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Leaflab 2+

System Manual

Hansatech Instruments

Leaflab 2+ System Manual

by Hansatech Instruments Ltd

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Leaflab 2+ System

1 Leaflab 2+ System

1.1 Introduction to Oxylab + Electrode Control Unit

The Oxylab+ oxygen electrode control unit is designed to provide PC control of oxygen uptake or evolution measurements across a broad range of applications from studies of mitochondrial and cellular respiration to measurements of isolated chloroplast suspensions or solid state leaf samples in photosynthesis research. In conjunction with user-friendly OxyTrace+ data acquisition and system configuration software, the Oxylab+ control unit provides an effective tool for the measurement of oxygen signals from the S1 Clark type oxygen electrode disc mounted in one of a range of liquid and gas-phase electrode chambers with quick and easy system calibration and configuration. Simultaneous recording of an optional auxiliary input signal (e.g. temperature, pH, chlorophyll fluorescence, TPP+ or other specific ion electrodes etc) is also possible using the appropriate apparatus.

A system comprises a maximum of 8 Oxylab+ control units linked via USB to a Windows® PC. The control unit features a built-in magnetic stirrer (for liquid-phase measurements) and all the electronics required to control and measure the signal from the oxygen electrode disc.

A maximum of 2 Oxylab+ systems can be used at any one time to perform quantum yield analysis.

The control unit also offers a blue LED screen to quickly view values such as set temperature, actual temperature, auxiliary and oxygen readings.

Oxylab+ is compatible with all existing liquid and gas-phase Hansatech oxygen electrode chambers and accessories. In addition, Oxylab+ features light source control capabilities providing automated intensity changes of Hansatech Instruments LED light sources LH11/2R and LH36/2R. Additional tools within the OxyTrace+ software package allow editing of PFD (photon flux density)

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tables providing complete automation of complex light intensity changes during light response curves.

OxyTrace+ software controls all major hardware and data acquisition functions with simple calibration routines for both liquid and gas-phase measurements. Data from the electrode disc, optional auxiliary input signal and temperature signal from the QTP1 PAR/Temperature probe sensor are plotted as a chart recorder emulations in "real-time" with post-measurement data analysis tools included within the program. Completed experiments are saved in Comma Separated Values (*.csv) format which then may be opened directly in other Windows® data analysis applications such as Excel®.

1.2 Leaflab 2 + System Inventory

What you have received and what additional items you will require.

Upon delivery, please check the items you have received against the checklist below. If you find that an item is missing, please contact Hansatech Instruments as soon as possible.

- OXYL1 - Oxylab electrode control unit
- LD2/3 - Oxygen electrode chamber
- S1 - Oxygen electrode disc
- QSRED - Large are PAR sensor for calibration of LH36/2R
- LH36/2R - 36 red LED light source
- S1/SMB - SMB-SMB Electrode cable
- A2 - Electrode disc membrane applicator tool
- S4 - 30m reel of PTFE membrane
- S9B - Spare O-rings set for LD2/3
- S14 - Set of 3 stainless steel discs, capillary matting and foam spacer
- S15 - 3-way gas taps and 1ml disposable syringe (packet of 4 taps+ 2 syringes).
- S16 - Electrode cleaning kit
- USB 2.0 A to B - 2m USB cable with A-plug to B-plug

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1.3 S1 Electrode Disc

1.3.1 Introduction

1.3.1.1 Introduction

The S1 electrode was designed in the early 1970's during a collaborative project between Prof. David Walker, Prof. Tom Delieu and Hansatech Instruments. The first S1 went on sale in 1974 and has remained largely unchanged in over 30 years. A range of complementary instruments have been developed over the years in order to meet the needs of emerging trends in the scientific world. Modern oxygen measurement systems incorporate the S1 electrode providing sensitive and reliable tools for a vast range of applications ranging from the demonstration of oxygen evolution from photosynthetic organisms to the analysis of mitochondrial respiration rates.

The Hansatech Instruments oxygen electrode disc is a specialised form of electrochemical cell known as a Clark type polarographic sensor which comprises a resin bonded central platinum cathode and a concentric silver anode. During measurements a 50% saturated potassium chloride (KCl) electrolyte solution forms a bridge between the 2 electrode elements via the aid of a fine paper wick and a fine layer of PTFE membrane which is selectively permeable to oxygen molecules. These are applied to the electrode disc during the preparation process and are held in place with an O-ring. Once preparation of the electrode is complete, the disc assembly is mounted into the base of an electrode chamber with the membrane covered dome section of the disc forming the floor of the reaction vessel. The electrode disc is connected to a control unit which supplies a polarising voltage of 700mV. In the presence of oxygen, a polarised disc generates a small current which is directly proportional to the concentration of oxygen in the reaction vessel above the cathode. This current is converted to a voltage, conditioned and digitised with high resolution before display by the electrode control unit.

