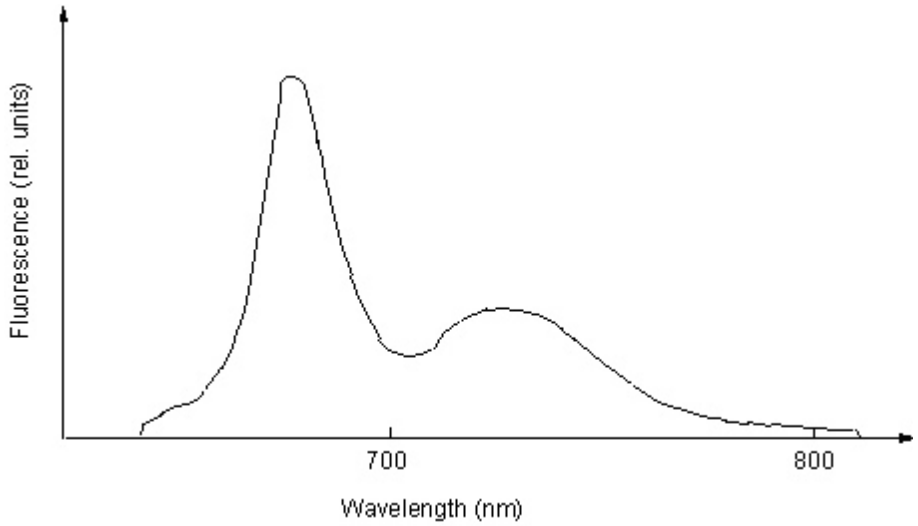

Fig. 1-1. Room temperature chlorophyll fluorescence emission spectra. (Adapted from Krause and Weis 1984)

Accurate determination of fluorescence requires a detection system which is sensitive to fluorescence wavelengths yet blind to the actinic (or excitation) light which drives photochemistry. In continuous excitation instruments such as the Plant Efficiency Analyser signal discrimination is achieved using a custom designed 650 nm actinic light source to drive photochemistry; optical filtering of the detector prevents detection of non-fluorescence wavelengths of light. Unfortunately such systems must be shielded from ambient light during use, otherwise the red / far-red component of day-light will be super-imposed on the fluorescence signal.

This problem is avoided by the FMS 2+ as it uses an electronic method, commonly referred to as modulated fluorometry, to separate actinic light from the fluorescence signal (Ögren and Baker 1985). During measurement the tissue sample is exposed to a pulsed amber (or optional blue) LED light source (the modulating beam) which induces a pulsed fluorescence signal from the sample under conditions where ambient light is excluded (A, Figure 2). When ambient light is applied the optical filtering of the FMS 2+ allows three types of light signal to reach the detector:

- ambient light of fluorescence wavelengths,
- non-pulsed fluorescence signal induced by the ambient light,
- and pulsed fluorescence signal induced by the modulating beam (B, Figure 1-2).

Addition of ambient light changes the physiology of the sample, altering the proportion of energy that is re-emitted as fluorescence. Thus the magnitude of the peaks of the pulsed fluorescence signal that are induced by the modulating beam will reflect changes sample physiology. It is these signals that the rapid pulse peak tracking electronics amplify and measure; ambient and non-pulsed fluorescence signal are discarded (C, Figure 1-2).2.

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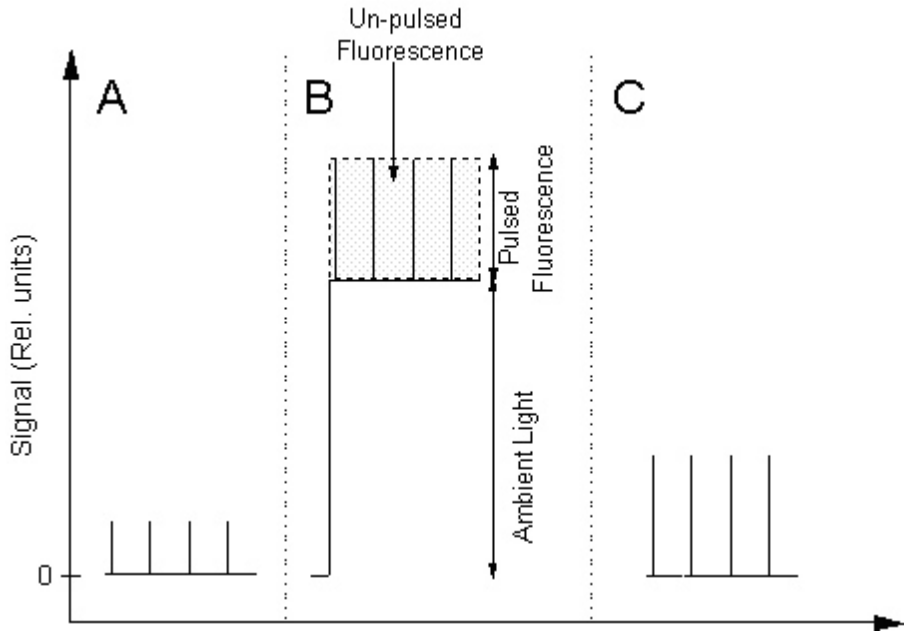

Fig. 1-2. Fluorescence signal detected by the FMS under modulating beam only conditions (A), under actinic light plus modulating beam before (B) and after (C) the rapid pulse peak tracking electronics.

The pulsed fluorescence that is induced by the modulation beam is effectively used to probe the efficiency of light-use for photosystem II photochemistry. Clearly it is important that the measuring technique itself does not induce changes in the physiology of the sample under investigation.

Exposure of a sample to a powerful pulsed light source may be expected to introduce a significant artifact into the measurement. However, FMS has been specifically designed to avoid such problems by using a modulating beam of very short duration pulses (1.8 µsec) with long off period between pulses. The net result is that the integrated amount of radiation incident upon the sample from the modulating beam is very small (<0.05 µmolm-2s-1), too small to induce significant physiological change in the sample (Schreiber et al. 1986), yet powerful enough to provide a reliable signal for fluorescence analysis.

The interpretation of fluorescence signals has developed considerably since the early experiments of Kautsky and Hirsch (1931) with several techniques employed to determine information about the photosynthetic performance of PSII (Baker 1991). The FMS hardware and software has been designed to incorporate these techniques into simple routines which store appropriate fluorescence measurements and automatically calculate key parameters. The principles and procedures behind these are discussed in relation to instrument control with the FMS Windows software and scripting language (Section 2).

Several “non-specific” parameter functions have also been included. The aim of these is to give sufficient flexibility to experiment and develop novel protocols. These are

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also discussed in Section 2.

2.2 Hardware

2.2.1 The Multi-Charger

The FMS 2+ is supplied with five lead-acid batteries which will probably need to be charged before the system can be made operational. A maximum of four batteries may be recharged simultaneously with the multi-charger unit.

The multi-charger should be placed in a flat, well ventilated area. Each battery inserts into a “slot” with the black upper surface facing out and the metal terminals facing forward to meet the charger contacts (Figure 1-3). The multi-charger has been designed so that the battery terminals and charger contacts can only meet in one orientation, so in normal use it should be virtually impossible to connect the battery incorrectly.

THE MULTI-CHARGER CAN CHARGE A MAXIMUM OF FOUR BATTERIES SIMULTANEOUSLY. THERE IS NO PROBLEM OPERATING THE UNIT WITH LESS THAN FOUR BATTERIES. EACH BATTERY “SLOT” IS AN INDEPENDANT CHARGING CHANNEL AND THERE IS NO DANGER ASSOCIATED WITH TOUCHING EXPOSED CONTACTS DURING OPERATION.

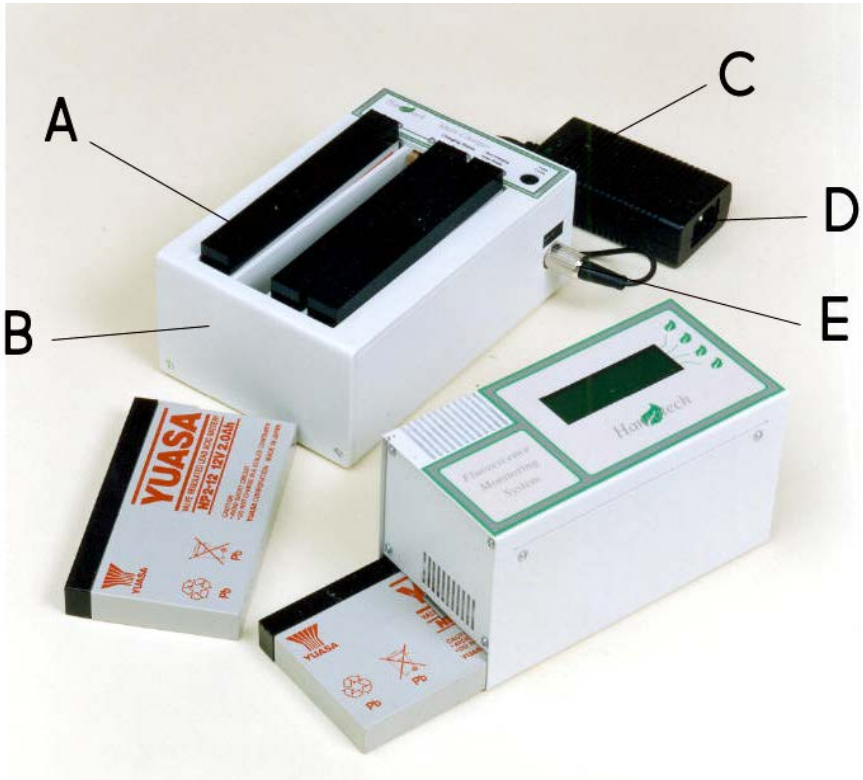

Fig. 1-3. The multi-charger unit (B) with batteries (A). Mains power is supplied from the mains adapter unit (C) which plugs into the side socket of the charger unit (E).

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The mains cable locates unit the front of the mains adapter unit (D)

Power is supplied to the multi-charger via the 15 V mains adapter. The 3-pin DIN output plug should be connected to the socket that is mounted in the side of the multi-charger (E, Figure 1-3). Mains power is connected through the mains cable input in the front of the mains adapter (D, Figure 1-3).

After power-up the LED indicators on each of the charging channels will take several seconds to settle and read the status of the batteries. The LED indicators will display one of two possible states:

- Red Light - indicates a discharged battery drawing current at the maximum regulated rate.
- Green Light - indicates that the battery is at least 98 % fully charged. A trickle current of 50 mA keeps the battery topped up until use.

AFTER INSERTING DISCHARGED BATTERIES IT IS GOOD PRACTICE TO WATCH AS THE LED INDICATORS SWITCH TO RED TO ENSURE THAT THE BATTERIES ARE RE-CHARGING. OLD OR DAMAGED BATTERIES WILL NOT DRAW CURRENT AND THUS THE INDICATORS MAY REMAIN PERMANENTLY GREEN.

Fully discharged batteries will require approximately 8 hours to reach maximum capacity. It is therefore advisable to run the charger over-night. Once the batteries are charged the multi-charger may be switched off and the batteries removed.

2.2.2 Inserting / Removing Batteries From the Control Unit

One of the batteries may be placed in the base of the FMS 2+ carrying bag; a second should be inserted into the FMS 2+ control unit. This is done by:

1. Orientating the battery at 45° to the battery compartment in the FMS 2+ control unit. The black-upper surface of the battery should face forward with terminals facing into the battery slot,
2. Lift the battery compartment catch with the corner of the battery,
3. Push the corner of the battery into the slot, re-align and fully insert (Figure 1-4).

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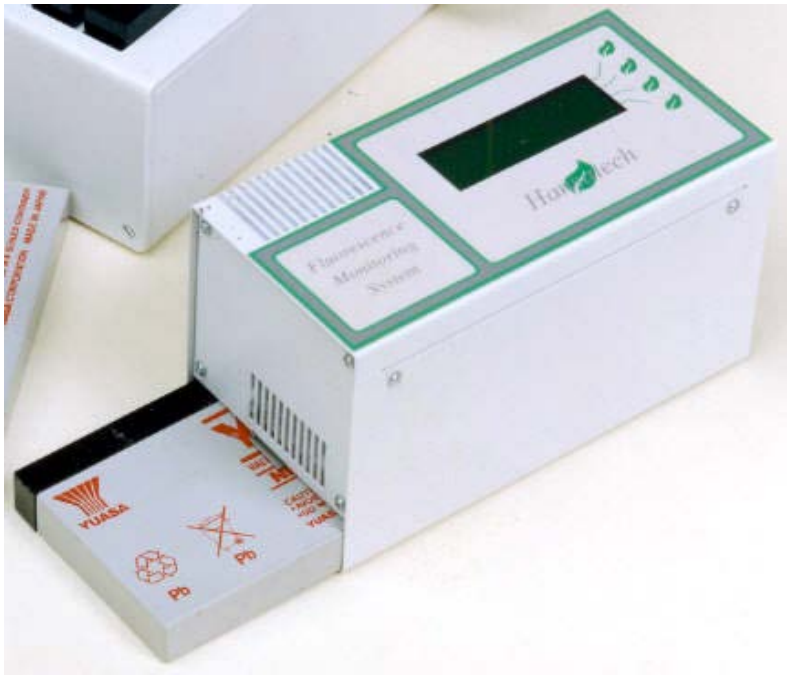

Fig. 1-4. Inserting the battery into the FMS 2+.

The spring loaded contacts inside the battery compartment will automatically make contact with the battery terminals whilst the catch on the end of the compartment should firmly hold the battery in position.

THE CONTROL UNIT CANNOT BE DAMAGED BY INSERTING THE BATTERY IN THE WRONG ORIENTATION AS THE BATTERY TERMINALS AND CONTROL UNIT CONTACTS WILL NOT MEET. HOWEVER THE INSTRUMENT WILL NOT OPERATE UNTIL THE BATTERY IS RE-INSERTED CORRECTLY.

The rate of battery discharge during operation is determined principally by the use of the LED Light Source. A high rate of pulsing or pro-longed use of high output actinic light will drain the battery relatively quickly. The following estimates of battery capacity and re-charge period should be considered when planning experiments:

- 5 hours of continuous actinic illumination of 3,500 µmol m-s s-1
- Approx 4,000 saturating pulses of 13,000 µmol m-2 s-1 at the sample surface

The electronics are designed to maintain constant light intensities throughout the discharge period of the battery. Warning messages are generated when the battery capacity reaches the threshold where this is no longer possible in both stand-alone (External User Interface) and PC operation (Section 2). At this point it is recommended that the instrument is switched off and a fresh battery installed.

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Battery removal in the field is simply a matter of switching the instrument off, lifting the catch on the battery compartment and tilting the control unit until the battery slides out. The instrument memory is protected by an internal capacitor. As a result data should not be lost even after several days without a battery.

IT IS ALWAYS GOOD PRACTICE TO UPLOAD DATA TO A PC AT THE END OF EACH DAY. THIS ALLOWS MULTIPLE COPIES OF THE DATA TO BE GENERATED FOR BACK-UP PURPOSES.

2.3 Connections to the FMS 2+ Control Unit

2.3.1 Electrical Connections

The following electrical connections can be made to ports in the right-hand side panel of the FMS 2+ control unit (Figure 1-5). It should be noted that for field-portable operation without a PC the instrument may be operated without connection to these sockets.

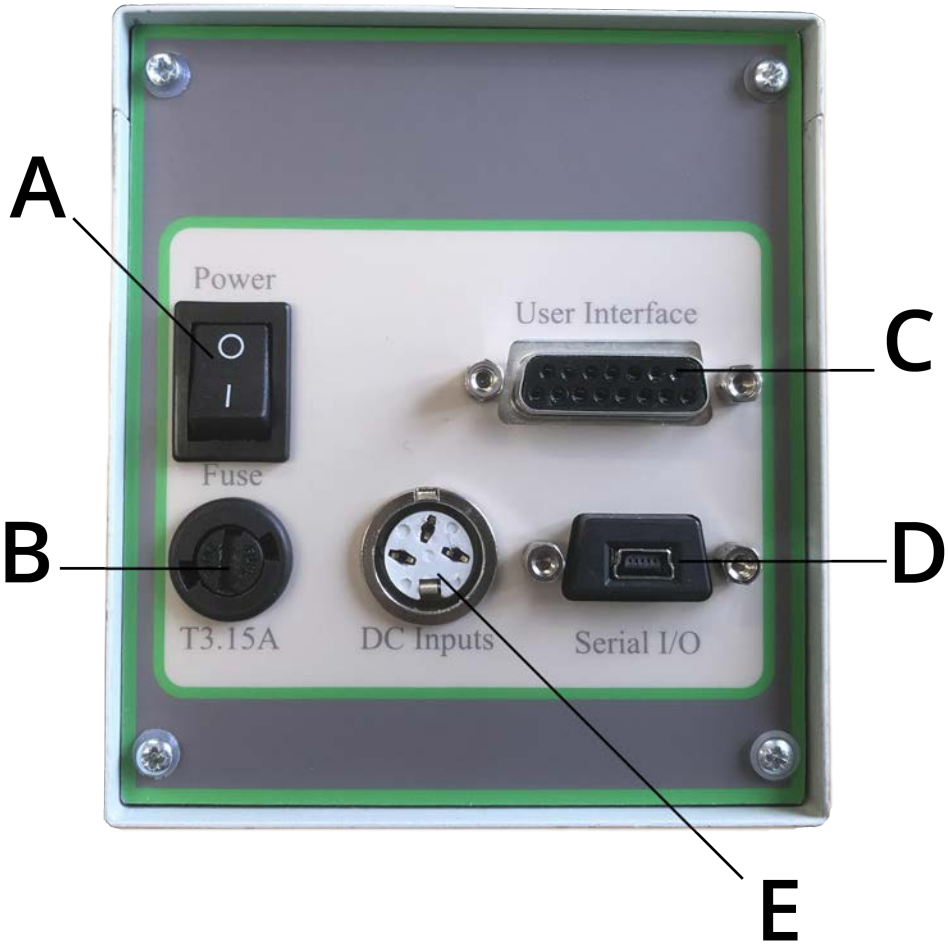

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Fig. 1-5. The right side panel of FMS 2+ contains the power switch (A), Fuse (B), "User Interface (C), Serial I/O (D) and DC input socket (E).

- Serial I/O socket (D) - Must be connected to a USB port via the supplied USB cable for computer controlled operation or data upload. Not required for "Local mode" use without a PC.
- DC Inputs socket (E) - Available for connection of external DC power supplies

such as:

12 V external battery via optional battery cable,
12 V output from optional mains adapter,
15 V mains adapter for re-charging the on-board battery without multi-charger.

- User Interface socket (C) - Provides access to four digital and one digital to analogue control line for optional control over external devices (Section 2).

2.3.2 Optical Connections

The control unit houses the light sources and detector which must be optically linked to the sample under investigation. This is achieved via a custom fibre-optic which locates onto the front of the control unit via the optic interface module.

When the instrument is first unpacked the interface module should be covered by a blanking plate which protects the optics in transit. The plate is removed by carefully pulling it from the control unit. The fibre-optic cable is correctly positioned on the optic interface module by inserting the small key-way block of the fibre-optic termination (A, Figure 1-6) into the corresponding groove in the optic interface module (B, Figure 1-6). The cable is secured by carefully threading the collar which surrounds the fibre-optic termination onto the optic interface module until it is hand-tight (D, Figure 1-6). This procedure ensures that the light guides within the fibre-optic are automatically aligned with the appropriate light source or detector within the control unit.

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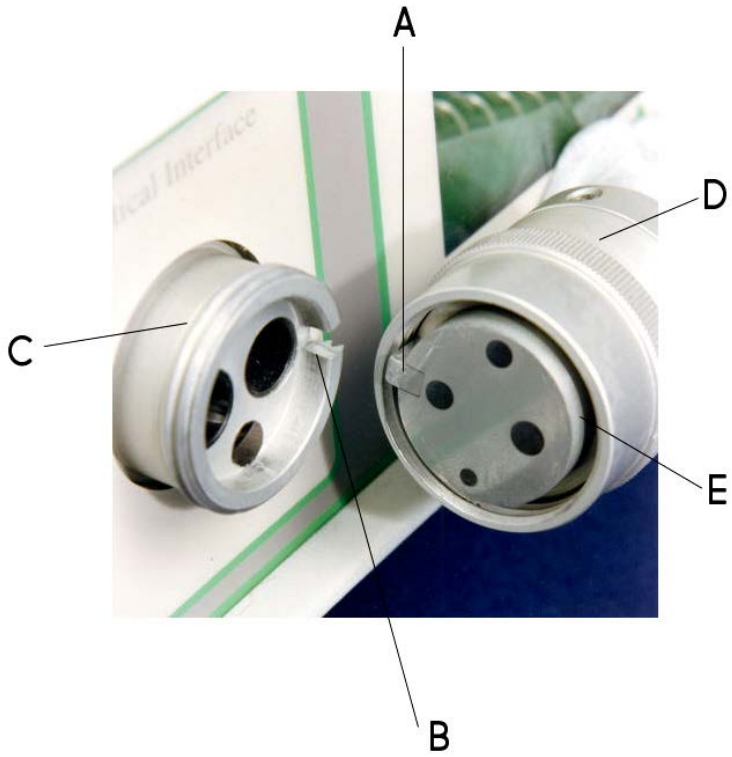

Fig. 1-6. A small key-way (A) on the end of the fibre-optic (E) locates into a complimentary groove (B) in the optics interface module of the FMS control unit (C). The fibre-optic is secured by gently threading the ferrule surrounding the fibre (D) clockwise onto the thread of the optics interface.

THE FIBRE OPTIC CABLE IS DESIGNED TO BE AS RUGGED AS POSSIBLE. HOWEVER ITS CORE IS MADE OF THOUSANDS OF INDIVIDUAL GLASS FIBRES WHICH CAN BREAK IF THE CABLE IS DROPPED OR FLEXED THROUGH TIGHT ANGLES. DAMAGE TO THE CABLE WILL SIGNIFICANTLY IMPAIR THE PERFORMANCE OF FMS.

Finally the sample end of the fibre-optic should be positioned close enough to the tissue under investigation to detect a strong signal. Typically it will have to be within 1.5 cm of the tissue, usually orientated in a manner which minimises fibre- nduced shading. A pair of leaf-clip systems are available for this purpose.

2.4 FMS 2+ Leaf-Clips

2.4.1 The PAR / temperature leaf-clip

The PAR / temperature leaf-clip is designed for measurements made under ambient light conditions. It consists of a sprung upper section which gently grips the tissue in a gentle clamping action (C, Figure 1-7A). A grooved neck mounted at 60° to the plane of the sample (B, Figure 1-7A) accommodates the fibre-optic (A, Figure 1-7A) which is slid into position. Marked graduations on

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the neck can be aligned with graduations on the fibre-optic termination to reference its position for future work, a retaining screw (B, Figure 1-7A) locks it into position throughout the experiment. The rest of the fibre may be looped over the leaf-clip and hooked to the rear of the clip to help support its weight.

An electrical connection to the “Leaf-clip” socket on the front panel of the control unit enables use of the remote trigger switch (F, Figure 1-7A) to activate / abort measurements in “Local” mode (Section 3) and connect on-board thermocouple and
light sensor (C, Figure 1-7A) to the control unit. The leaf-clip may be held in the hand if multiple samples are being studied or mounted on a standard tripod mount (E, Figure 1-7A) via a thread in the lower clip section for fixed-position work.

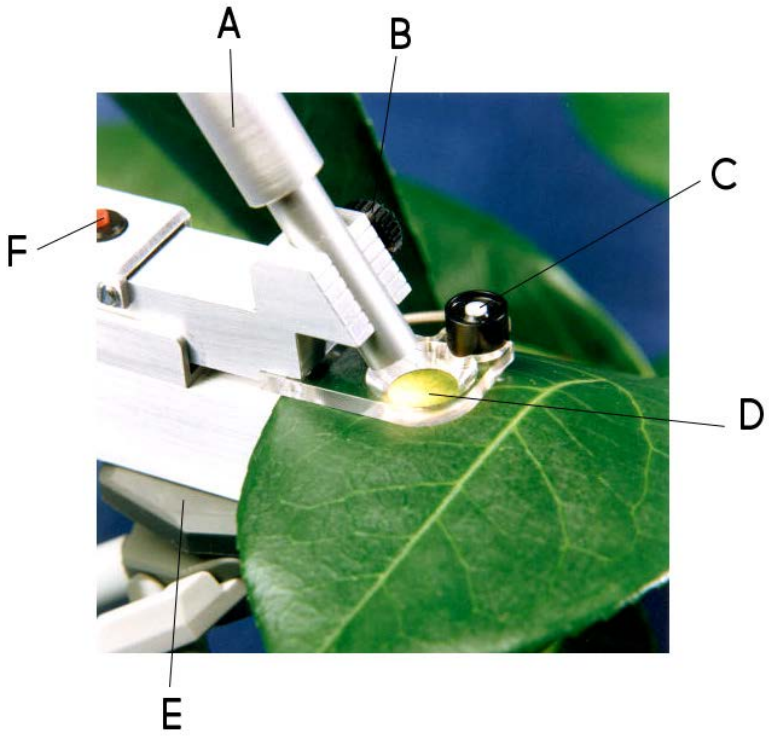

Fig. 1-7A. A broad leaf sample under investigation in the PAR / temperature leaf-clip. The fibre-optic (A) is retained in the grooved neck by a screw (B) whilst the sample (D) is gripped between the upper and lower clip sections. Simultaneous measurements of ambient light are made with the co-sine corrected PAR sensor (C) and sample temperature monitored by a thermocouple in contact with the underside of the sample at (D). The leaf-clip is supported by a tripod mount (E) and measurements initiated with the remote trigger button (F).

The PAR sensor has been designed for both recording of ambient light intensities during fluorescence analysis and measurement of FMS actinic and saturating light sources during instrument set-up. It may be mounted in one of two positions on the leaf-clip:

- Upper side mount - prevents shading by broad leaf samples, suitable for ambient light measurement during analysis (Figure 1-7A).

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- Lower front mount - suitable for ambient light measurement during analysis of narrow leaves which do not shade the sensor (Figure 1-7B). - aligns sensor level with sample for calibration of fibre-optic light output when upper section slid forward (see below).

Fig. 1-7B. The PAR sensor (A) located in the cup on the lower section of the clip. This position is suitable for actinic light calibration (described below) or simultaneous PAR monitoring during analysis of narrow samples (B) which do not cover the sensor in this position.

Prior to commencing an experiment it is often useful to measure the output of the saturating LED and reference the intensities of actinic light settings that are to be used. In order to do this the fibre-optic and PAR sensor must be placed in the same geometry and distance that the sample and fibre would occupy during fluorescence analysis. This can be achieved simply by gently sliding the upper section of the clip down and forward through its full range of travel so that the PAR sensor is under the fibre's field of view. The various light-sources can then be activated and readings noted from the software display. When light calibration is complete the upper section should be pushed down and back over the full range of travel to return the fibre to its measurement position.

Sample temperature is also monitored simultaneously with PAR and fluorescence. A thermocouple which stands slightly proud of the lower section is designed to maintain contact with the lower surface of the sample.

IT IS IMPORTANT TO NOTE THAT THE THERMOCOUPLE IS PROUD OF THE

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LOWER LEAF-CLIP SECTION. THEREFORE CARE SHOULD BE TAKEN TO AVOID SNAGGING THE THERMOCOUPLE WHEN SAMPLES ARE BEING INSERTED OR REMOVED.

2.4.2 The Dark-Adaptation Leaf-Clip and Fibre-Optic Adapter

A separate leaf-clip system has been developed for situations where ambient light is to be excluded from the sample during measurement. This is suitable for experiments requiring dark-adapted measurements e.g. screening applications measuring Fv/Fm or situations which require adaptation of tissue to standardised doses of actinic light.

The first step in the process is to attach the fibre-optic adapter (Figure 1-8) to the sample end of the fiber optic. This is done by sliding the fibre-optic (A, Figure 1-8) into the adapter mounting collar (B, Figure 1-8) and securing it in position by tightening the side mounted retaining screw (C, Figure 1-8). The adapter / fibre-optic assembly can then be sealed into the top of the dark-adaptation clip (D, Figure 1-8) and the position of the fibre-optic fine-tuned by unthreading the retaining screw in the adapter collar and sliding the fibre-optic into the desired position before locking it by re-tightening the collar screw.

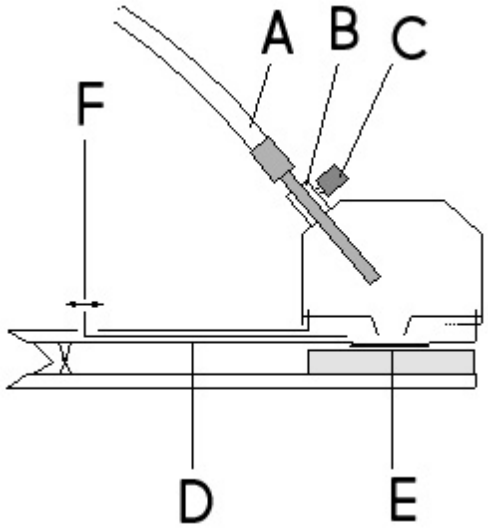

Fig. 1-8. The dark-adaptation leaf-clip and fibre-optic adapter. The fibre-optic (A) is held in the adapter mounting collar (B) by the side-mounted retaining screw (C). The adapter / fibre combination locate onto a dark-adaptation clip (D). The sample (E) is supported on a foam pad and exposed to the FMS when the sliding shutter (F) is opened.

THE GEOMETRY OF THE FIBRE-OPTIC WILL DETERMINE HOW EVENLY THE SAMPLE WILL BE ILLUMINATED AND THE STRENGTH OF FLUORESCENCE SIGNAL RECEIVED. IT MAYBE USEFUL TO MAKE SEVERAL TRIAL MEASUREMENTS WITH SAMPLE IN ORDER TO OPTIMISE THE SYSTEM FOR

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EACH PARTICULAR APPLICATION.

When the fibre-optic has been locked into position the dark-adaptation clip can be removed and clipped into the experimental sample. The tissue is dark-adapted by sliding the shutter plate forward (F, Figure 1-8). After a suitable period the tissue should be dark-adapted and ready for measurement. The adapter / fibre-optic assembly can then be sealed over the dark-adaptation clip and the shutter blade opened to expose the tissue to the FMS. The adapter ensures that the sample is completely shielded from ambient light throughout the measurement.

IN RAPID SCREENING MEASUREMENTS IT MAY BE MORE EFFICIENT TO DARK-ADAPT A NUMBER OF SAMPLES SIMULTANEOUSLY WITH SEVERAL CLIPS. REPETITIVE MEASUREMENTS CAN THEN BE MADE BY MOVING THE FIBRE-OPTIC FROM CLIP TO CLIP.

2.5 Hardware Maintenance

The hardware is designed with a minimum of consumable components. The most likely components to require attention are the LED Light Source, battery and fuses. This section details the procedures to replace both of these items.

2.5.1 Fuse Replacement

The instrument is protected by two fuses. An external fuse is located beneath the “On / Off” switch on the right-hand side panel. This protects the instrument from external 12 V supplies, such as car batteries, that have been connected through the “DC input” socket. If this fuse fails the external battery and associated wiring to the “DC input” socket should be checked before replacement. The fuse is replaced using a flat blade screw driver to open the fuse cover by pushing in and gently turning anti-clockwise. The old fuse should be replaced with a 3.15 A anti-surge fuse.

An internal fuse protects the circuitry from the on-board power supply and mains adapter connection to the 12 V “DC input” socket. The fuse is accessed by removing the right-hand side panel (panel with “On /Off” switch) with a screw driver. The fuse is located under a label in the black acetal battery block at the bottom of the control unit. It can be gently levered out with a flat blade screw-driver and replaced with a 5 A medium-blow 20mm fuse.

A BLOWN FUSE MAY BE A SYMPTOM OF A MORE SERIOUS FAULT WITH THE INSTRUMENT. HANSATECH INSTRUMENTS LTD SHOULD BE CONTACTED IF A REPLACEMENT FUSE BLOWS WITHIN A SHORT PERIOD OF TIME.

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2.6 The External User Interface

Measurements with the FMS can be triggered from external devices via one of four digital control lines which are connected to the User Interface Socket of FMS (B, Figure 1-5). The Digital IN connections are presented below:

Channel Number:	Pin Connection:
0	4
1	5
2	6
3	7
Ground	3 and 11

The FMS may also be used to trigger measurements using the Digital OUT connections:

Channel Number:	Pin Connection:
0	12
1	13
2	14
3	15
Ground	3 and 11

Analogue output channel connections are:

	Pin Connection:
Analogue Output	10
Analogue Ground	2

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Section 2: Software Installation, System Operation, Data Handling

3 Section 2: Software Installation, System Operation, Data Handling

3.1 FMS Software

The FMS is supplied with two Windows software applications:
“Modfluor32” - Instrument control and trace data presentation program.
“Parview” - Parameter data upload and viewer program.

All of the software and on-line help documents are supplied on a USB drive. This also contains reference lists and links through to the Hansatech Instruments web site where updated versions of software may be downloaded as and when they are released.

3.1.1 Installing FMS Software

The installation is launched from the USB drive supplied with the unit. Follow the on-screen prompts in order to install Modfluor32 and Parview on your PC.

The installation automatically sets up a program group called “Hansatech Instruments”. This contains a sub directory with icons for running both the “Modfluor” and “Parview” applications, an “Uninstall” program and links to the help documentation.

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3.1.2 Connecting the FMS to Modfluor Software

3.1.2.1 Adjusting COM port and Latency Settings

The software is only operational when the FMS is set to a port number from 1-8. This is very simple to view/change if needed by following the instructions below.

1. Enter your PC's 'Device Manager', this can be located via the control panel.

2. Expand the "Ports (COM & LPT)" section of this menu by pressing the down arrow located to the left of it. The FMS will be present as "USB Serial Port" (picture below)

3. Right click "USB Serial Port" and select "Properties", navigate to the "Port Settings" tab. Click "Advanced"

4. There will be a drop down box containing the COM port number. If this is numbered 1-8 then this does **not** need to be changed. Make a note of this COM number and continue onto the next step. If however, this does need alteration click the drop down box and select a COM port number from 1-8 which is not currently in use. Make a note of this COM number.

5. In this same window under "BM options" there is a drop down box labeled "Latency Timer (msec). Change this value to 1.

6. Press OK, then OK again in the properties window (you may be asked to restart the PC to save these changes).

7. If the COM port selected is COM1 then ensure the instruments is in "PC mode" and initiate the software.

8. Locate (or create) the "Modluor" shortcut on your desktop. Right click this and select "Properties"

9. The writing in the "Target" box will read "C:\Program Files\Hansatech Instruments Ltd\FMS\Modfl32.exe" . In order to run the software, the port number will need to be added to this. Scroll to the end of the line after the last quotation mark, click here. You will need to enter a **space** followed by /x (whereby the x is the COM port number you selected previously).

For example the line for COM port 2 would look like: "C:\Program Files\Hansatech Instruments Ltd\FMS\Modfl32.exe" /2

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10. Click Apply then OK

11. Ensure the instruments is in "PC mode" and initiate the software.

3.1.3 Executing “Modfluor” and “Parview”

Both programs may be run without connection to the FMS. However in most circumstances they will be used to communicate with the control unit. The USB serial cable must be connected to the PC and the control unit should be switched on and “PC” button pressed to select the “PC-mode” of operation (PC Mode of Operation Under Modfluor).

During initialisation the software tests the communications link. If the software is unable to communicate a prompt box will be generated requesting to use “demonstration mode” or a re-test.

Most commonly communications problems occur because the control unit is not switched on or set to “PC-mode”.

3.2 Operating the FMS

The FMS is designed to accommodate two different operating modes:

• “PC mode” - the control unit is linked to the PC with commands issued from “Modfluor” under “real-time” control from the PC
• “Local mode” - (Section 3) the control unit is used autonomously with measurement commands issued via pre-loaded scripts accessed via the four-button keyboard on the control unit display panel.

When the instrument is first used it will only contain two default scripts for measurement of Fv/Fm and PSII. It is therefore sensible to use “PC mode” to test the system and generate further scripts which are more suited to individual applications. “Local mode” operation is discussed in Section 3.

3.2.1 “PC Mode” Operation Under Modfluor

Once the “Modfluor” program has successfully communicated with the control unit, it loads the data recording screen.

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3.2.2 The Recording Screen

The default recording screen which is generated when the software is first initialised consists of four areas:

- The data recording area (A, Figure 2-1),
- Instrument monitor bar (B, Figure 2-1),
- "Drop-down" menu bar (C, Figure 2-1),
- Instrument tool bar (D, Figure 2-1).

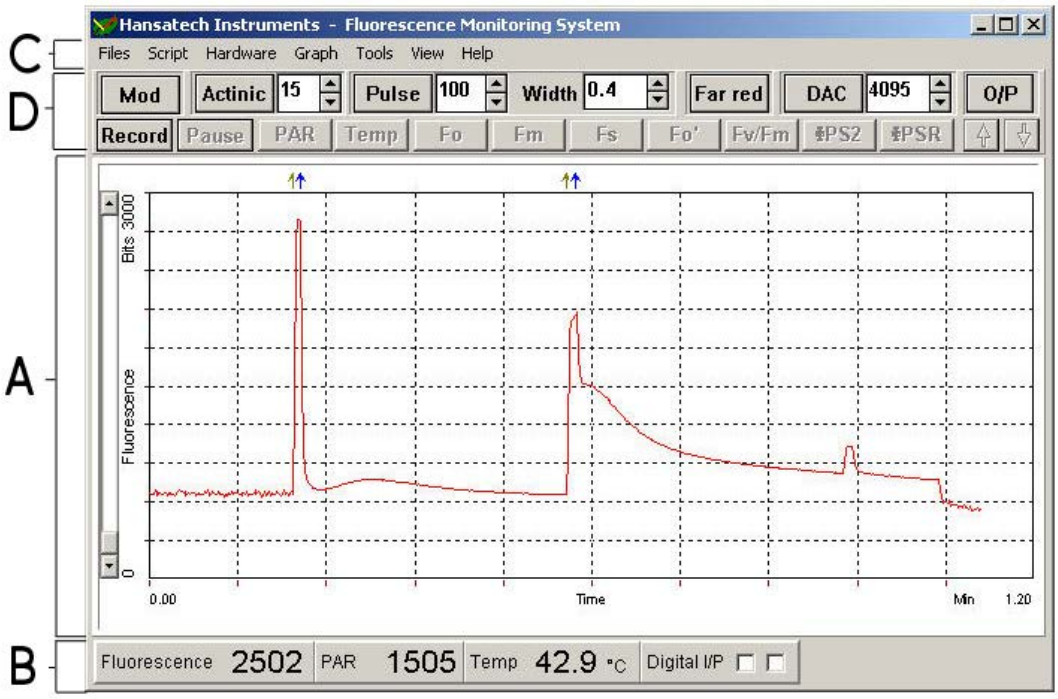

Fig. 2-1. The data recording screen contains the data recording area (A), instrument monitoring bar (B), "drop-down" menu bar (C) and instrument toolbar (D).

The data recording area is a graphic display area where acquired fluorescence data expressed as output from the analogue to digital (A/D) converter in bits (Y-axis) is plotted against time (X-axis) to give a "real-time" chart recorder emulation. Both axes may be scaled either automatically to display all data or manually to zoom regions of particular interest via the Graph function in the "drop-down" menu (Graph Functions). The maximum range of the A/D converter is 4095 bits; the plot area above this value is greyed-out to indicate that any output above 4095 bits is overscale (Testing the FMS).

Automatic scrolling of the plot ensures that data collection is continuous up to a maximum length of approximately 50 minutes. The PAUSE button in the instrument toolbar may be used to suspend data collection, operating as the computer equivalent of lifting the pens off a chart recording. If

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this function is used to limit data logging to specific periods of interest much longer experimental protocols may be run.

Data is digitally displayed throughout the experiment on the instrument monitoring bar (B, Figure 2-1). Fluorescence information is continuously up-dated even when the PAUSE function is used to suspend data logging, allowing estimation of steady state fluorescence prior to resumption of logging during a measurement. Data from external devices such as the PAR and temperature leaf-clip are also presented on the monitoring bar along with information relating to the status of digital control lines. In the event that external devices are not being used it is possible to switch off the monitoring bar via a toggle switch in the View function of the "drop-down" menu bar (View Functions).

Software configuration, file handing and instrument programming facilities are grouped according to function on the "drop-down" menu. They are accessed via the mouse or accelerator keys.

Example:

Files **S**cript **H**ardware **G**raph **T**ools **V**iew **H**elp
Graph functions are opened by pressing ALT **G** together.

When a group is selected a sub-menu of related functions is opened. The required function can be selected with accelerator keys (Key) or by moving the blue-highlight with UP and DOWN cursor keys to the desired function and pressing RETURN.

Navigation to other the sub-menus is achieved with LEFT and RIGHT cursor keys or mouse. Specific details of each of the menu function are given in Drop Down Menu Functions.

Direct control of FMS light sources and FMS measurement operation is made through the instrument tool bar (D, Figure 2-1). Light sources are activated by clicking in the appropriate toggle button with the left hand mouse button. The modulation beam, actinic and far-red light sources remain illuminated (indicated by an indented button) button until their respective buttons are re-pressed. The extent of actinic illumination is determined by the numeric value (0 to 50) in the edit box located next to the "Actinic" button (The Actinic Light Source).

CHANGES IN THE ACTINIC OUTPUT LEVEL ONLY TAKE EFFECT WHEN THE ACTINIC SOURCE IS INITIALISED. THE ACTINIC LIGHT MUST BE TOGGLED OFF AND THEN ON TO EFFECT AN INTENSITY CHANGE (THE ACTINIC LIGHT SOURCE).

The saturation-pulse is activated each time the "Pulse" button is pressed. The button remains indented during pulse application and automatically re-sets upon termination. Both pulse intensity (0 - 100) and duration (0.3 - 5.0 sec) are set from accompanying edit boxes on the toolbar (The Saturating Light Source).

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Measurement functions are only available whilst the FMS is recording. A fresh recording is started by pressing the “Record” button. This automatically triggers the modulation beam which is essential for fluorescence detection and disables menu functions that should be kept constant during the recording e.g. instrument configuration settings. Common fluorescence parameters may be logged via the various parameter buttons located on the instrument toolbar. Detailed descriptions of each measurement function are given in Measurement Functions. The digital values of each parameter are written to a “Parameter Window” within the software. Under default settings the “Parameter Window” is not displayed to save screen space. However the software display may be customised to suit individual requirements via the View function in the “drop-down” menus (View Functions).

Throughout the recording period individual FMS-driven illumination events are marked by arrows to mark light on and arrows to mark light off events. User events such as herbicide treatment etc. may be marked using the buttons on the instrument toolbar (External Device Interface). All markers tag corresponding data in the tabulated data output from FMS by colour-coding data for easy reference (Event Marker Colour).

The next stage in operating the FMS is to test the system with spare sample and configure the instrument to achieve optimum performance.