1.3.1.2 Electrode Disc Theory

The oxygen electrode is a specialised form of electrochemical cell which consists of two electrodes immersed in an electrolyte solution. Typically, a 50% saturated solution of KCl is used in oxygen electrode systems. This is prepared by dissolving 17.5g of anhydrous salt in 100ml of de-ionised water at 25°C. Application of a polarising voltage of 700 mV ionises the electrolyte and initiates current flow via a series of electrochemical reactions. In the case of KCl electrolyte the following reactions occur:

Equation 1.

$$Ag \longrightarrow Ag^{+} + e^{-}$$
$$\downarrow$$
$$Ag^{+} + Cl^{-} \longrightarrow AgCl$$

Equation 2.

$$O_2 + 2H_2O + 2e^{-} \longrightarrow H_2O_2 + OH^{-}$$
$$\downarrow$$
$$H_2O_2 + 2e^{-} \longrightarrow 2OH^{-}$$

Oxygen is consumed during the electrochemistry, thus the magnitude of the current flow is related to the oxygen concentration of the surrounding media. This type of electrode

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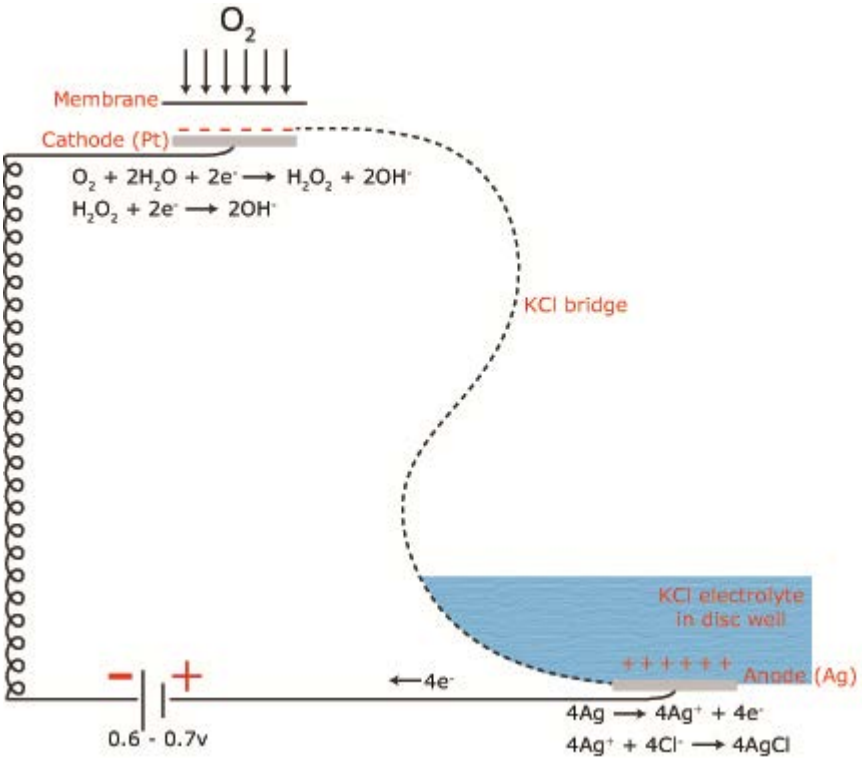
sensor was first developed by Clark (1956) to measure oxygen in blood samples. As a result it is often referred to as The Clark Electrode.