3.2.3 Testing the FMS

Before running an experiment several functional aspects of the instrument should be checked:

- The signal is sufficiently strong to be acceptable,
- The modulation beam does not change the physiology of the sample,
- The saturation pulse is sufficiently powerful to saturate the sample, and the maximum fluorescence yield is within the range of the instrument.

The following section briefly describes two experiments which can be run to verify each of these aspects on spare sample tissue.

1. Dark-adapt a piece of spare experimental tissue using the dark-adaptation clip or by covering the sample in the PAR / temperature clip.
2. Start a new recording by pressing the “Record” button on the instrument toolbar. The dark-adapted fluorescence yield should plot across the data recording area. This is the minimum fluorescence signal from the sample and should be within the range of 10 - 700 bits for reliable measurement. If the signal is too small the test recording should be stopped (re-press the “Record” button) and the sensitivity of the instrument adjusted by increasing gain (Config Settings) or by moving the fire optic closer to the sample. If the signal is too great (>700 bits) the instrument will probably over-scale when the saturating pulse is used. Instrument gain should be reduced. This effect can also be achieved by moving the fibre-optic away from the sample.

It is also important to check that the signal recorded in the experiment above remains constant. In the example below (Figure 2-2.) the dark-adapted fluorescence signal is around 500 bits (suitable

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for further measurement) but gradually increases with time. This indicates that the modulation beam setting is too high and that it is inducing a physiological effect on the sample. The most common symptom is a gradual rise in the fluorescence signal known as “variable” fluorescence which occurs as the modulating beam drives a significant amount of photochemistry (Fo Fluorescence Origin).

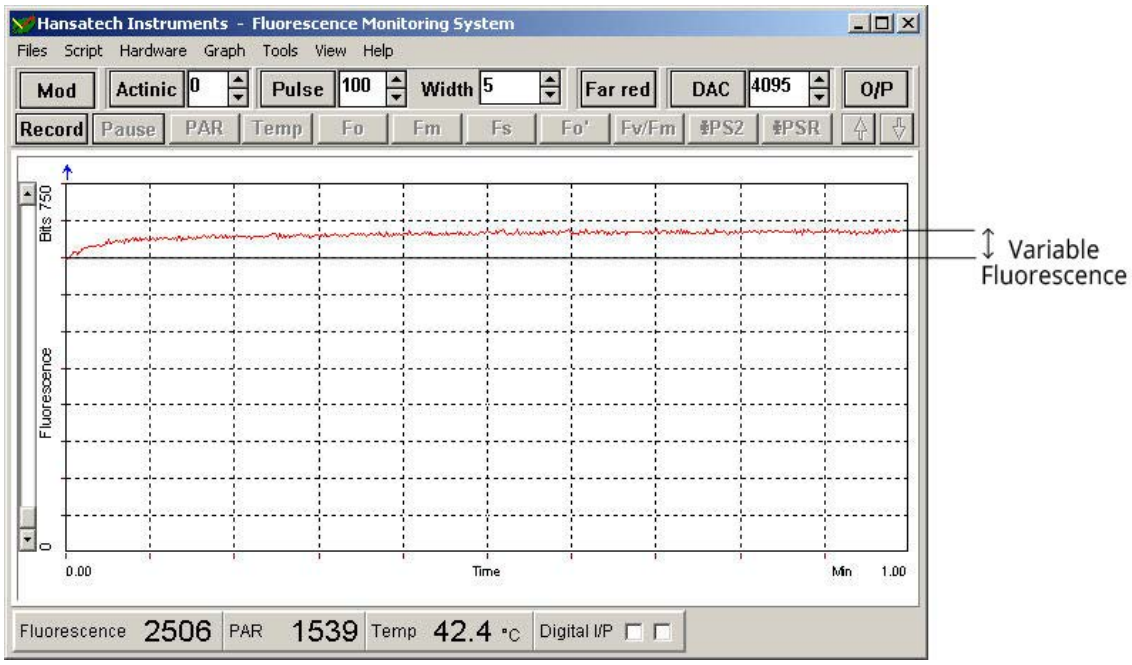

Fig. 2-2. The sample in this experiment is maintained in the dark under the modulation beam. The gradual rise in the fluorescence signal is attributable to the modulation beam driving significant PSII photochemistry. This increase in fluorescence is known as variable fluorescence.

This phenomena will cause an error in the estimation of several commonly referenced fluorescence parameters (Fo Fluorescence Origin). Corrective action requires a reduction in the frequency of the modulating beam by using a lower modulating beam setting (Config Settings) or a reduction in modulation intensity by increasing the distance between the sample and fibre optic.

It is also important to check that the instrument is correctly configured to record the maximum level of fluorescence when the saturation lamp is used. This may achieved with the following short experiment:

1. Dark-adapt a second piece of spare tissue for a minute of two.
2. Press the “Record” button to start a fresh recording.
3. Set the “Pulse” intensity to medium value in the edit box (e.g. 50 units) and initiate a pulse by pressing the “Pulse” button. After a further 30 seconds increase the pulse intensity by 10 units and pulse. Repeat this procedure incrementing the pulse intensity up to 100 units.
4. After the final pulse stop the recording and autoscale the trace (under the

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Graph function in the “drop-down” menu).

A typical trace from this test on a healthy leaf is shown below (Figure 2-3).

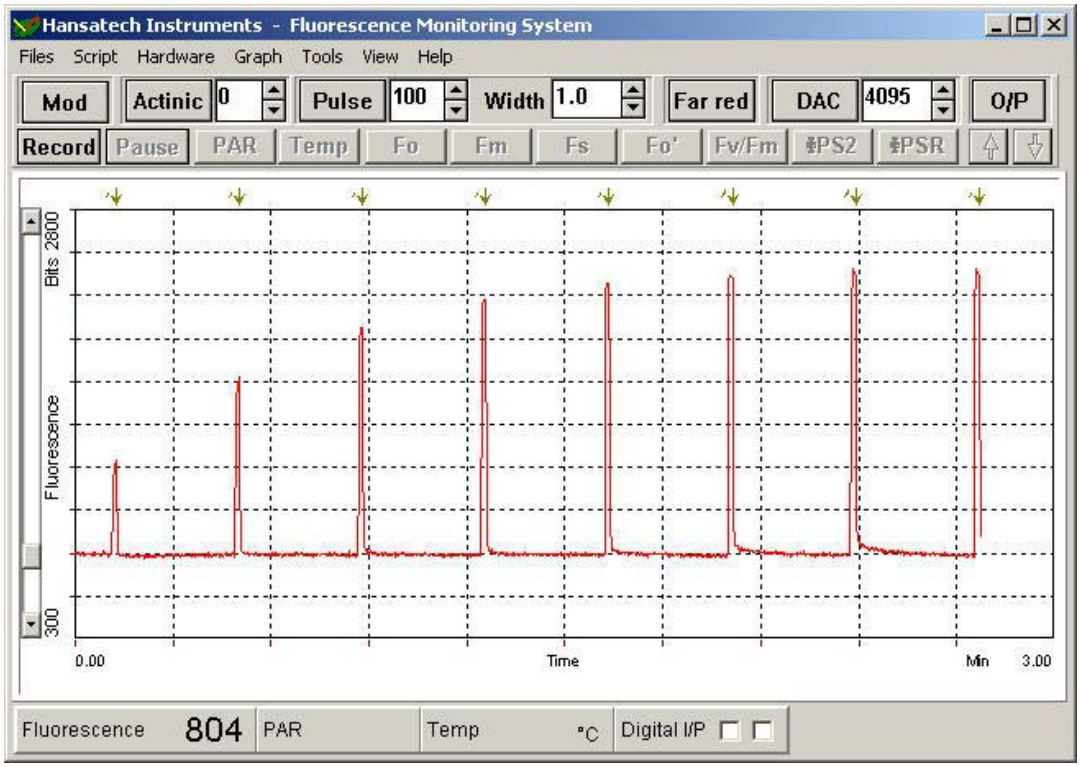

Fig. 2-3. Fluorescence trace from a healthy leaf sample. Initially the sample is adapted to the modulating beam and then subjected to pulses of increasing intensity to determine the correct setting to achieve light saturation of PSII.

The maximum fluorescence yield from the sample is generated by application of a saturating pulse of light (Fm Fluorescence Maximum). In this test several pulses are applied whilst the power of the saturating lamp is incremented between pulses. The resulting trace contains six fluorescence peaks (Figure 2-3) which correspond to pulses of 50, 60, 70, 80, 90 and 100 units respectively. The gradual increase in the magnitude of the pulsed fluorescence signal up to the 80 unit pulse indicates that a pulse intensity smaller than 80 units is not powerful enough to saturate PSII. Consequently the pulse intensity should be set to 80 units to ensure accurate measurement of Fm and PSII.

A second important function of the light pulses is to check that the maximum fluorescence signal is within the detection range of the instrument. The A/D works over the range 0 - 4095 bits. Any signal below zero or above 4095 will appear as a flat line at the bottom or top of the data recording area. The electronics of the FMS are designed to prevent a negative signal. However if the fibre-optic is oriented close to the sample it is possible that the detected signal will exceed 4095 bits and border the greyed-out area of the recording area. An example trace is shown in figure 2-4.

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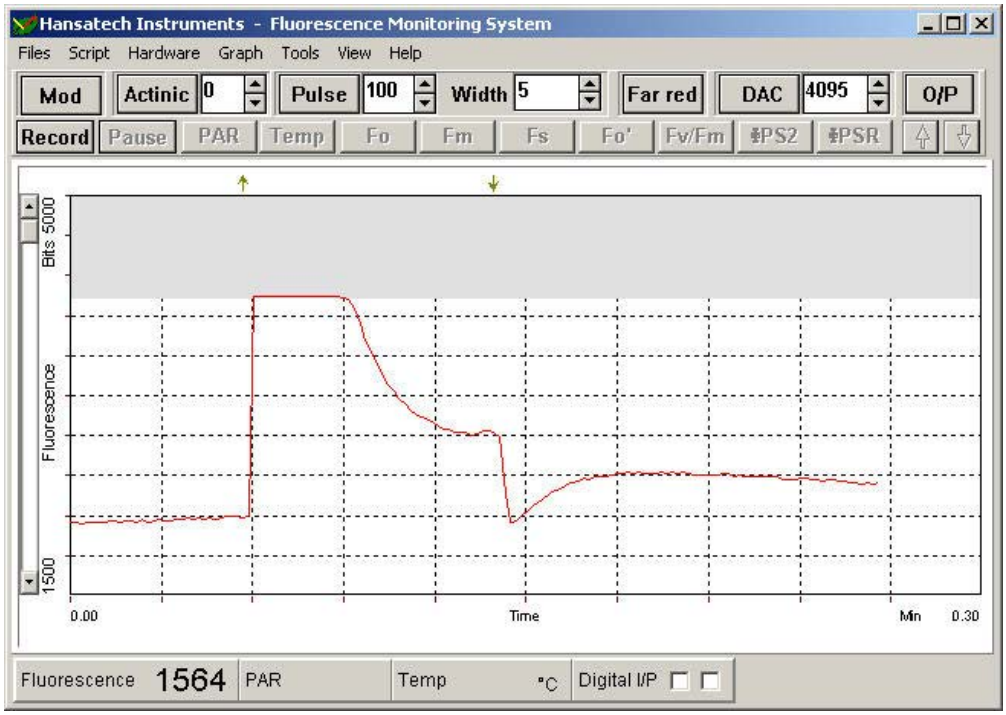

Fig. 2-4. During the saturation pulse (marked by the arrows) the fluorescence signal exceeds the range of the A/D converter. The peak of the pulse is cut from the trace but sampled (incorrectly) as 4095 bits by the software.

The peak fluorescence signal is effectively cut from the recording. This situation may be corrected by reducing instrument gain (Config Settings) or re-orienting the fibre-optic further away from the sample. If left un-corrected erroneous measurements of Fm and PSII will be stored when measurements are made.

As referenced above, all of the FMS configuration and data management facilities are accessed from the “drop-down” menu (C, Figure 2-1) whilst execution of measurements and on-line control of instrument light sources is initiated via the instrument toolbar (D, Figure 2-1). The following sections detail the configuration and data management facilities available from the “drop-down” menu. Description of each item on the instrument toolbar is given in The Instrument Toolbar.

3.3 Drop-Down” Menu Functions

The “drop-down” menu contains seven groups of related functions under the following headings:

Files Script Hardware Graph Tools View Help

In this section the groups are described in three broad catagories:

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• Configuration of hardware & software (Hardware, Graph, Tools and View)
• Data Handling & Export (Files)
• Instrument Programming via User-Defined (Script)
Scripts

3.3.1 Configuration of Hardware and Software

3.3.1.1 Hardware

When the Hardware menu is selected the following sub-menus are presented:

Hardware
Status
Clear memory
Set Time / Date
Config settings

3.3.1.2 Instrument Status

Before operating FMS in a remote situation it is useful to know what user-defined script files are loaded into the memory of the instrument, how much memory capacity is available for data storage and whether the battery holds sufficient charge to complete the experiment. The status command interrogates the instrument to generate a status window (Figure 2-5).

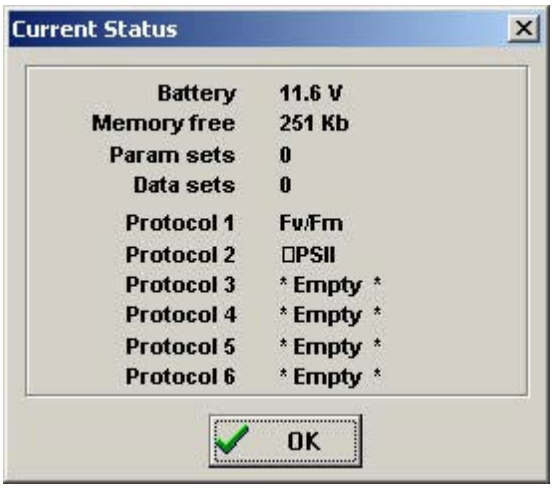

Fig. 2-5. Instrument Status Display.

Appropriate action can then be taken using other commands to prepare the instrument such as clearing memory for new data.

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3.3.1.3 Clear memory

Before initiating a new series of experiments it may be necessary to clear old files from the FMS to make space for new data. In this case the clear memory should be selected. The software will prompt to establish which files should be deleted (Figure 2-6).

Fig. 2-6. Delete files selection prompt.

The software will delete from the file number entered by the user to the end of the memory space. If file number 5 is entered into the above prompt box (Figure 2-7), files 5 through 10 (inclusive) will be deleted (Figure 2-7).

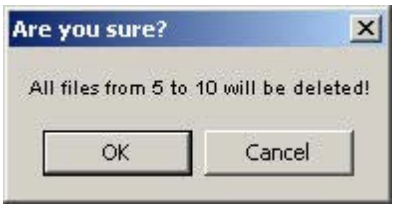

Fig. 2-7. File deletion prompt indicating that files 5 through 10 (inclusive) will be deleted.

3.3.1.4 Set Time / Date

The FMS has an on-board timing chip which can be used to log when experiments are executed in “Local” mode. The Set Time / Date hardware function allows the information within this chip to be up-dated. The software will initially prompt to set the FMS clock to match that of the PC. If this is not acceptable the user can manually enter the appropriate information into the time / date prompt box that is generated (Figure 2-8).

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Fig. 2-8. Insert time / date prompt box

3.3.1.5 Config settings

The Config settings sub-menu contains information on the current modulation beam setting and amplification (gain) settings used by the instrument (Figure 2-9).

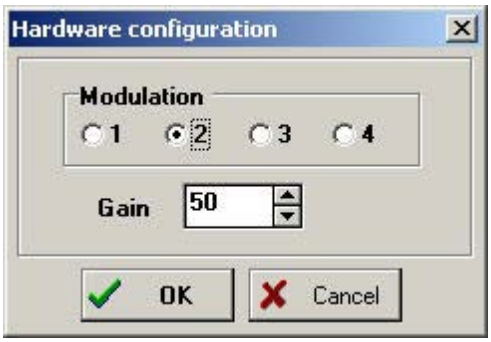

Fig. 2-9. Hardware configuration settings menu with modulation intensity setting of 3 (maximum) and 50 % gain applied to the signal

Default settings which should provide acceptable performance in most circumstances (Figure 2-9) are automatically loaded when the software is first executed. It is recommended that the tests described in section 2.3. are carried out before any experiments in order to verify that the settings are suitable. Adjustments to the settings are written to the MODFLUOR.INI file when the program is exited and re-applied when the program is next operated.

Once the instrument has been configured it is useful to set the software for a clear display of the data as it is acquired. These options are located under the View menu.

3.3.2 View Functions

When the program is first run the default screen displays both the instrument monitoring bar (B, Figure 2-1) and instrument toolbar (D, Figure 2-1); the rest of the screen is devoted to the data recording area. At the of end experiment when recording is finished the instrument monitoring bar is no longer required. The View menu allows the each of these items to be de-activated individually. The tick mark next an item indicates that it is displayed; de-activation is achieved by clicking on the item to remove the tick mark.

View
- Toolbar
- Monitoring bar
- Parameters window

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The parameters window is also activated from the View menu. This is a separate window used to log fluorescence parameters as their measurement is initiated from the instrument toolbar. The window may be dragged to any convenient position on the display and re-sized by stretching the box (Figure 2-10). Unwanted parameters may be removed by high-lighting with the mouse and pressing the “Delete” key.

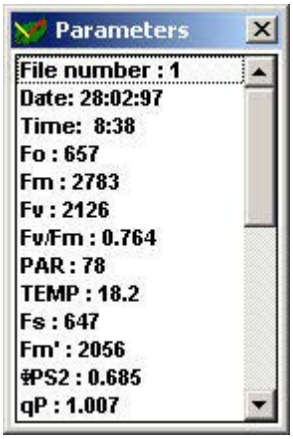

Fig. 2-10. “Parameters Window”

A scroll bar allows data browsing in the event that the list of data exceeds the dimensions of the window (Figure 2-10). All of the parameters in the window are available for export using the Params to clipboard function (Params to Clipboard Function) in the Files menu (File Management) to copy the data into the Windows® clipboard for pasting into another Windows® package. Each of the parameters measured by FMS is defined in Measurement Functions.

3.3.3 Graph Functions

Initial data recording screen settings are designed to display to the full fluorescence detection range of the instrument on time-base of 0 to 2 min. Data from experiments that exceed this time-base is automatically scrolled during recording so that some of the data is off-screen. The Graph menu contains several commands which may be operated either during or post recording to clarify the chart-recorder emulation and remove unwanted data from the display.

Graph
- Clear
- Autoscale
- Zoom
- Set axis

3.3.3.1 Clear

The Clear command can be used to remove all data from the recording screen and the parameters window (View Functions). It is important to note that such data is not retrievable unless it has

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been saved to a data file on floppy or hard drive (File Management. A prompt is generated to remind the user each time the command is used (Figure 2-11).

Fig. 2-11. Clear trace prompt

3.3.3.2 Autoscale

The Autoscale function re-sets both fluorescence and time axis to accommodate all of the currently recorded data on a single screen. It may be used at any time after a recording to "un-scroll" the chart-recorder emulation.

3.3.3.3 Zoom

After selecting the zoom function the mouse cursor should be moved to the area of interest in the data recording area. The left mouse button should be held down whilst the area to be zoomed is defined by dragging the cursor across the data recording area. A box is generated to outline the area of the zoom before it is enlarged to fill the data recording area.

3.3.3.4 Set Axis

Manual control over axis setting and other plotting attributes are accessed via the Set axis sub-menu (Figure 2-12).

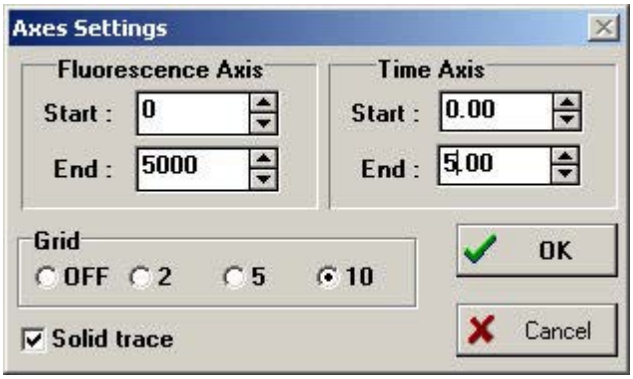

Fig. 2-12. Axis settings menu. The fluorescence axis set over the range 0 to 5000 bits with recording from time zero to 5 min. A solid trace is selected with 10 gridlines per axis.

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The on-screen ranges for both axes are set by entering “Start” and “End” points for each data range on the appropriate axis “edit” boxes, allowing a user-defined degree of trace zoom. Other display attributes include control over the background grid and whether data is plotted as individual data points or a solid trace chart-emulation.

THE SHORT DURATION OF PULSES MEANS THAT THERE ARE A RELATIVELY SMALL NUMBER OF PLOTTED DATA POINTS DURING EACH PULSE. WHEN INDIVIDUAL DATA POINTS ARE PLOTTED IT MAY BE DIFFICULT TO VISUALISE THE PULSE WITHOUT ZOOMING THE TRACE.

The individual data points which make up the recording may be accessed from the Tools menu.

3.4 Tools

The Tools menu contains items which relate to the raw data output from the A/D converter.

Tools
- Tabulate
- Event marker colour

3.4.1 Tabulate

Raw data can be presented during, or after recording in tabulated form by selecting the Tabulate function. The bit values of each data point are presented in rows of ten data points (A, Figure 2-13) against time (B, Figure 2-13) with a scroll bar to allow movement throughout the trace. Data is exported for pasting into other Windows packages with the Data to clipboard function (Data to Clipboard) which is located under the Files menu (File Management).

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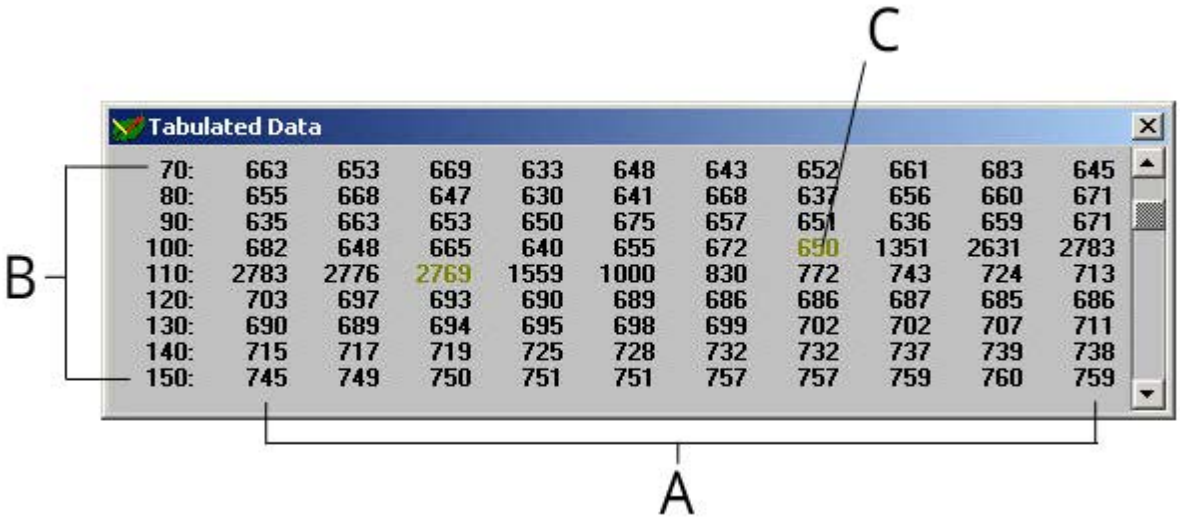

Fig. 2-13. Tabulated output of data collected from the A/D converter. Data is presented in rows of 10 data points (A) with the time of the first point at the start of each line (B). Event-marked data are colour-coded for easy reference (C).

Data points which correspond to trace event marks are tagged with colour-coded labels (C, Figure 2-13). The colours associated with each type of event marker can be defined by the user.

3.4.2 Event marker colour

The addition and removal of FMS light sources to the sample is event marked on the data recording screen with a different coloured arrow for actinic, pulse and far-red light sources. The default colours and data tags (in the tabulated output) for these are yellow, white and red respectively with blue reserved for user-applied event markers. The colour of each type of marker can be specified by selecting the appropriate marker type from the sub-menu presented and double-clicking on the required colour from the palette that is generated and selecting OK (Figure 2-14).

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Fig. 2-14. Paint Palette used for definition of event marker colour.

The final colour settings in place when the program is shut-down will automatically be saved to the MODFLUOR.INI file to become default values when the FMS is next is used.

3.5 Data Handling and Export

Once an experiment has been completed the data recording area should contain a trace with associated parameters stored in the “Parameters Window” (View Functions).

It is important to store this information to disc and to be able to retrieve previous work.

3.5.1 File Management With the Files Menu

The Files menu contains the following functions. Many of which are common to any computer package.

Files

- Save data
- Load data
- Upload to screen
- Upload to file
- Text search
- Params to clipboard
- Params to ASCII

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Data to clipboard
Data to ASCII
Print graph
Print params
Set printer page
Exit

In the case of FMS, “Modfluor” is used to transfer trace data which has been stored to the control unit’s on-board memory (Upload to screen, Upload to file). These functions are discussed in relation to “Local mode” use of the instrument in Section3

3.5.1.1 Save Data

Data can be stored on floppy or hard drive using the Save data function. The software will prompt to determine the final storage destination via the “Save data” prompt box (Figure 2-15).

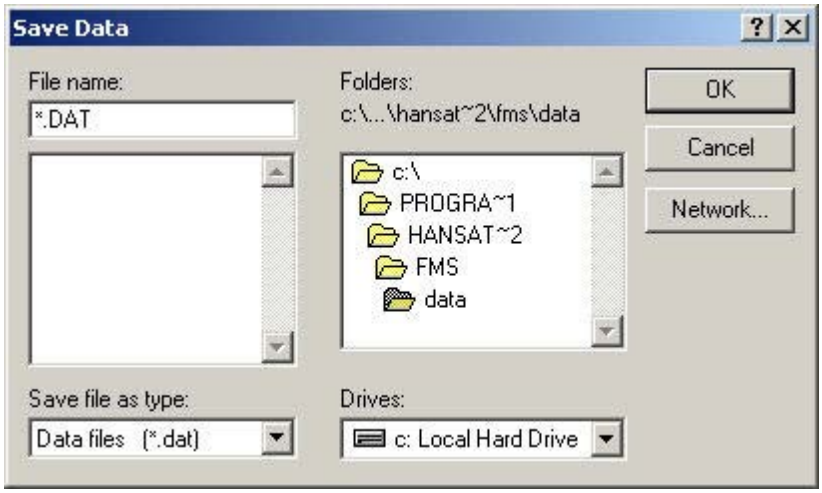

Fig. 2-15. Save data prompt box.

The mouse should be used to specify the drive and directory for data storage with the file name typed into the File name box. When correctly set the OK button closes the “Save data” prompt box and opens the “File description” prompt box (Figure 2-16).

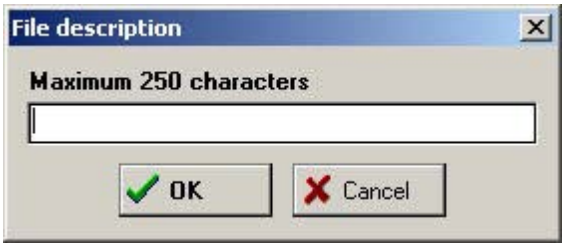

Fig. 2-16. File description prompt box.

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Up to 250 characters of text may be attached to the file as a reference which can be searched on during data retrieval (Text Search). When finished both trace and associated parameter data (if measured) are saved to file which is stored with a *.DAT extension

3.5.1.2 Load Data

Data files can be retrieved into the FMS software package using the Load data function. If a trace is already present on screen the program will prompt to overlay the existing data.

The first step in the process is to select the file to be opened by defining the file path in the “Load data” prompt box which is generated (Figure 2-17).

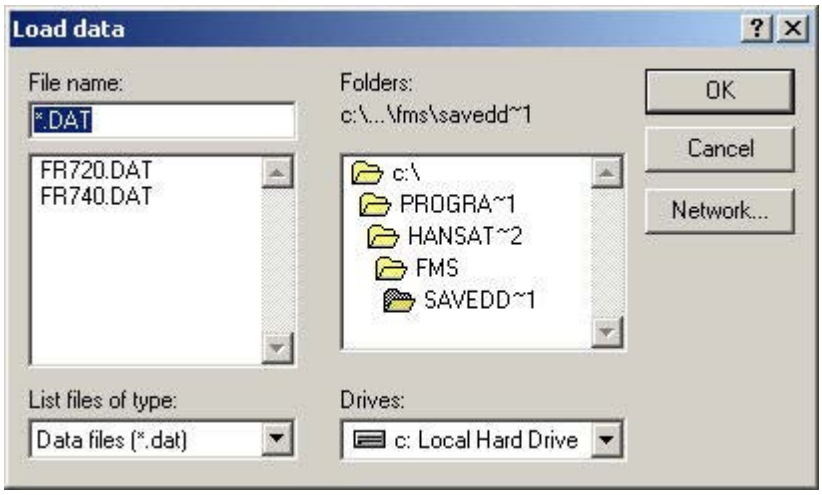

Fig. 2-17. Load data prompt box

In the above example the software is set to load a file named fr720.dat which is stored on drive C: in a directory called FMS. When the OK button is selected a further prompt box displays any text reference that was attached to the file when it was saved and requests confirmation of the action (Figure 2-18). Once confirmed the file will load with data recording area and “Parameters Window” updated accordingly.

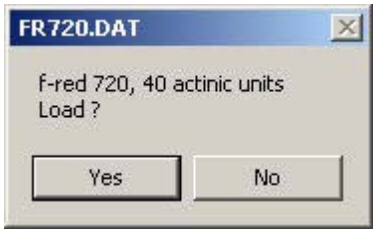

Fig. 2-18. Confirmation box for file loading.

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3.5.1.3 Text search

As described in section 2.4.5.1.1. a descriptive text reference can be attached to each data file as it is saved. The Text search function is provided for searching these references for a particular text string in the event that a particular file name is forgotten. When selected the “File notes” prompt box is displayed (Figure 2-19).

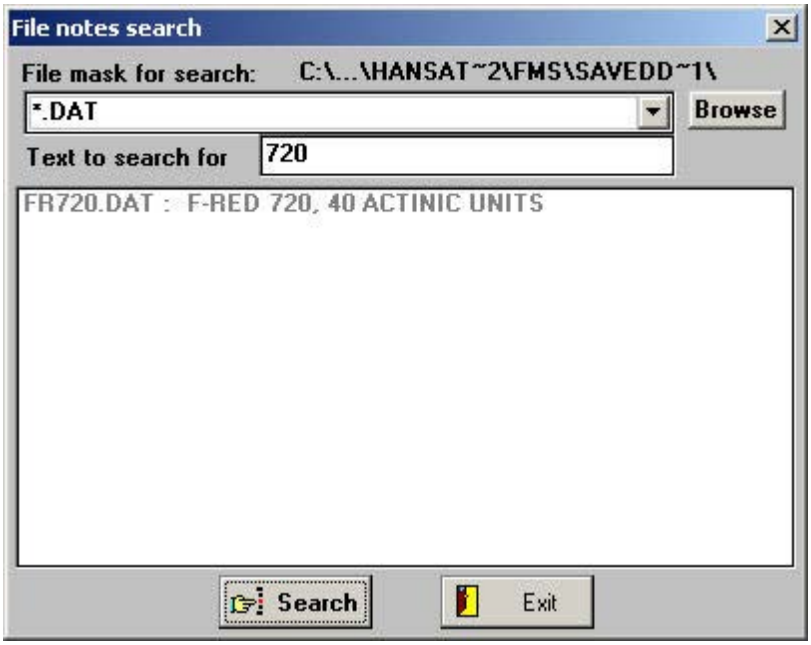

Fig. 2-19. File notes search prompt box.

Initially the Browse button should be used to view the PC drive and directory structure and the search directory selected by clicking on it and selecting OK. The text string of interest should then be entered or the default wildcard (*) used. The search initiated by pressing the “Search” button. In the above example all files contained in the C:\FMS directory were searched from the string “720”. Only one file had “720” in its associated reference; the name of the file (FR720.DAT) and the full reference (F-RED 720, 40 ACTINIC UNITS) are presented in a search results window (Figure 2-19).

3.5.2 Exporting Data to Other Windows® Applications

The data recording screen contains two types of information at the end of a “PC mode” experiment: trace data is graphically displayed as a chart-recorder emulation and parameter data is stored in the “Parameters window”. Both types of information can be cut to the Windows® clipboard for pasting into other applications via the Params to clipboard and Data to clipboard functions or exported as ASCII files via the Params to ASCII and Data to ASCII finctions.

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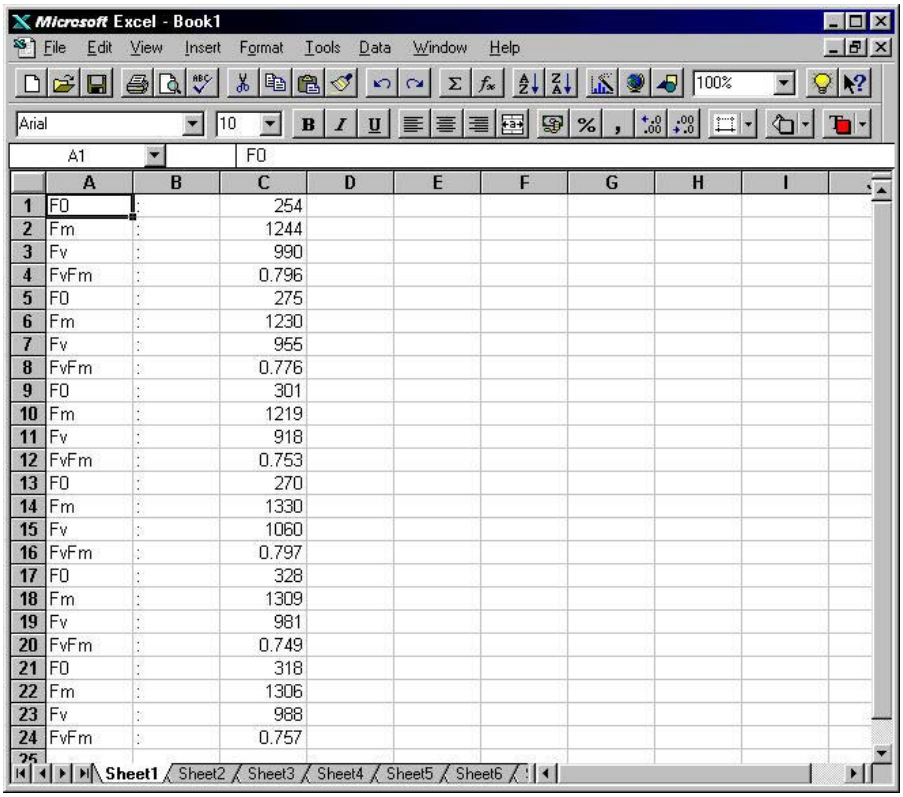
3.5.2.1 Params to clipboard and Params to ASCII

When selected the Params to clipboard function copies the contents of the "Parameters window" as text to the clipboard. The updated contents of the clipboard are suitable for pasting into data analysis or spreadsheet packages such as "Lotus" or "Excel". The example below refers to Microsoft Excel but the procedure is virtually the same for all Windows® applications.

Once recording is completed select the Params to clipboard function; all of the parameters are automatically copied to the clipboard. The ALT + TAB keys should then be used to switch to the destination application or in the event that the destination application is not running, it should be initiated from the Windows® taskbar. The contents of the clipboard are then pasted into the destination application.

In the case of "Excel" the "paste" function is located in the Edit menu. The contents of the clipboard should appear on the spreadsheet (Figure 2-20).



	A	B	C
1	F0	:	254
2	Fm	:	1244
3	Fv	:	990
4	FvFm	:	0.796
5	F0	:	275
6	Fm	:	1230
7	Fv	:	955
8	FvFm	:	0.776
9	F0	:	301
10	Fm	:	1219
11	Fv	:	918
12	FvFm	:	0.753
13	F0	:	270
14	Fm	:	1330
15	Fv	:	1060
16	FvFm	:	0.797
17	F0	:	328
18	Fm	:	1309
19	Fv	:	981
20	FvFm	:	0.749
21	F0	:	318
22	Fm	:	1306
23	Fv	:	988
24	FvFm	:	0.757

Fig. 2-20. FMS parameters data pasted directly into Excel.

The parameters are listed as they were measured in descending order with the final parameter at the bottom of the list. This basic format means that data from several replicate experiments can be pasted next to earlier data in the same spreadsheet. With replicate data pasted in a row across the spreadsheet calculation of mean and standard errors becomes a simple matter of typing the appropriate formulae and high-lighting the range of replicate data to be included. Once analysis of

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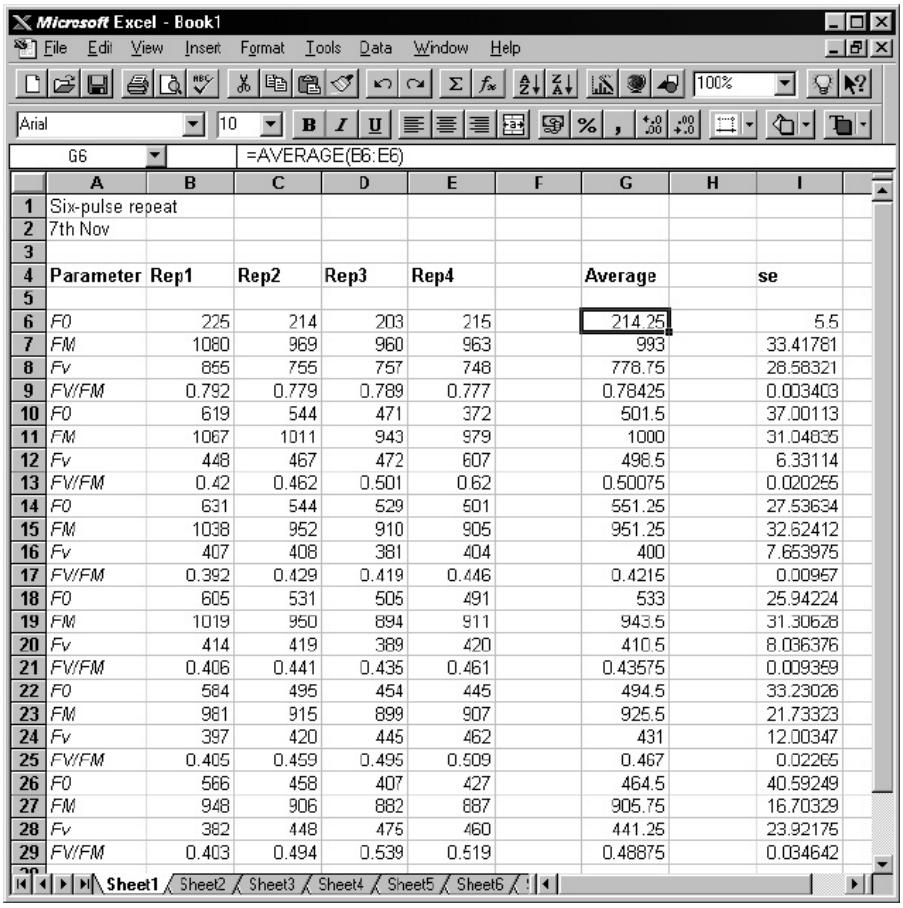
the top row has been completed calculations for all susequent rows can be made automatically using the Fill Down command (Figure 2-21).



Microsoft Excel - Book1

File Edit View Insert Format Tools Data Window Help

Arial 10 100%

G6 =AVERAGE(B6:E6)

	A	B	C	D	E	F	G	H	I
1	Six-pulse repeat								
2	7th Nov								
3									
4	**Parameter**	**Rep1**	**Rep2**	**Rep3**	**Rep4**		**Average**		**se**
5									
6	*F0*	225	214	203	215		214.25		5.5
7	*FM*	1080	969	960	963		993		33.41781
8	*Fv*	855	755	757	748		778.75		28.58321
9	*FV/FM*	0.792	0.779	0.789	0.777		0.78425		0.003403
10	*F0*	619	544	471	372		501.5		37.00113
11	*FM*	1067	1011	943	979		1000		31.04835
12	*Fv*	448	467	472	607		498.5		6.33114
13	*FV/FM*	0.42	0.462	0.501	0.62		0.50075		0.020255
14	*F0*	631	544	529	501		551.25		27.53634
15	*FM*	1038	952	910	905		951.25		32.62412
16	*Fv*	407	408	381	404		400		7.653975
17	*FV/FM*	0.392	0.429	0.419	0.446		0.4215		0.00957
18	*F0*	605	531	505	491		533		25.94224
19	*FM*	1019	950	894	911		943.5		31.30628
20	*Fv*	414	419	389	420		410.5		8.036376
21	*FV/FM*	0.406	0.441	0.435	0.461		0.43575		0.009369
22	*F0*	584	495	454	445		494.5		33.23026
23	*FM*	981	915	899	907		925.5		21.73323
24	*Fv*	397	420	445	462		431		12.00347
25	*FV/FM*	0.405	0.459	0.495	0.509		0.467		0.02265
26	*F0*	566	458	407	427		464.5		40.59249
27	*FM*	948	906	882	887		905.75		16.70329
28	*Fv*	382	448	475	460		441.25		23.92175
29	*FV/FM*	0.403	0.494	0.539	0.519		0.48875		0.034642

Sheet1 Sheet2 Sheet3 Sheet4 Sheet5 Sheet6

Fig. 2-21. Sorted and formatted experimental data.

The Params to ASCII function writes the contents of the Parameters Window to a user-named ASCII file in two column, comma delimited format:

Parameter label	Value
Fo	400 etc

These files are suitable for import as columns into most DOS and Windows data packages.

3.5.2.2 Data to Clipboard and Data to ASCII

The raw fluorescence data which forms the recorder-emulation trace is also available for export via the Data to clipboard command. This operates in a similar manner to parameter export (Params to Clipboard) with numeric values transferred as a text string.

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The procedure for pasting to the clipboard contents remains unchanged. However, the increased quantity of data involved means that several passes through the clipboard may be required.

When pasting to the clipboard the software will prompt for the first and last data point in the range of data to be transferred. If the specified range is small enough all of the data is pasted to the clipboard in one pass. If it is too large the software will paste as much data as possible and report the number of the last point pasted. The clipboard contents should then be pasted into the destination application and a second clipboard pass made using the reported data number as the first point in the second pass. A maximum of four passes may be required to paste a maximum length trace.

The data is pasted in the destination application as a single of column of sequential readings (Figure 2-22).

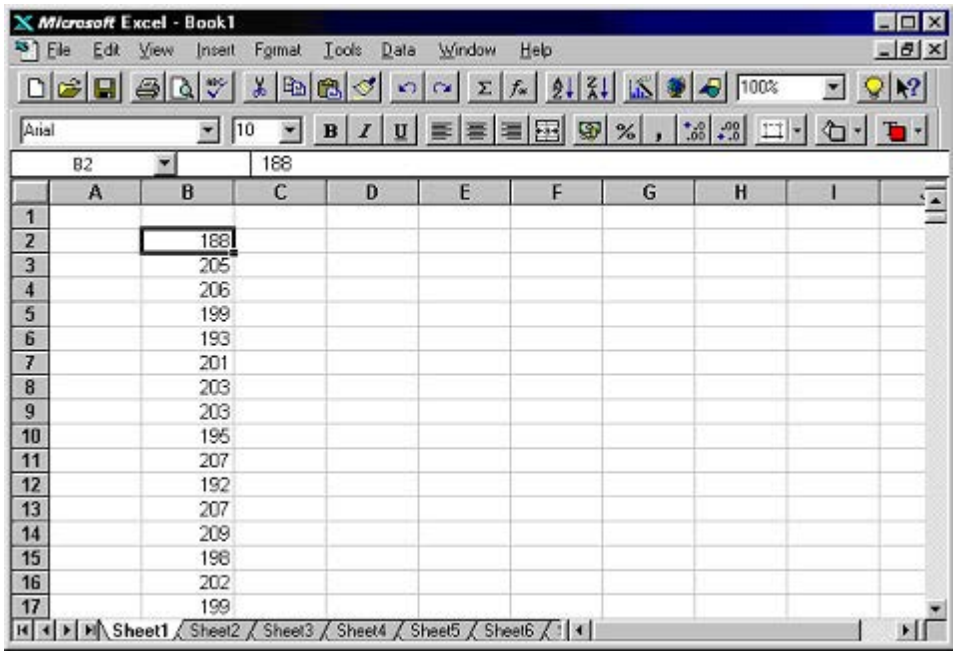

Fig. 2-22. Tabulated trace pasted directly into into Excel.

Data points are stored every 0.11 sec. As a result a column of time information may be added by entering 0.11 in the cell next to the first data point (A2, Figure 2-23) and the formula (=A2+0.11) to the cell below. This formula can be “filled down” to add a time-base to all of the data (Figure 2-23).

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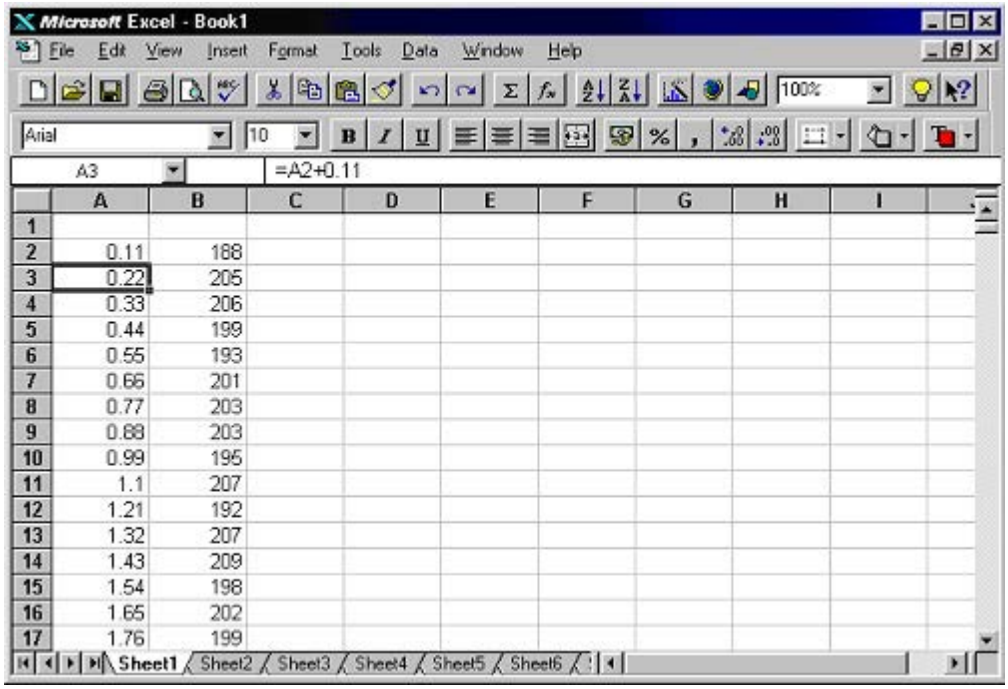
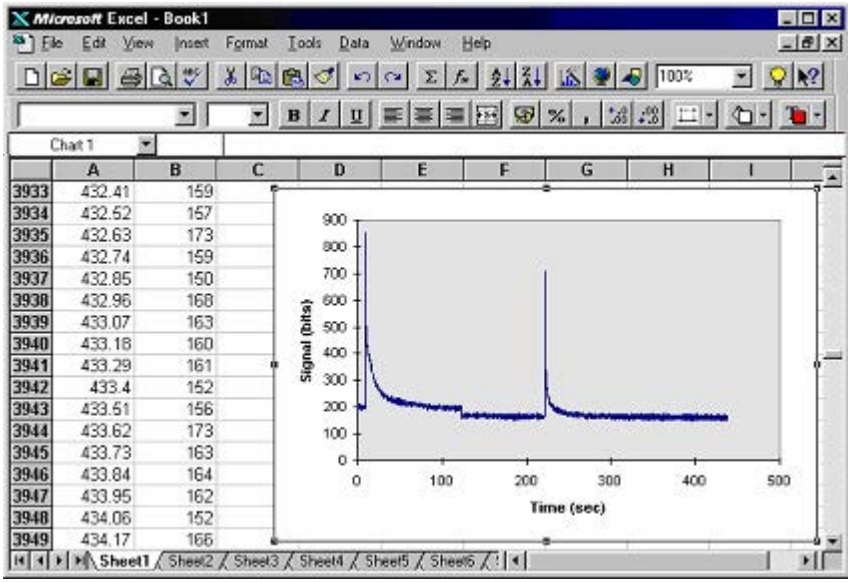


	A	B
2	0.11	188
3	0.22	205
4	0.33	206
5	0.44	199
6	0.55	193
7	0.66	201
8	0.77	203
9	0.88	203
10	0.99	195
11	1.1	207
12	1.21	192
13	1.32	207
14	1.43	209
15	1.54	198
16	1.65	202
17	1.76	199

Fig. 2-23. Time-base added to exported data.

Once in this format it is simple to plot the data as an X,Y plot for presentation (Figure 2-24).



	A	B
3933	432.41	159
3934	432.52	157
3935	432.63	173
3936	432.74	159
3937	432.85	150
3938	432.96	168
3939	433.07	163
3940	433.18	160
3941	433.29	161
3942	433.4	152
3943	433.51	156
3944	433.62	173
3945	433.73	163
3946	433.84	164
3947	433.95	162
3948	434.06	152
3949	434.17	166

Fig. 2-24. Data plot of signal against time using Excel “Chart-Wizard”.

The Data to ASCII function writes all of the trace data to a user-named ASCII file in two column, comma-delimited format:

Time of reading (sec Signal value (bits)

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0.11	100
0.22	101 etc

The files are suitable for importation into most DOS and Windows manipulation packages

3.5.3 Printing

The contents of the data recording screen (A, Figure 2-1) and the "Parameters Window" may be printed to most Windows® printer drivers using the Print graph and Print params commands respectively. When selected the print dialogue box shows the current default printer driver and permits the selection of colour or monochrome printing depending upon the type of printer selected (Figure 2-25).

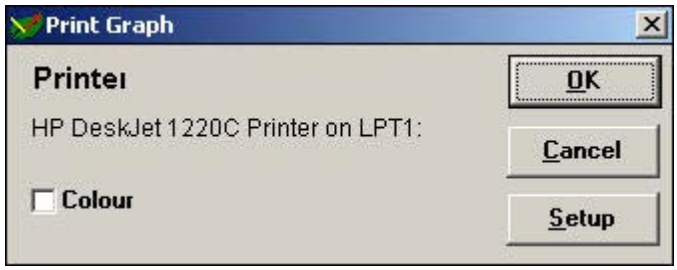

Fig. 2-25. Print dialogue box.

The selected printer settings can be changed through the "Print setup" dialogue box (Figure 2-26) which is accessed from the Setup button.

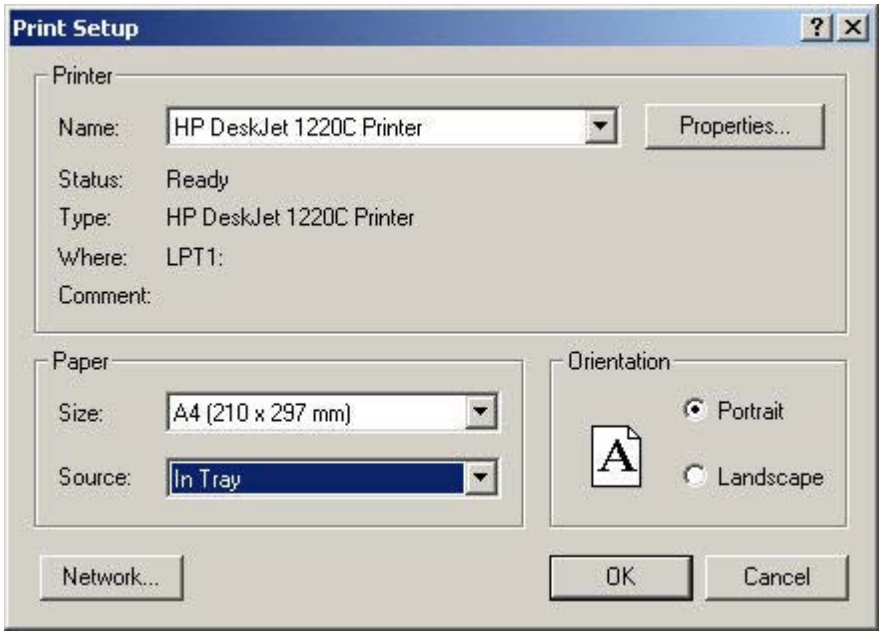

Fig. 2-26. "Print setup" dialogue box.

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The Print graph function prints all of the graphics information which is displayed within the boundary of the data recording area plus event markers whilst the Print params functions outputs each of the parameters in a single column in descending order as they were recorded.

The size of the print output can also be adjusted to fit different areas on the page with the Set printer page feature (Figure 2-27).

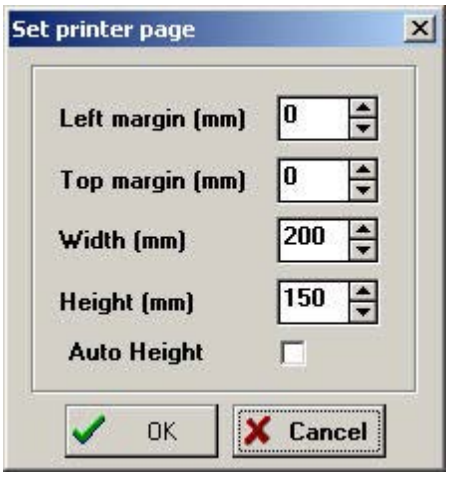

Fig. 2-27. Set printer page dialogue box.

The default settings give minimal margins. The print area may be changed by altering the height and width settings with a constant height / width aspect ratio maintained by ticking the “Auto Height” option box. The printer page settings apply to output from both Print graph and Print params functions.

3.6 Instrument Programming Via User-Defined Scripts

The FMS has been developed in parallel with Hansatech Scripting Language (HSL). The latter consists of a simple, yet extremely versatile, method for pre-programming a maximum of six user-defined control event / measurement sequences (referred to as protocols or scripts) into the instrument. Once programmed the protocols are available for operation by a single-key press under “Local mode” outside of the laboratory or from the Windows software. The Script function on the “drop-down” menu allows access to the Script Editor for creation / editing of protocols, the Script Recorder which logs commands used in “PC mode” to automatically generate a script and the script Run command for Windows operation.

3.6.1 The Script Recorder

The principles behind the Script Recorder are very similar to those of “Macro Record” functions found in spreadsheet packages. When active the recorder logs the sequence and timing of every command that the user issues during the experiment and automatically creates a script which during play-back precisely replicates the original series and timing of commands that were issued.

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The Script Recorder is activated / de-activated by clicking the Record option under the Script menu. A tick signifies that the recorder is set to operate as soon as a new recording is initiated from the “Record button” on the instrument toolbar (Recording Functions).

Script
- Run
- Edit
- Record

Script recording is automatically stopped when the experiment is terminated from the instrument toolbar (Recording Functions). The software then prompts before loading the newly recorded script into memory as any currently loaded script will be overwritten. Once loaded the script may be viewed, Run, saved to file or edited using the
script editor.

TIME INTERVALS BETWEEN COMMAND STATEMENTS ARE ALWAYS INTERPRETED BY THE SCRIPT RECORDER AS WAIT ELEMENTS OF APPROPRIATE DURATION. THERE ARE SEVERAL DIFFERENT TYPES OF “SCRIPT PAUSE” ELEMENT WHICH MAY BE MORE SUITABLE FOR “LOCALMODE” APPLICATIONS. THESE MUST BE MANUALLY EDITED INTO RECORDED SCRIPTS USING THE SCRIPT EDITOR (THE SCRIPT EDITOR).

3.6.2 Running a Script Under Windows

The Run command executes the current script that is loaded in the script editor with data presented in the data recording area and parameters window (if open, View Functions) as it is acquired.

Most of the individual script elements (Hansatech Scripting Language Elements) are designed to operate in the same manner regardless of whether they are executed from the control box in “Local mode” (Section 3) or whether they are operating under the Windows software. There are however some exceptions in the way Script Pause and plot identification elements (Hansatech Scripting Language Elements) are interpreted. These differences are necessary to account for the different operating environments and may be need to be considered when creating scripts with the script editor.

Element	Windows Operation	Local Operation
WAIT	Pauses script for defined duration.	Delays execution of subsequent script elements for a fixed duration.
WKEY	Pauses script indefinitely until the SPACE-BAR is pressed.	Delays execution of subsequent script elements until a key is pressed.

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WMON	Pauses script indefinitely until the SPACE-BAR is pressed	Delays execution of subsequent script elements until a key is pressed whilst displaying updated fluorescence level.
WLEV	Pauses script indefinitely until signal changes by less than a defined amount.	Delays execution of subsequent script elements until rate of signal change is smaller than defined amount.
ID	Not operated	Requests plot identification information each time the script is executed in “Local-mode” (Creating an Example Script).

It is recommended that the TEXT element is used to insert information before use of certain Pause Elements to indicate that user-intervention is required for the script to proceed (e.g. WKEY, WMON, WDON, WOFF). The following example displays a text prompt (“Press SPACE-BAR“) above the instrument monitoring bar whilst the script is paused and clears the text after user input has been received:

TEXT : Press SPACE-BAR
WKEY :
CLR :

3.6.3 The Script Editor

The script editor is a graphics-based window where the contents of a script are listed and made available for editing with the iconised tool functions (Figure 2-28).

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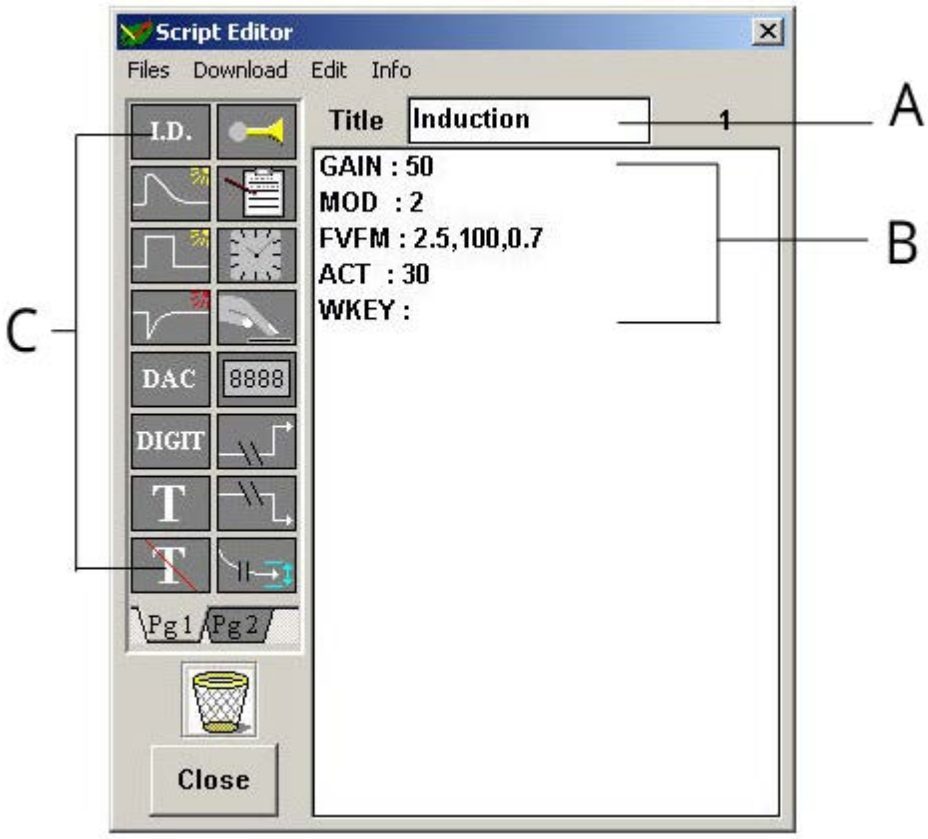

Fig. 2-28. The script editor is used to display and edit each script. The script title is presented in the title box (A) with the command lines or script elements displayed in a box below (B). Further script elements can be added by "dragging" them from the iconised selection (C).

Each script consists of two parts: a title, presented in the title box (A, Figure 2-28), which is used to identify the script when loaded into the FMS and a sequence of script elements which are to be executed in descending order during the experiment (B, Figure 2-28).

Editing, down-loading and script file handling functions are contained within the "drop-down" menu whilst further commands may be added using a "drag and drop" procedure to select items from the two pages of iconised script elements (C, Figure 2-28).

The best way to appreciate the principles behind the script editor is to create an example script

3.6.3.1 Creating an Example Script

The first step is to create a new file from the Files group on the "drop-down" menu. The editor will prompt for a script title (maximum of 10 characters) which is also used to identify the script on the FMS display screen after it has been downloaded.

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As a result the title should be as descriptive as possible. In this example the script is called “Test” (Figure 2-29).

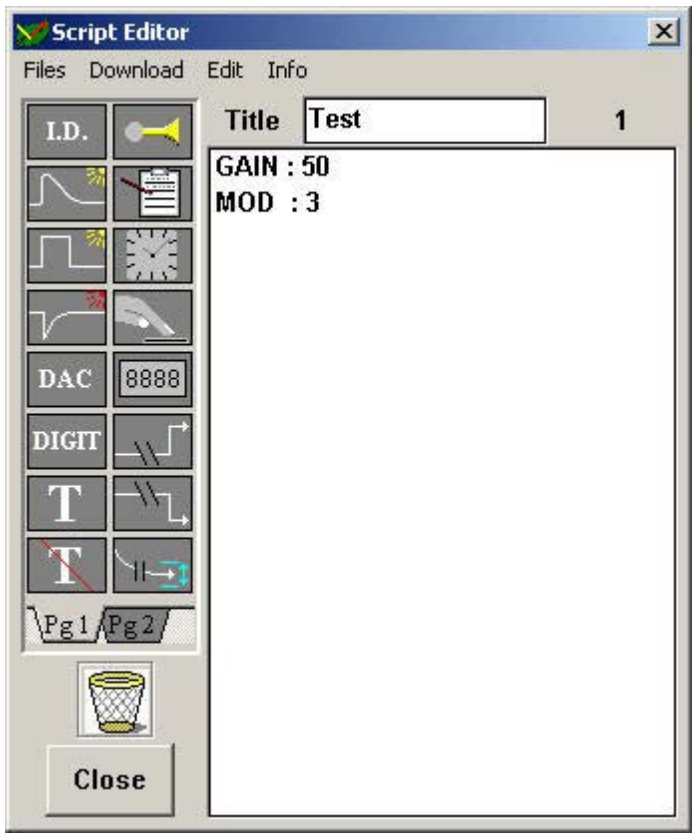

Fig. 2-29. “Test” example script with first elements automatically added.

A pair of essential script elements are always added at the beginning of each script automatically. These are GAIN and MOD which set instrument amplification and modulation beam intensity respectively. The editor uses the following format when listing all script elements:

Name of script element	Element parameters
GAIN	50
MOD	3

The name of the script element is listed, followed by a colon (:) and numerical parameter(s) which define the element’s function. In the above case gain is applied at instrument setting 50 and the modulating beam is switched on at setting 3.

Further elements may then be added to the script in the desired order of execution. Available elements are presented as two pages of icons next to the developing script. Page one contains control events such as actinic light control and event timers such as “wait for X secs” or “wait for instrument key press” before continuing script whilst page two contains direct measurement functions including temperature, PAR and common fluorescence parameters. The purpose of the

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Script Editor is to allow the user to string individual elements together in any order, generating complex experiments which can be operated from a single key-press on the FMS when away from a PC or executed automatically under the Windows software.

A full description of each script element, the control events initiated with associated parameters and the data that is measured during its operation are listed in Appendix A. It is important to remember that each element is an independent unit and thus can be inserted in any position within a script after the initial GAIN and MOD functions.

In this example the script for a simple light-response measurement will be created and the FMS will be commanded to LOG all trace data to give a full chart recorder emulation of the experiment upon return to the laboratory. The FMS will also be set to request experimental plot identification information with the ID script element.

Once the new script file has been created (Figure 2-29) the next step is to select the ID element. This function is added to the script by clicking on the “Page 1” tab to open the correct icon page and moving the mouse cursor over the ID icon. After a second, a yellow box beneath the icon automatically opens to provide further information on the icon. The element is incorporated into the script by holding the left mouse button down and “dragging” the ID icon beneath the MOD element in the script and releasing the left mouse button. A prompt box is generated requesting a pair of values M,N. These can be considered as number of treatments (M) and number of replicates (N). Thus 5,5 would denote a 25 plot experiment with 5 treatments replicated 5 times. When entered the following line is added to the script:

ID : 5,5

When this script is run in “Local mode” (Section 3) the FMS will prompt each time the script is run for a plot number. The first run the default number will be 1,1 indicating that treatment 1, replicate 1 is to be measured. This can be accepted or an alternative plot ID entered with the keys. The entered plot information is saved with the file at the end of the measurement and displayed in the parameters window when the data is uploaded. When the script is executed a second time the default plot number is automatically incremented (1,2 , 1,3 , 1,4 , 1,5 , 2,1 etc.).

The next element to add to the script is the command to LOG all of the trace data for detailed analysis on upload. The LOG element (clipboard and pen icon) should be dragged beneath the ID element and the parameter “1” entered into the parameter prompt box.

A LOG element with parameter “0” may be used to stop anywhere during the script. In our script logging is required until the end of the script. Hence the following line is added:

LOG : 1

Once the set-up elements have been entered the measurement elements should be added. The first such measurement is dark-adapted Fv/Fm. Upon addition the editor will prompt for parameter

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values which define the Fv/Fm measurement, giving default numeric values which should be suitable for most circumstances (Figure 2-30).

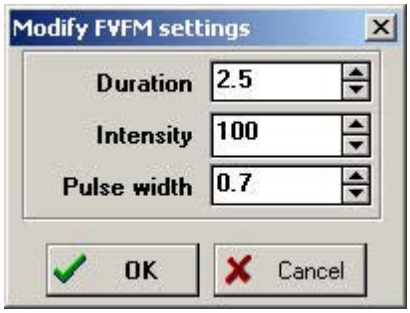

Fig. 2-30. Modify Fv/Fm settings.

The measurement of Fv/Fm requires two control events: averaging of data to determine Fo and application of a saturating light pulse to determine Fm.

Consequently the parameters are:

Parameter	Definition	Default Value
Duration	Period over which element runs	2.5 secs
Intensity	Intensity of saturation light pulse 100 sec (1-100 units)	100 secs
Pulse Width	Duration of light pulse 0.7 sec (0.7 - 5.0 sec)	0.7 secs

If the default values are accepted the following command line is added to the script:

FVFM: 2.5, 100, 0.7

The need for defined parameters adds complication to script development but their presence makes it possible to customise scripts to suit individual requirements and thus optimise the performance of the instrument. A complete listing of the parameters associated with each script element is given in Hansatech Scripting Language.

The next stage is to switch on the actinic light at a predetermined level. This is done by selecting the “Actinic Light” icon from page 1 of the script elements and “dragging it below the Fv/Fm measurement (Figure 2-31).

Fig. 2-31. Actinic Light Icon

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Once released the program will prompt for the intensity of illumination with a default value of 30 units. In this case light should be set to 10 units by entering 10 and pressing RETURN. The following command line should be added:

ACT: 10

The fluorescence signal normally passes through a series of changes immediately after illumination whilst the photosynthetic processes become light adapted. Measurement is often delayed until the sample has reached “steady state”. The scripting language contains several “Wait” commands which are suitable in different contexts e.g. waiting for a key-press from the user, waiting for a defined period of time or waiting for a digital input from some external apparatus (Hansatech Scripting Language). In this example the “wait for a fixed period of time” command will be used by “dragging” the clock icon from page 1 to the script (Figure 2-32); the program will prompt for the wait duration in units of seconds.

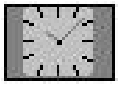

Fig. 2-32. Wait for fixed period Icon.

A value of 100 sec should be entered to generate the following command line.

WAIT: 100.0

After 100 sec the next command will be executed. This will be a measurement of the quantum efficiency of PSII (fPSII) which is available from page 2 of the icons. The control functions required for fPSII measurement are similar to those of Fv/Fm. Hence the same types of parameters are used. Although it should be noted different values for any parameter may be used. If default parameters are used the following command line should be inserted:

ϕPSII : 2.5, 100, 0.7

The final script should appear similar to that in figure 2-33.

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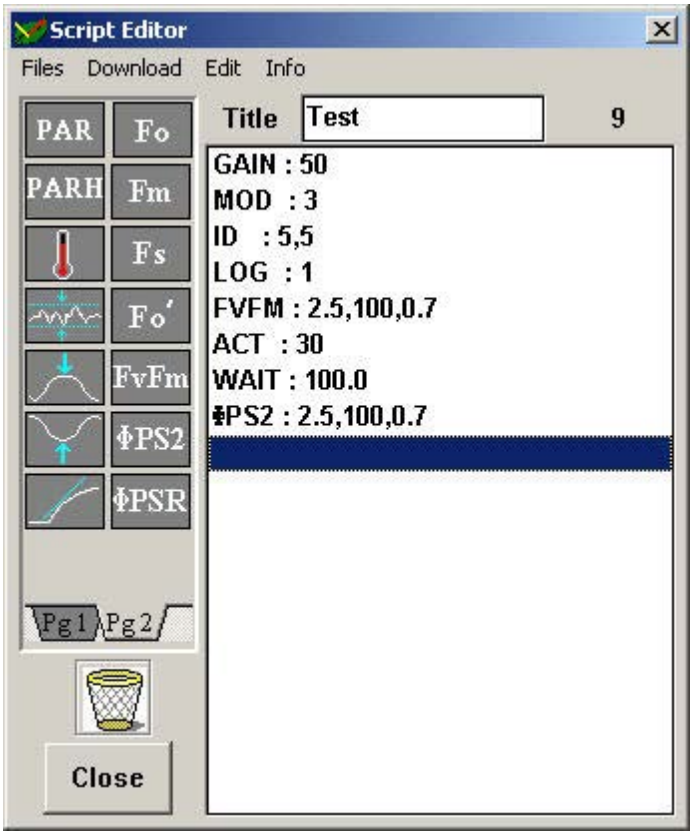

Fig. 2-33. Final "Test" example script.

The above example is rather trivial containing only eight commands. The full potential of HSL caters for around 30 script elements which may be assorted into a maximum script of 500 bytes (equates to approximately 80 Fv/Fm elements). When dealing with larger scripts some of the editing and file handling functions are extremely useful. These are accessed from the "drop-down" menu.

3.6.3.2 Script Editor "Drop-Down" Menu

The "drop-down" menu in the Script Editor operates in a similar manner to the main "drop-down" menu in the recording screen; the individual commands are divided into groups and accessed under headings which broadly describe their function. There are three headings: Files, Download and Edit.

Edit

The Edit menu contains commands suitable for adding, deleting or changing the parameters of commands within a script.

Edit

Add

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Insert
Modify
Delete

Some of these functions can be duplicated using the mouse to “drag” in new commands from the icons or delete existing commands by “dragging” then to the waste paper basket.

Files

Once a script has been created it is sensible to store it to disc. Scripts may be stored to or retrieved from *.SCR files using options under the Files menu.

Files
New Script
Save Script
Load Script
Text Search
Close

The procedures for operating Files are the same as those described in File Management

Download

A maximum of six script files can be down-loaded to the FMS and executed in the absence of a computer via “Local mode” operation (Section 3). The Download menu is used to transfer the script that is currently displayed in the Script Editor into one of the six protocol locations within the control unit. The appropriate location is selected from the Download menu with the mouse or accelerator key; a prompt indicates successful script transfer to the control box.

Download
Protocol 1
Protocol 2
Protocol 3
Protocol 4
Protocol 5
Protocol 6

When the instrument is first used Protocols 1 and 2 will contain default screening scripts which measure Fv/Fm and fPSII. These, and any other script can only be over-written if they are unused and thus do not have experimental data stored with them. This safe-guard is essential as the FMS uses the loaded script to interpret the data that is stored on-board. Hence it is only possible to re-use script locations if the data files from the instrument are up-loaded (Section 3) and the memory cleared (Clear Memory).

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3.7 The Instrument Toolbar

In “PC mode” FMS light sources and measurement features are selected from buttons presented on the instrument toolbar (Figure 2-34).

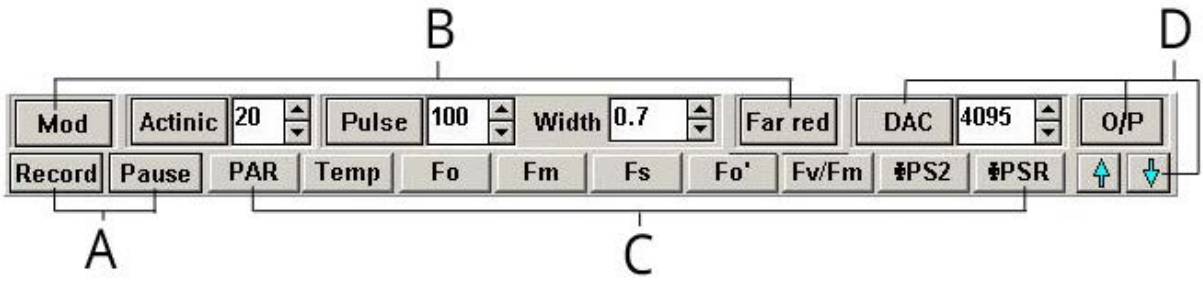

Fig. 2-34. Instrument toolbar.

The features can be categorised into four functional groups:

- Recording Functions (A, Figure 2-34),
- Light Source Controls (B, Figure 2-34),
- Measurement Commands (C, Figure 2-34),
- External Device Interface and Event Markers (D, Figure 2-34).

3.7.1 Recording Functions

Recordings are initiated with the “Record” button. This also triggers the modulation beam, essential for fluorescence detection (Section 1), and if the script record function is ticked (The Script Recorder) will start recording the sequence of measurement commands that are issued from the toolbar during the experiment. A good analogy is the recording of key strokes by spreadsheet software to create re-usable “macro” programs.

Throughout each experiment data is plotted to the data recording screen and presented on the instrument monitoring bar (if active) every 0.11 sec for a maximum of approximately 50 minutes. Experiments may be run over longer periods if the “Pause” button is used to conserve memory space by suspending data logging during periods of the experiment where measurements are not required (e.g. duringadaptation to a new light regime). Fluorescence readings are continuously up-dated on the instrument monitoring bar even if data logging is “Paused”.

Recording is terminated by re-pressing the record button. All active light-sources are switched off and script recording (if active) is stopped. A dialogue box will prompt to determine whether the newly recorded script should be allowed to over-write the script which is currently open in memory. Further details regarding script creation and editing are located in Creating an Example Script.

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3.7.2 Light Source Controls

The FMS control unit has four integral light sources which are controlled from the instrument toolbar.

3.7.2.1 The Modulation Beam

For most experiments the modulation beam must be switched on at the beginning of the recording and switched off at the end. This is done automatically during initiation and termination of recording with the “Record” button. However, it is possible to turn the modulating beam off during an experiment by clicking the “Mod” button (Figure 2-34). When this is done the fluorescence signal will drop to zero bits and remain at this level until the beam is restored. This function may be useful in a experimental situation where it is important to stop all photochemistry for a period; the only certain method for achieving this is to prevent any light from reaching the sample. It may be useful to pause data logging in this situation as the instrument is unable to detect a fluorescence signal.

The modulating beam can be operated at one of three set intensities with control provided through the Hardware function of the “drop-down” menu. The fluorescence signal that is detected by FMS is directly proportional to the intensity of the modulating beam incident upon the sample. Only by keeping this constant during an experiment is it possible to monitor changes that relate to sample photochemistry. As a result the optimum modulation intensity should be configured prior to starting the experiment on a piece of test tissue (Testing the FMS), as the FMS software will not allow changes whilst recording is in progress.

3.7.2.2 The Actinic Light Source

Actinic light is provided by operating the LED Light Source at reduced power. The actinic output range is 0 to 3500 $\mu molm^{-2}s^{-1}$ (measured at the end of the fibre optic) selectable in 50 steps via the numeric value entered in the edit box next to the “Actinic” button. The “Actinic” button initiates illumination and remains indented until re-clicked to switch the lamp off. It is important to note that the lamp must be switched off and re-started with a new numeric value in the edit box to change illumination intensity.

Whilst the actinic lamp is active the contribution from the modulating beam to the total amount of absorbed irradiation by the sample becomes negligible. As a result it is possible to increase the rate of modulation beam pulsing without significantly affecting the extent of variable fluorescence (Testing the FMS.). The FMS automatically performs this to maximise the signal to noise performance of the instrument. The extra data generated by this process is averaged to maintain an overall data logging rate of 10 readings per second.

IF THERE IS A LARGE DISTANCE BETWEEN THE FIBRE-OPTIC AND THE SAMPLE IT IS POSSIBLE THAT THE INCREASED MODULATION RATE USED

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WHEN THE ACTINIC LIGHT IS ACTIVE CAN BE USED TO IMPROVE THE SIGNAL TO NOISE RATIO WITHOUT INDUCING VARIABLE FLUORESCENCE IN DARKADAPTED SAMPLES. THIS CAN BE TRIED BY SWITCHING ON THE ACTINIC LIGHT WITH A SETTING OF ZERO INTENSITY.

3.7.2.3 The Saturating Light Source

The on-board LED Light Source is also used to deliver saturating pulses to the sample. The pulse can be set to deliver from 180 to 13,000 $\mu molm^{-2}s^{-1}$ selectable in 100 steps via the numeric value entered into the "Pulse" edit box (Figure 2-34). The minimum pulse duration is 0.3 sec with 0.1 sec increments up to a maximum of 5 sec; pulse duration is determined by the numeric value entered in the second "Pulse" edit box (Figure 2-34).

Pulses are initiated by pressing measuring buttons which require pulses (Measurement Functions.) or the "Pulse" button. Whilst the pulse lamp is active the button remains indented to signify that it is in use. The button is automatically re-set when the pulse terminates.

3.7.2.4 The Far-Red Light Source

Far-red light is provided by an LED source with typical peak wavelength 735 nm. It is activated via the "Far-red" button (Figure 2-34) in a similar manner to the actinic source. When active the LED operates at maximum output; the small output in relation to ambient or other actinic light sources means that care should be taken when using the far-red lamp to ensure that its effectiveness is not diminished by the presence of other light.

3.7.3 Measurement Functions

The FMS incorporates routines to execute commonly referenced fluorescence measurements. The most common fluorescence parameters can be classified into two distinct groups: those made on dark-adapted tissue and those made on tissue which is actively engaged in photosynthesis under ambient or FMS actinic light source. Both sets of measurement are initiated by pushing the appropriate button on the instrument toolbar (C, Figure 2-34).

3.7.3.1 Dark-adapted Measurement Routines

Dark-adaptation inhibits all light-dependent reactions. The resulting absence of photochemistry for a sufficient length of time allows complete re-oxidation of PSII electron acceptor molecules, opening PSII reaction centres and thus maximising the probability that absorbed light can be used for photochemistry. Commonly measured parameters from tissue in this state are used to calculate the maximum quantum efficiency of PSII and are usually used to reference measurements made on light adapted samples.

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Fo (fluorescence origin)

The fluorescence origin (Fo) is defined as the fluorescence yield following dark adaptation when all of the PSII reaction centres and electron acceptor molecules are fully oxidised and hence "open" for photochemistry. As a result Fo is often measured at the beginning of an experiment when only the modulating beam is illuminated.

The "Fo" button triggers data averaging over a 1.6 sec period with the result stored as Fo in the "Parameter Window". It is important to note that this function cannot determine whether the sample is correctly dark adapted. Hence if it is used in the presence of ambient light or if the output from the modulating beam is bright enough to drive significant photochemistry (Testing the FMS), the Fo value stored will not relate to the common definition of Fo.

Fm (fluorescence maximum)

The maximum fluorescence yield (Fm) is attained when the dark-adapted sample is exposed to an intense saturating pulse of light. This temporarily reduces all PSII electron acceptors preventing PSII photochemistry. The temporary absence of competition from photochemistry for absorbed energy ensures maximum fluorescence emission from the sample. The "Fm" button initiates a saturating pulse
of light using the intensity and duration settings specified in the respective "Pulse" edit boxes (The Saturating Light Source). During the pulse the fluorescence rise is monitored with the highest average of two consecutive points stored as the Fm value in the "Parameter Window". As a result the Fm value that is presented will be slightly different to the maximum fluorescence value observed in a tabulated output of the full data set. This procedure is used to minimise the effects of random noise on the accuracy of the measurement.

Fv/Fm (Maximum quantum efficiency of PSII photochemistry)

The difference between the Fo and Fm fluorescence yield relates the maximum capacity for photochemical energy quenching by the sample (Figure 2-35) and is defined as variable fluorescence (Fv).

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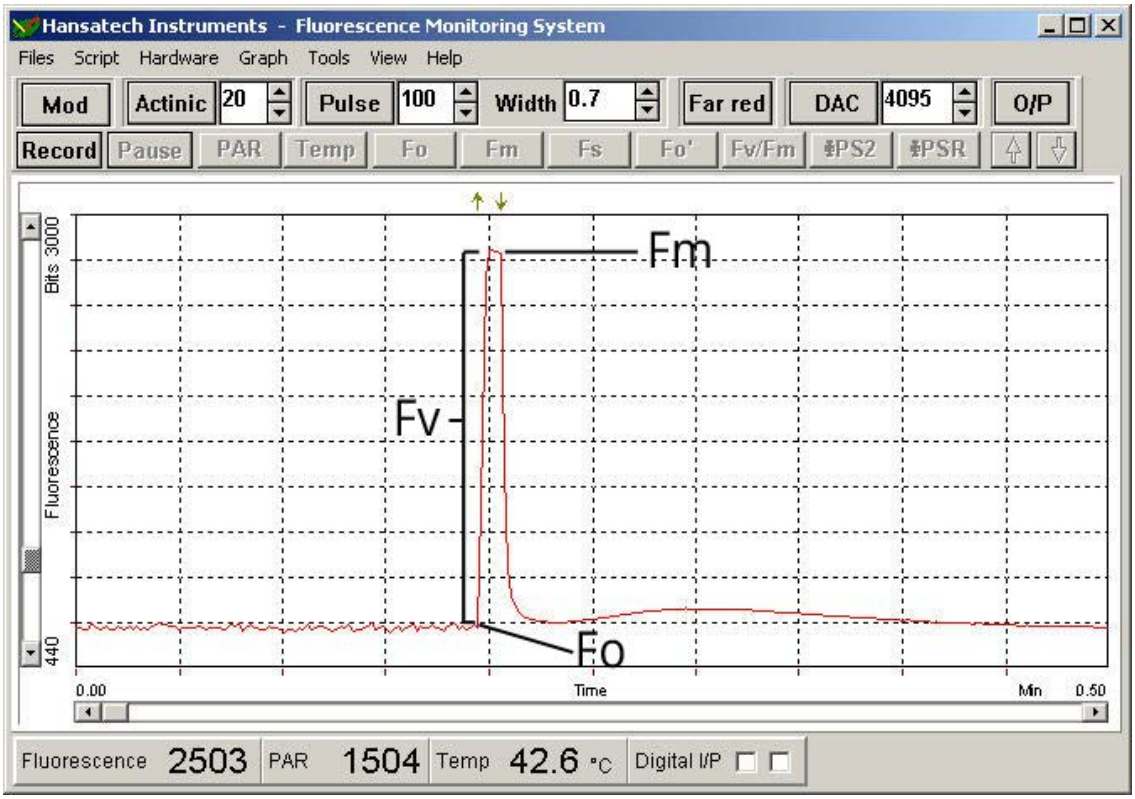

Fig. 2-35. Fluorescence trace following measurement of Fv/Fm

Calculation of the rate constants for competing energy dissipation pathways in tissue under dark-adapted (Fo) and light-saturated (Fm) conditions have shown that the ratio of variable to maximal fluorescence (Fv/Fm) is directly proportional to the quantum efficiency of PSII photochemistry (Butler 1977, 1978). Close correlation with other measures of quantum efficiency of photochemistry in a wide range of species (Björkman and Demmig 1987) has resulted in widespread use of Fv/Fm as a screening parameter for stress response.

The “Fv/Fm” button initiates a routine to measure Fo, Fm and calculate Fv, Fv/Fm. The routine operates in three stages:

1. The modulation beam only signal is averaged for 1.6 sec to determine Fo,
2. A saturation pulse is applied to determine Fm (Figure 2-35),
3. Fv and Fv/Fm are automatically calculated and presented with Fo and Fm in the “Parameters Window” (Figure 2-36).

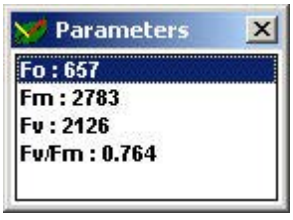

Fig. 2-36. “Parameters Window” following measurement of Fv/Fm

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3.7.3.2 Light-adapted measurement routines

In the case of light-adapted tissue, a proportion of PSII electron acceptors are reduced, “closing” some PSII reaction centres. Hence the probability that absorbed energy is used for photochemistry is not maximal as competing non-photochemical processes are operating (Section 1). The instrument toolbar allows access to routines which determine the relative extent of each of these processes. When coupled with measurement of the light-adapted ratio of variable to maximal fluorescence ratio they permit the estimation of PS II quantum efficiency (PSII) using the model of Genty et al. 1989.

The phi PSII function

Several measurements of fluorescence yield from the sample in different defined states are required to estimate ϕPSII. Initially the fluorescence yield of the sample under the ambient light regime is required (Figure 2-37). Such measurements are often made after a sample has adapted to a particular light-regime or environment and is operating at steady state. Consequently the measurement is often referred to as the “steady state” fluorescence yield, Fs (Figure 2-37). A saturating pulse is then required to “close” all of the PSII reaction centres; the temporary inhibition of PSII photochemistry ensures that the maximal fluorescence yield is achieved, Fm’ (Figure 2-37). If a previous dark-adapted measurement of Fv/Fm has been made the extent of photochemical and non-photochemical quenching processes can be determined from the equations of Schreiber et al. 1987.

The ϕPSII function automatically executes the protocol shown in boxed area of figure 2-37:

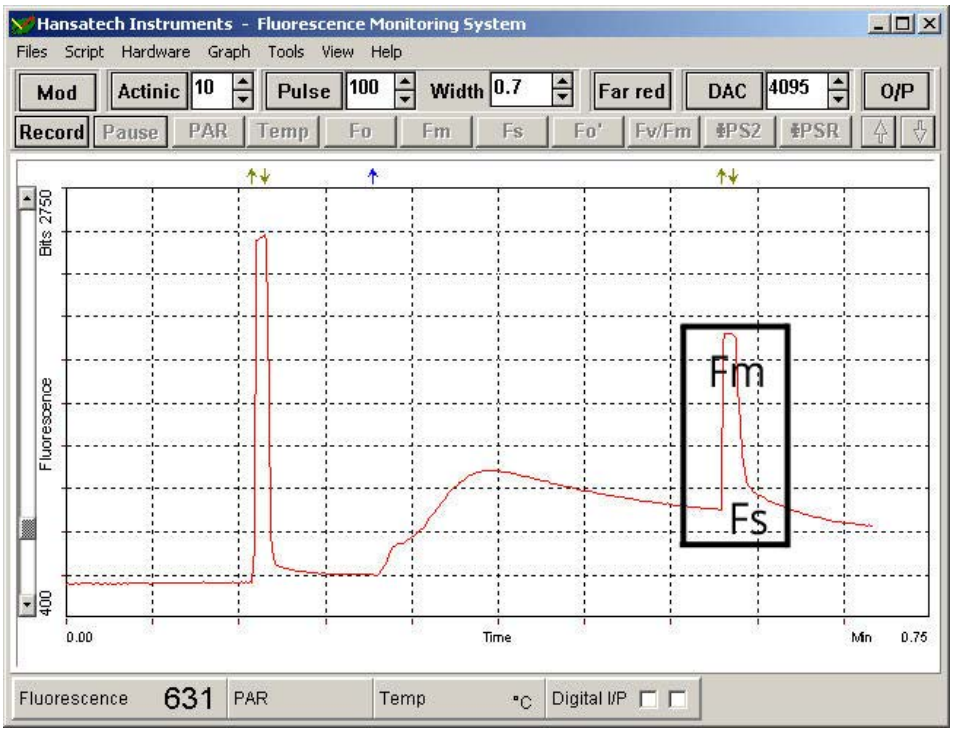

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Fig. 2-37. Fluorescence trace observed after Fv/Fm , adaptation to actinic light and ϕPSII measurement (boxed area).

1. Values of PAR and temperature are logged from the leaf-clip (these will be zero if the clip is not connected),
2. The steady state fluorescence value (Fs) is determined by averaging the fluorescence signal under the ambient light,
3. A saturation pulse, defined by the intensity and duration values in the "Pulse" edit boxes, is applied and Fm' stored as the average of the highest consecutive values. At the end of the function the following measured and calculated parameters are presented in the "Parameters Window":

Parameter	Measurement	Units	Derivation
PAR	Incident photosynthetically active radiation	$\mu molm^{-2}s^{-1}$	
Temp	Incident photosynthetically active radiation	°C	
Fs	Steady state fluorescence yield	Bits	
Fm'	light-adapted fluorescence maximum	Bits	
ϕPSII	quantum efficiency of PSII	No Units	= (Fm'-Fs) / Fm' (Genty et al. 1989)
qp	photochemical quenching co-efficient	No Units	(Fm'-Fs) / (Fm'- Fo)
qNP	Non-photochemical quenching co-efficient	No Units	= (Fm-Fm') / (Fm-Fo)
NPQ	Alternative definition of non-photochemical quenching	No Units	= (Fm-Fm') / Fm'
ETR	Electron transport rate	No Units	=PAR * 0.5 * ϕPSII * 0.84

It is important to note that in some of the derived parameters such as the quenching coefficients reference data that is not encompassed within the fPSII measurement function. As a result it is possible to make perfectly valid measurements of fPSII (which does not require referencing to data outside of the measurement function) whilst calculating invalid measurements of quenching coefficients (which require referencing to a dark-adapted measurement of Fo and Fm). Error trapping routines have been built into FMS which operate according to the following rules:

1. Fm must ≥ Fm'
2. Fs must ≥ Fo

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3. PAR ≥ 1 $\mu molm^{-2}s^{-1}$ (effectively
checks presence of the PAR sensor)
4. Fs must ≤ Fm'

If the measured values used during the calculation of a derived parameter do not obey the above rules any suspect calculated parameter will be flagged in the format "?? calculated number". The cause of such flags is usually easy to trace by manually calculating the parameter from the "raw" data that is always presented in the "Parameters Window".

The phi PSIIR function

Adaptation to high irradiance can involve significant changes in the conformation of the photosynthetic apparatus which result in non-photochemical energy dissipation in the PSII antennae, before energy reaches the reaction centres (Horton et al. 1991 Horton and Ruban 1994). Failure to account for this effect can lead to inaccuracies in calculation of the relative contributions of photochemical and non-photochemical energy dissipation. This problem can be overcome by transiently shading the sample and using a far-red light source to preferentially excite PSI relative to PSII; electrons
are drawn through the electron transport chain effectively "opening" PSII reaction centres and allowing measurement of a light-adapted Fo, usually given the notation Fo'. The ϕPSIIR function initiates the following sequence of events to make these measurements:

1. Values of PAR and temperature are logged from the leaf-clip (these will be zero if the clip is not connected),
2. The steady state fluorescence value (Fs) is determined by averaging the fluorescence signal under the ambient light,
3. A saturation pulse, defined by the intensity and duration values in the "Pulse" edit boxes, is applied and Fm' stored as the average of the highest consecutive values,
4. 3 sec after the pulse the sample should be temporarily dark-adapted by shading the sample (the FMS actinic source is automatically switched off), and a 5 sec far-red pulse is applied ,
5. The fluorescence decline is tracked during the far-red pulse with the minimum yield stored as Fo',
6. At the end of the function the FMS actinic source is switched back on.

The ϕPSIIR function calculates and writes the following parameters (using the measurement of Fo') to the "Parameters Window". The error trapping rules described in The phi-PSII function. are used during calculation of derived parameters with the addition Fo' must ≤ Fo.

Parameter	Measurement	Units	Derivation
PAR	Incident photosynthetically active radiation	$\mu molm^{-2}s^{-1}$	

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Temp	Incident photosynthetically active radiation	°C	
Fs	Steady state fluorescence yield	Bits	
Fm'	light-adapted fluorescence maximum	Bits	
ϕPSII	quantum efficiency of PSII	No Units	= (Fm'-Fs) / Fm' (Genty et al. 1989)
qp	photochemical quenching co-efficient	No Units	(Fm'-Fs) / (Fm'- Fo)
qNP	Non-photochemical quenching co-efficient	No Units	= (Fm-Fm') / (Fm-Fo)
NPQ	Alternative definition of non-photochemical quenching	No Units	= (Fm-Fm') / Fm'
ETR	Electron transport rate	No Units	=PAR * 0.5 * ϕPSII * 0.84

Fs

The steady state fluorescence yield (Fs) is automatically measured when the PSII and PSIIR functions are active. However, there may be occasions where it is desirable to log the Fs value without initiating a saturating pulse. This should be done by clicking the "Fs" measurement function. When activated the following 1.6 sec of data are averaged with the result written as Fs in the "Parameters Window".

Fo

Measurements of Fo' can be made independently of the PSIIR function by operating the "Fo'" button. When active the actinic light is switched off, far-red light initiated for 5 sec and the fluorescence decline tracked. The minimum value is written to the "Parameters Window" as Fo'. At the end of the far-red pulse the actinic light is switched back on to its original setting.

THE SAMPLE SHOULD BE SHADED FROM AMBIENT LIGHT DURING THE FAR-RED ILLUMINATION TO ENSURE THAT PREFERENTIAL PSI EXCITATION OCCURS. IF TOO MUCH AMBIENT LIGHT IS PRESENT THE EFFECT THE FAR-RED ILLUMINATION WILL BE SWAMPED.

Logging PAR and Temperature

The "PAR" and "Temp" functions on the instrument toolbar allow logging of incident PPFD and temperature at the sample. They write a latest measurement displayed on the instrument

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monitoring bar to the “Parameters Window”. It is therefore important that the instrument monitoring bar is displayed (View Functions). A value of zero will be recorded for both parameters if the leaf-clip electronics are
not connected to the control unit.

3.7.4 External Device Interface and Event Markers

The remaining buttons on the instrument toolbar control the external device interface and application of user-specified event markers.

3.7.4.1 External Control Interface

The FMS1 is provided with a digital control interface to aid interfacing of controls with third party equipment. It is possible to trigger external devices which are connected to line 0 directly from the instrument toolbar whilst experiments are being run using “O/P” button or during a script using the DIGIT element (Hansatech Scripting Language Elements).

This approach is suitable for triggering an external device to log an event in parallel with FMS. Conversely the FMS accepts up to four digital inputs which trigger the FMS in response to control from an external device. The status of input lines 0 and 1 is shown on the instrument monitoring bar during manual operation (B, Figure 2-1) and may be used to advance scripts that have been paused using the WDON, WOFF elements (Hansatech Scripting Language Elements).

A digital to analogue (D/A) line is also provided for control of devices such as external LED light sources which require variable voltage (0 to 4095 mV) control. The D/A line is toggled on / off from the “DAC” button with required voltage output (in mV) set in the adjacent edit box. Control is also possible during script operation using the DAC element with the required voltage as the element parameter. N.B. A DAC value of 0 switches off the channel.

Utilisation of either DAC or digital control lines requires appropriate wiring connections between instruments. Pin connections to the FMS are explained in The External User Interface . A bare ended connector cable is also available upon request from Hansatech Instruments Ltd.

3.7.4.2 Event Markers

Whilst recording data all FMS-controlled illumination events are event-marked with coloured arrows. The Up and Down arrow buttons on the instrument toolbar can be used to mark user-specified events such as addition of herbicide or change of temperature etc. Each time one of the event marker buttons is used an arrow is displayed above the data recording area and the corresponding fluorescence data in the tabulated output is coloured-coded for easy retrieval. The colour-codes may be customised to suit individual applications (Event Marker Colour).

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3.8 Hansatech Scripting Language Elements

The are a total of 32 script elements which make up the Hansatech Scripting Language. This page contains a complete listing of:

- The script editor icon,
- Its control functions,
- User-defined settings for respective control functions,
- And logged measurements that are saved to on board memory in “Local mode” or written to the “Parameters Window” under “PC” operation.

For clarity the list is separated into four groups of elements.

Instrument Control Elements - set instrument functions and light sources without logging data.

Element:	Editor Icon:	Function(s):	Required Setting(s):	Logged Measurement Parameter(s):
GAIN	None	Sets gain	Gain setting (1-100)	Gain setting (1-100)
MOD	None	Mod beam on	Mod level (0-3)	None
ID		Use plot ID in “Local mode”	N= Treatment no. M= Replicate no.	Plot location (Section 2.5.3.1.)
ACT		Actinic lamp	Intensity (0-50), zero switches off.	None
SAT		Sat lamp	Intensity (0-100), N.B. automatically shuts down after 5 sec if left on.	None
RED		Far- red source	N = 1 ON N = 0 OFF	None
BEEP		Beeper	Beep duration (sec)	None
DAC		User Interface Socket	Voltage output (mV)	None
DIG		User Interface Socket	N= 0 sets all lines off N= 1 sets line 1 N= 2 sets line 2 N= 4 sets line 3 N= 8 sets line 4 *e.g.* N= 12 sets lines 3 + 4	None
TEXT		Display text	20 characters max.	None
CLR		Remove text	None	None

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Script Pause Elements - halt script execution until a condition is fulfilled. The LOG function sets the system to log data during each pause for full trace analysis.

Element:	Editor Icon:	Function(s):	Required Setting(s):	Logged Measurement Parameter(s):
LOG		Store data during script pauses	N = 1 Log data N= 0 No logging	None
WAIT		Pause script for period of time	Pause duration (sec)	None
WKEY		Pause script until key-press	None	None
WMON		Pause script until key-press Display data during pause	None	None
WDON		Pause script until digital input goes high.	N= 1 waits for line 1 N= 2 waits for line 2 N= 4 waits for line 3 N= 8 waits for line 4 *e.g.* N= 12 waits for 3 + 4	None
WOFF		Pause script until digital input goes low.	N= 1 waits for line 1 N= 2 waits for line 2 N= 4 waits for line 3 N= 8 waits for line 4 *e.g.* N= 12 waits for 3 + 4	None
WLEV		Pause script until change in signal is less than N *i.e.* pause until steady state	N = max. size of signal change (bits)	None

Trace Logging Elements - used in conjunction with the PAR / temperature leaf-clip to log sample environment and basic parameters relating to the trace.

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Element:	Editor Icon:	Function(s):	Required Setting(s):	Logged Measurement Parameter(s):
PAR		Store PAR measurement	None	PAR reading
PARH		Store high range PAR measurement	None	PAR reading
TEMP		Store temperature measurement	None	TEMP reading
FLAV		Average data for period, Store average value	Duration of averaging (sec)	FLAV (average)
FMAX		Track signal over period, Store max value	Duration of tracking (sec)	FMAX (maximum)
FMIN		Track signal over period, Store min value	Duration of tracking (sec)	FMIN (minimum)
RATE		Track signal over period, Calculate and store rate of signal change	Duration of rate calculation (sec)	RATE

Fluorescence Parameter Logging Elements - automatically measure, calculate & store commonly referenced chlorophyll fluorescence parameters

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Element:	Editor Icon:	Function(s):	Required Setting(s):	Logged Measurement Parameter(s):
Fo	F0	Average data over period, Store average as Fo	Duration of signal averaging (sec)	Fo (Section 2.6.3.1.1.)
Fm	Fm	Sat Pulse Store max. signal as Fm	Duration of sat pulse (0-5 sec) Intensity of pulse (0-100)	Fm (Section 2.6.3.1.2.)
Fs	Fs	Average data over period, Store average as Fs	Duration of signal averaging (sec)	Fs (Section 2.6.3.2.3.)
Fo'	F0'	Actinic light off, Far-red pulse, Track signal, Store minimum signal as Fo'	Duration of signal tracking (sec)	Fo' (Section 2.6.3.2.4.)
FvFm	FvFm	Average data over period, Store average as Fo, Sat pulse Store max signal as Fm	Duration of element (sec) Intensity of saturating pulse (0-100) Pulse width (0-5 sec)	Fo, Fm, Fv/Fm (Section 2.6.3.1.3.)
ϕ_{PSII}	ΦPS2	Log PAR, Log TEMP, Average data over period, Store average as Fs, Sat pulse Store max signal as Fm'	Duration of element (sec) Intensity of saturating pulse (0-100) Pulse width (0-5 sec)	PAR, TEMP, Fs, Fm', Φ_{PSII} (Section 2.6.3.2.1.), QP, qNP, NPQ ETR
ϕ_{PSIIR}	ΦPSR	Log PAR, Log TEMP, Average data over period, Store average as Fs, Sat pulse Store max signal as Fm' Switch off actinic Far-red Pulse Track and store minimum signal as Fo' Re-apply actinic.	Duration of element (sec) Intensity of saturating pulse (0-100) Pulse width (0-5 sec)	PAR, TEMP, Fs, Fm', Fv', Fv'/Fm' ϕ_{PSII} (Section 2.6.3.2.2.), qP QNP NPQ ETR

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Section 3: Stand-Alone Instrument Operation and Data Upload

4 Section 3: Stand-Alone Instrument Operation and Data Upload

4.1 Local Mode" Operation Without a PC.

Once the FMS has been loaded with user-defined scripts it can be disconnected from the computer and the external power supply for operation outside of the laboratory. This mode of operation has been termed "Local mode" as instrument functions are accessed via the control unit's built-in firmware.

Work outside the laboratory is often carried out by relatively untrained personnel in sub-optimal environmental conditions. As a result the philosophy behind the FMS has been to prepare as much of the difficult work in the laboratory in the form of down-loaded scripts, with data gathering in the field controlled by single key-strokes from a simple menu system. User-input to the control unit is made via the four-button keypad on the display panel of the control unit; the function of each button being defined by the label displayed on each of the four lines of the LCD display. Thus as the user works through the menu structure the function of each key is up-dated. In practice the menu structure has been kept as simple as possible with affirmative actions generally initiated from the upper key and negative responses initiated from the lower key.

The FMS defaults to "Local mode" when it is switched on. A brief introductory screen presents the instrument version identification (essential information when contacting Hansatech Instruments Ltd for customer support) before entering the "Run" menu.

4.1.1 The "Run" Menu

The "Run" menu always defaults to the last script that was executed with the upper key reserved to initiate the script (Figure 3-1). (When the instrument is switched on for the first time it will automatically default to script number 1). It displays information on memory availability and the length of time it will take to complete the script.

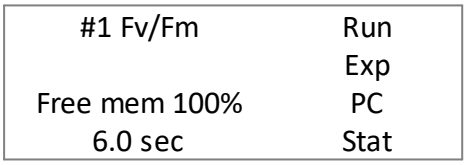

Fig. 3-1. Run menu

There are four options available to the user:

- Run - initiates the currently displayed script (Running a Script Under Windows.),
- Exp - choose an alternative script for execution (Selecting a Different Script),
- PC - set the control box as a slave in "PC mode",
- Stat - view instrument status (Checking Instrument Status).

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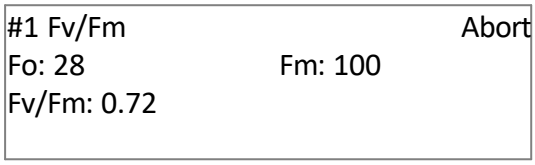
A flow diagram of the overall menu structure is shown in "Local Mode" Menu Structure.

4.1.1.1 Running the script

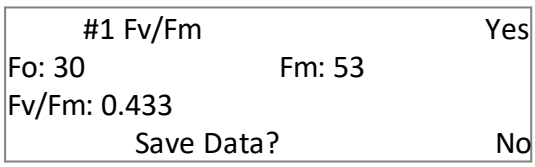
The script that is displayed in the "Run" menu is executed by pressing the "Run key". The FMS will then scroll through the script sequentially executing each of the functional elements. Those elements which record fluorescence parameters (fluorescence parameter logging elements, Hansatech Scripting Language Elements) write their respective parameters on the middle pair of lines on control unit display as the element is completed. This allows the operator to monitor the progress of the experiment and resume the script where key-presses are required to complete script functions (Hansatech Scripting Language Elements). In the example below the default Fv/Fm script is being operated.



The script consists of three lines:

GAIN: 50
MOD: 3
FVFM: 2.5, 100, 0.7

These set the gain, modulating beam and initiate the fluorescence measurement. The FVFM element is set to operate the element over a 2.5 second period, averaging the Fo signal for (2.5-0.7) = 1.8 sec before applying 100 unit saturating pulse over a 0.7 sec duration to determine Fm. Thus the Fo, Fm and calculated Fv/Fm values are presented at the end of the script. Any subsequent fluorescence logging element would then initiate and the displayed data would be replaced. Despite being replaced on screen, all of the data is stored to memory in "Full trace" mode (Checking Instrument Status.),
whilst "Parameters only" mode stores parameters without any trace data (Checking Instrument Status.). At the end of the script the operator must confirm the data storage in memory by selecting "Yes" at the "Save Data?" prompt.



When confirmed the data is given a file number and saved as "Full data" or "params only" depending upon the setting in the "Status" menu (Checking Instrument Status).

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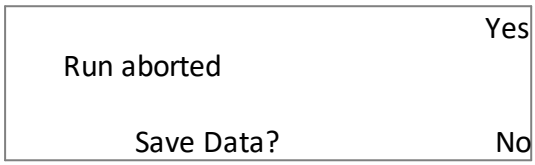
Throughout the script the FMS makes checks to determine that the instrument has sufficient power to finish the script and that the recorded signals are within the range of the A/D (Local Mode Error Codes). If the FMS fails one of these checks it automatically terminates the script and prompts to determine whether data should be saved from the partially completed script. A complete list of the error codes and the appropriate remedy is given in Local Mode Error Codes. In virtually all cases the memory will retain data for down-load to a PC. Even in the event of battery failure an internal capacitor will power the memory for several days.
Provision has also been made for the operator to terminate a script; the upper-key (labelled "Abort" during script execution) or the remote trigger on the leaf-clip terminates the script giving the option to store the partial data set.



4.1.1.2 Selecting a different script to run

There are six memory locations in the FMS control unit which can accommodate a single experiment each. Thus under "Local mode" operation it is important to be able to change location and execute any of the stored scripts. The "Exp" function under the "Run" menu initiates the "Select" menu which can be used to select a particular experiment for display and hence execution from the "Run" menu.

The "Next" key toggles forward through the six memory locations, incrementing the memory location number i.e. in the above example the next location is number 2, ϕPSII whilst the "Prev" key toggles backward, decreasing the memory location number. The title of each resident script is shown on the display; if a location does not contain a script the title display will show "* Empty *".

When the desired script is displayed it is selected by pressing the "OK" button. The FMS returns to an up-dated "Run" menu with the experiment location number and script title shown on the display.

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It is possible to exit the “Select” menu at any time without changing the script selection; the “Esc” (lower key) returns the instrument to the “Run” menu without changing the selected script.

4.1.1.3 Checking instrument status

The status of the FMS may be checked at any time between scripts from the “Run” menu by pressing the lower key (labelled “Stat”). The following information is displayed:

Full Data	← →
Files 10	
Free mem 99%	More
Battery 12.0V	Esc

“Full Data” refers to the information that is saved each time a script is executed. In “Full Data” mode all of the data is recorded during measurement elements plus associated parameters. When uploaded (Data Upload to PC) the data set is displayed as a chart recorder emulation in the data recording area with parameters written to the “Parameters Window”.

DATA IS ONLY RECORDED DURING SCRIPT ELEMENTS WHICH INVOLVE MEASUREMENTS. IF A FULL TRACE (INCLUDING FLUORESCENCE SIGNAL DURING SCRIPT PAUSE ELEMENTS) IS REQUIRED THE “LOG” SCRIPT ELEMENT MUST BE USED.

The default instrument setting stores full data information. However if fluorescence trace information is not a priority it is possible to save memory space by selecting “Params only” mode with the upper key (labelled ← →). In this mode only the measurement parameters associated with logging elements (Hansatech Scripting Language Elements) are recorded. They should be uploaded into “Parview” Uploading Parameter Data to Parview) feature is useful for screening procedures where the emphasis is on a high throughput of numerical information rather than in-depth analysis of fluorescence trace features.

The reduced requirement for data storage during the execution of each script also ensures that a much larger number of scripts can be performed between instrument down-loads.

The “Status” menu also shows the number of data files currently stored in memory, the available memory space and a spot reading of the instrument operating voltage. The instrument will automatically issue a warning message before shutting down if the operating voltage drops below 10.5 V

The “More” key displays a new screen with real-time updated values of fluorescence, PAR and temperature

Par: 1200	Esc
Temp: 10.1	

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This may be helpful for checking environmental conditions prior to executing a script. The lower key can be used to toggle the display back-light on and off to configure the display for maximum visibility.

4.1.2 Data Upload to PC

After “Local mode” operation it is sensible to up-load data from the control unit as soon as possible. This allows multiple copies of data to be generated for “back-up” purposes and enables clearance of instrument memory for further experimentation.

Full trace data must be uploaded into “Modfluor” as the trace plotting functions are required to display the chart-recorder emulation of trace data. Parameter-only data should be uploaded into “Parview” which has facilities to tabulate data from multiple replicates (Uploading Parameter Data to Parview).

Before data transfer can begin the PC serial link to the control unit must be connected and the unit set as a slave device for interrogation by the computer in “PC mode”.

This is done via the “PC” button in the “Run” menu. Further information to troubleshoot communications problems may be found in Software Operation Section of this manual (Section 2).

4.1.2.1 Uploading Trace Data to “Modfluor"

There are two different methods for transferring data from the control unit. Individual data files may be selected by file storage number for display on screen (Upload to Screen) or multiple files may be selected by script type and stored to sequential data files on disc (Upload to File). Both data transfer functions are located under the Files group on the “drop-down” menu.

Upload to Screen

Individual data files may be up-loaded for display in the data recording screen with the Upload to screen function. When this function is selected both the data and script used to generate the data are loaded into the FMS software together. As a result the software will initially prompt to verify that any existing script can be over-written.

The “OK” button should be pressed to proceed with the transfer. The next stage is to enter the ID number of the file to be transferred into the “Select File” prompt which is generated (Figure 3-2).

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Fig. 3-2. Select file prompt box.

The file ID number refers to the number given to the data when it was saved during "Local mode" operation. File numbers are issued chronologically in the sequence that recorded data is saved to memory as files. Hence in the above example, file number 1 contains data which was gathered when the first script was executed in "Local mode".

Once a valid file number has been entered the computer will interrogate the instrument's memory to retrieve a copy of both the appropriate data set and the experimental script that was used to generate the data. If "full data" mode has been used trace data is presented in the data recording area. The "Parameters Window" is up-dated with the file number, date and time of script execution and associated parameters.

The data is then available to further analysis, storage or export via a "drop-down" menu in "Modfluor" software. The script that was used to generate the data can be viewed or refined with the "Script Editor".

EDITED SCRIPTS CANNOT BE DOWN-LOADED INTO THE CONTROL UNIT TO OVER-WRITE EXISTING SCRIPTS UNTIL ALL OF THE DATA HAS BEEN CLEARED. BEFORE DOWN-LOADING EDITED SCRIPTS FOR FURTHER EXPERIMENTATION IT IS ADVISABLE TO DOWN-LOAD THE INSTRUMENT AND CLEAR MEMORY.

Upload to File

The Upload to screen function only allows single files to be retrieved in each data transfer session. Multiple files may be retrieved in a single session using the Upload to file function. The following file selection screen is generated (Figure 3-3):

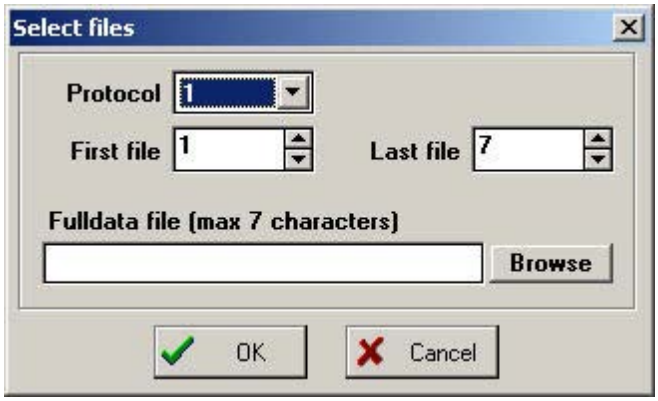

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Fig. 3-3. Select files prompt box from the Upload to files function.

Files may be selected by protocol type (i.e. the script that was used to generate the data) or as a complete memory upload by opening the protocol edit box and selecting the “All” option. Data retrieval can be limited to a specified range of files by setting retrieval limits in the “First file” and “Last file” edit boxes.

A destination file / file series must be specified on the PC. The desired name should be entered into the fulldata file box. The first file uploaded will be saved to name01.dat, second to name02.dat etc. Each of these files can be loaded into the software and reviewed individually.

4.1.2.2 Uploading Parameter Data To “Parview”

“Parview” is a stand-alone utility designed to allow easy upload and transfer of multiple parameter files to a spreadsheet program. After initialisation the default parameter screen is loaded (Figure 3-4).

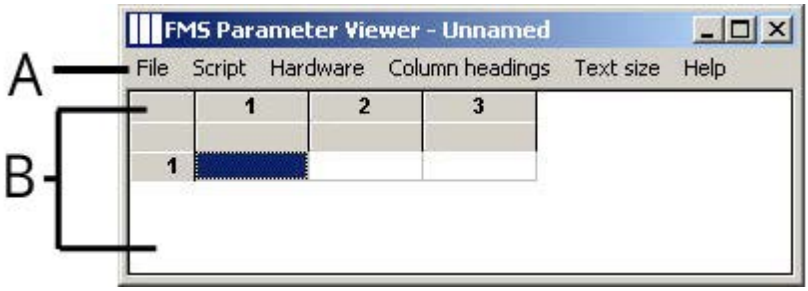

Fig. 3-4. The “Parview” default parameter screen with main menu (A) and parameter display area (B)

Data access and manipulation functions are located under the main menu (A, Figure 3-4) with parameters presented in spreadsheet form in the parameter display area beneath (B, Figure 3-4). The data upload and storage functions are located under the Files menu.

The File Menu

The Files menu contains the following data handling options:

File

- Clear
- Load data
- Save data
- Upload params
- Text Search
- Convert to ASCII
- Print

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Exit

Initially data must be opened within the program before parameters can be viewed. This can be achieved by searching and loading previously saved parameter (*.PAR) files from disc with the Text search and Load data functions. These operate in the same manner as “Modfluor” (Section 2) Alternatively new data must be uploaded from the control unit with the Upload params function.

Upload params

When selected the Upload params function interrogates the control unit to list the instruments memory contents. Files can be uploaded by protocol type with defined ranges using the “first file” and “last file” settings (Figure 3-5).

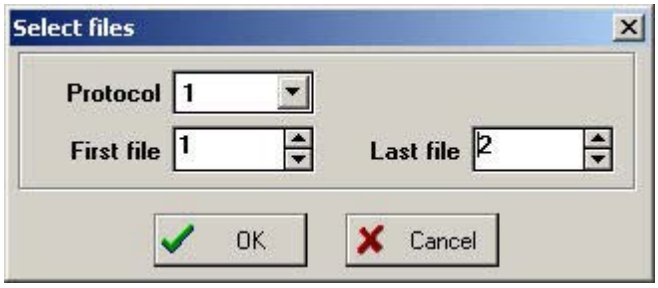

Fig. 3-5. “Select files” for upload prompt.

When “OK” is pressed the selected files are uploaded to the “Parview” program. At this stage only three columns of file identification parameters are displayed in columns within the parameter display area; experimental replicates are loaded by row (Figure 3-6).

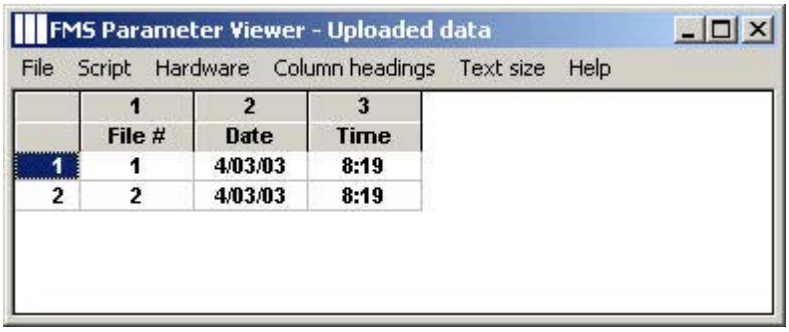

Fig. 3-6. Parameter presentation after upload. The file identification parameters of File number, Date of execution and Time are presented in three columns. Replicates are presented by row.

At this stage the full set of parameters plus a copy of the script used to generate them are resident in PC memory. They may be stored to disc as a *.PAR file using the Save data function.

The parameter display area must be configured to display further parameters. Appropriate functions are located under the Column headings menu.

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Configuring the Parameters Display

When a script is executed the various parameters associated with each script element are stored in the sequence in which they were measured. Each parameter can be assigned to a column within the parameter display area; “Parview” can display a maximum table of 255 parameter columns by 12,800 replicate rows. Parameters are assigned to columns via the “Set Parameter Columns” prompt box (Figure 3-7).

Fig. 3-7. Set Parameters Prompt Box.

In the above example the script used to generate the data contains a simple Fv/Fm measurement. The default setting on upload assigns columns 1,2 and 3 to display File #, Date and Time respectively. The Autoset button assigns columns 4 through 255 to the next 255 parameters in the sequence. If the sequence contains more than 255 parameters blocks of 255 parameters may be displayed at a time (e.g. parameters 1 - 255 then 256 - 510) by following the prompts that are generated.

Individual types of parameter can be included or excluded from the data set by pressing the Parameters button. This generates a list of types of parameters measured during a script which is split into those parameters types that ate currently displayed (Used) and those which have been excluded (Available) (Figure 3-8).

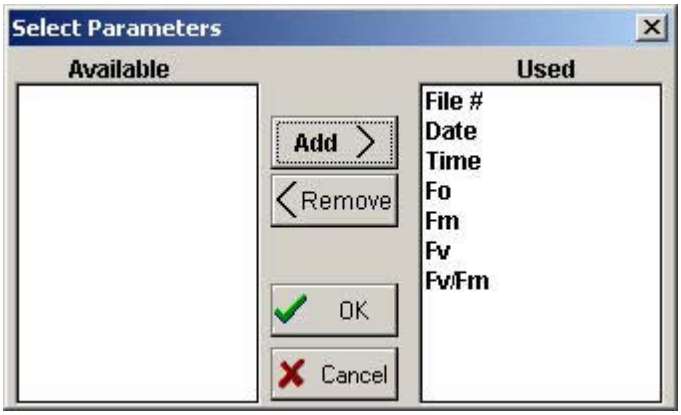

Fig. 3-8. Selecting which parameter types are displayed.

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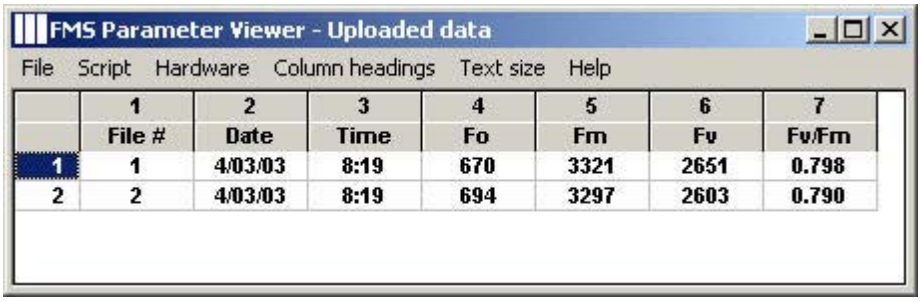
Specific parameter types may be selected with the mouse and transferred between lists with the appropriate “Add” and “Remove” buttons. The example above displays all of the parameters recorded during the Fv/Fm script (Figure 3-9).



	1	2	3	4	5	6	7
	File #	Date	Time	Fo	Fm	Fv	Fv/Fm
1	1	4/03/03	8:19	670	3321	2651	0.798
2	2	4/03/03	8:19	694	3297	2603	0.790

Fig. 3-9. Displayed parameters after parameter selection.

The font size of the displayed text may be adjusted to improve readability with the Text size command on the main menu.

Parameter Data Export From “Parview”

ASCII for export into other data analysis applications. This can be achieved with the Convert to ASCII command under the Files menu. The command operates in a similar manner to that used in “Modfluor” (Section 2) and generates a comma de-limited file containing all of the data in the parameter display area. Such files may be opened or imported into most common spreadsheet packages such as “Excel” or “Lotus”.

IT IS IMPORTANT TO NOTE THAT PARAMETERS WHICH HAVE BEEN EXCLUDED FROM THE DISPLAY AREA ARE NOT INCLUDED IN THE ASCII FILE OUTPUT. THE ASCII FILE OUTPUT WILL ONLY CONTAIN DISPLAYED PARAMETER TYPES.

If more than 255 parameters have been measured during a script it will be necessary to export the data in at least two ASCII files. The first 255 parameters should be configured and exported to file1.asc then the display should be re-configured to show parameters 256 through 510. The latter display should be exported to file2.asc.

Other “Parview” Functions

The Files menu additionally contains a Print option which allows all complete rows and columns of data visible within the current window view to be printed at the selected Text Font size.

The main menu also contains commands to view and edit scripts (Script), and to check the status of the control unit (Hardware). Both of these commands operate in the same manner as the “Modfluor” program and are described in Section 2.

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4.2 “Local Mode” Error Codes

The FMS contains several functions which check both the status of the instrument and the validity of the data that is collected. A brief list of possible error codes and their appropriate remedies is given below.

System Errors

Error:	Problem:	Action:
Battery Low	Battery potential has dropped below 10.5 V. The current script has been aborted. It is not possible execute further scripts.	Connect external 12 V supply, re-charge battery by connecting mains power supply.
Memory Full	Data storage memory full. The current script is aborted and no further scripts may be run. It is possible to save parameters from the aborted script.	Upload data to a PC and clear instrument memory.
Memory Fail	The control unit has been subject to a hard re-set or the data has become corrupted.	Clear instrument memory. Re-load scripts. If problem persists contact Hansatech Instruments
HANSA01 + HANSA02	Hardware device failures	Please contact Hansatech Instruments

Data Errors

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Error:	Problem:	Action:
Overscale	Saturation pulse signal exceeds 4095 bits	Reduce gain, increase fibre-optic / sample distance or reduce modulating beam frequency.
Underscale	Modulating beam only signal (Fo) or far-red (Fo') signal is less than 10 bits	Increase gain, modulating beam intensity or bring sample closer to fibre-optic.
Fm < Fo	Sample absent of moved during measurement.	Check orientation of sample.

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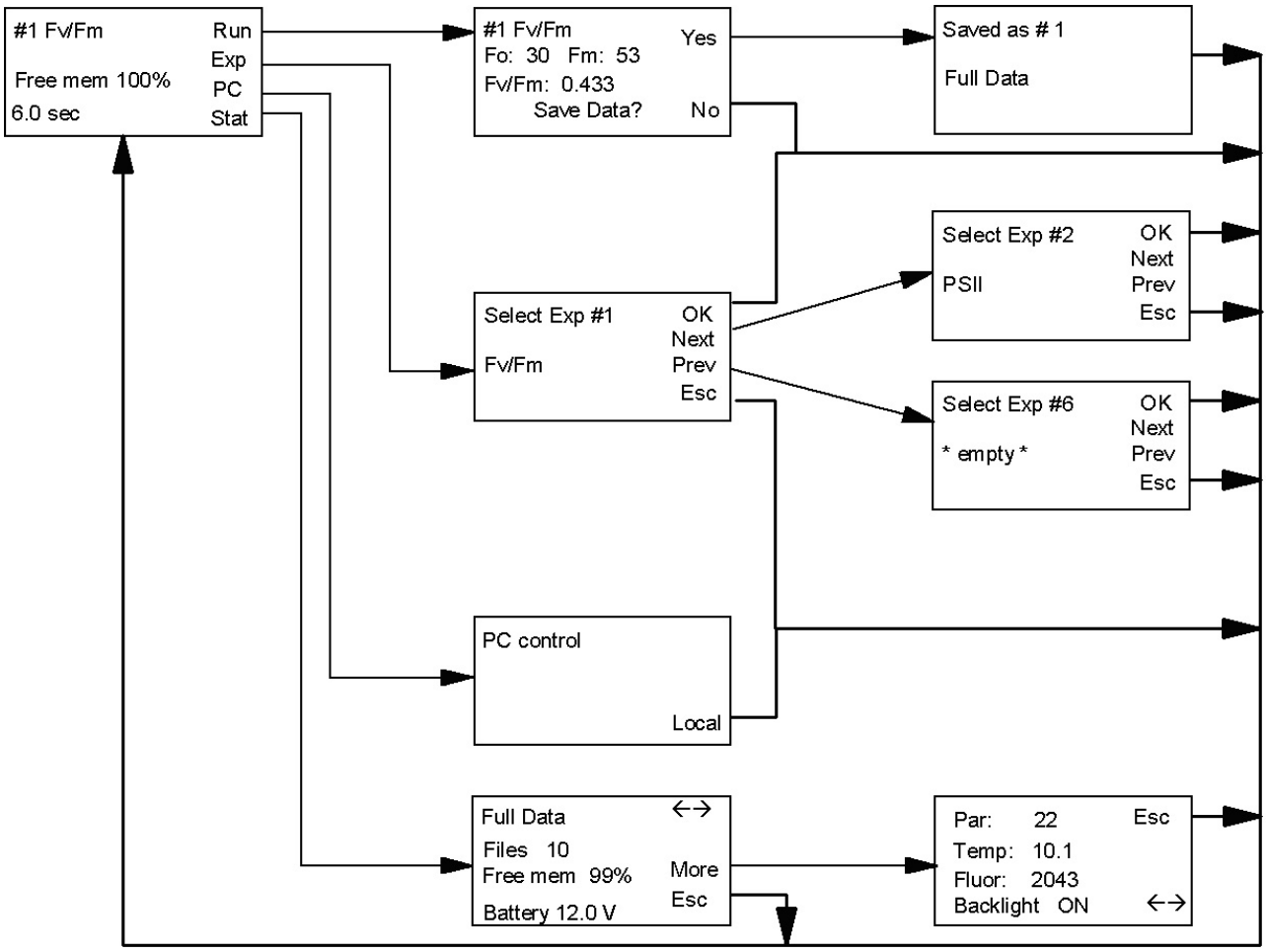
4.3 "Local Mode" Menu Structure



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Horton, P. and A. Ruban (1994). The role of light-harvesting complex II in energy quenching. In *Photoinhibition of photosynthesis from molecular mechanisms to the field* ed. N. R. Baker and J. R. Bowyer pp. 111-128. Oxford: BIOS Scientific Publishers Ltd.
Kautsky, H. and A. Hirsch (1931). Chlorophyllfluoreszenz und Kohlensäureassimilation. *Naturwissenschaften*, 19, 964. Krause, G. H. and E. Weis (1984). Chlorophyll fluorescence as a tool in plant physiology. II. Interpretation of fluorescence signals. *Photosynthesis Research*, 5, 139-157.
Krause, G. H. and E. Weis (1991). Chlorophyll fluorescence and photosynthesis: the basics. *Annual Review of Plant Physiology and Plant Molecular Biology*, 42, 313-349. Ögren, E. and N. R. Baker (1985). Evaluation of a technique for the measurement of chlorophyll fluorescence from leaves exposed to continuous white light. *Plant, Cell and Environment*, 8, 539-547. Schreiber, U., Schliwa, W. and U. Bilger (1986). Continuous recording of photochemical and non-photochemical chlorophyll fluorescence quenching with a new type of modulation fluorimeter. *Photosynthesis Research*, 10, 51-62.
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