1.3.1.3 Operational Principles

The following text and image are based on the chapter "The Principle of Oxygen Measurement" from the book "The Use of the Oxygen Electrode & Fluorescence Probes in Simple Measurements of Photosynthesis" by Prof. David A. Walker (Oxygraphics Ltd. 1987). Hansatech Instruments oxygen electrode systems for photosynthesis and respiration measurements take one of 2 forms, liquid or gas-phase. The measurement techniques for each form are quite different, however, the underlying principles of the measurement of oxygen remain the same. Oxygen dissolved in the reaction vessel of liquid-phase systems or that which accumulates in the sample chamber of gas-phase systems is detected polarographically by either the S1 or S1/MINI (in Oxytherm oxygen measurement systems) Clark type electrodes (Clark, 1956). Both oxygen electrode discs comprise a relatively large (2mm diameter) platinum cathode and a concentric silver anode immersed in, and linked by, an electrolyte solution. Both electrodes are set in an epoxy resin disc; the cathode at the centre of a dome and the silver anode in a circular groove named the well or electrolyte reservoir, surrounding the dome. The electrodes are protected by a thin PTFE (Teflon) membrane which is permeable to oxygen. The purpose of the dome is to stretch the membrane smoothly over the surface of the platinum cathode and to allow it to be secured in position by an O-ring. The membrane also traps a thin layer of electrolyte (a solution, which usually contains potassium chloride) over the surface of the electrodes. A paper spacer is placed beneath the membrane as a wick in order to provide a uniform layer of electrolyte between anode and cathode.



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When a small voltage is applied across these electrodes, so that the platinum is made negative with respect to the silver, the current which flows is at first negligible and the platinum becomes polarised (i.e. it adopts the externally applied potential). As this potential is increased to 700 mV, oxygen is reduced at the platinum surface, initially to hydrogen peroxide H2O2 so that the polarity tends to discharge as electrons are donated to oxygen (which acts as an electron acceptor). The current which then flows is stoichiometrically related to the oxygen consumed at the cathode. The diagram above represents the oxygen electrode reactions. When a potentiating voltage is applied across the two electrodes, the platinum (Pt) becomes negative (i.e. becomes the cathode), and the silver (Ag) becomes positive (the anode). Oxygen diffuses through the membrane and is reduced at the cathode surface so that a current flows through the circuit (which is completed by a thin layer of KCl solution or other electrolyte). The silver is oxidised and silver chloride deposited on the anode. The current which is generated bears a direct, stoichiometric, relationship to the oxygen reduced and is converted to a digital signal and recorded by an electrode control unit.

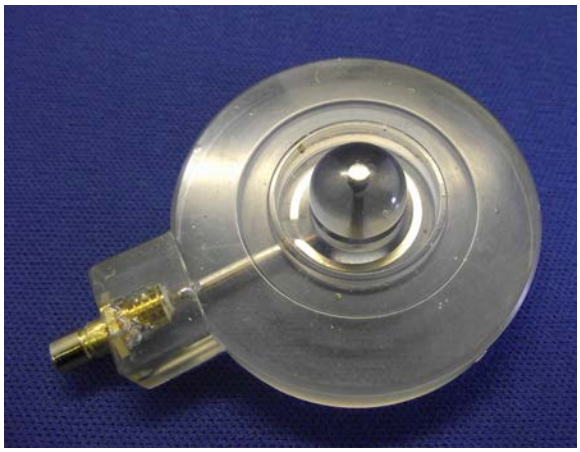
1.3.2 Electrode Preparation

1.3.2.1 Electrode Disc Preparation

Before use, the electrode disc needs to be prepared in such away that an electrolyte bridge is formed between the anode and cathode in order for current to flow in the presence of oxygen. Various different compositions of electrolyte have been used however, Hansatech Instruments recommend a 50% saturated solution of potassium chloride which is proven to work well in many different applications.

Stage 1

Ensure the electrode disc is clean paying particular attention that the silver anode is free from any deposits of silver chloride (brown deposit) or silver oxide (black deposit). If traces of these substances are present, clean the electrode thoroughly according to our recommended cleaning procedure.



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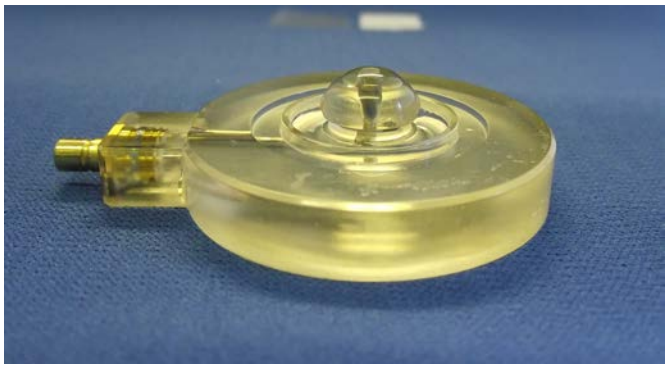
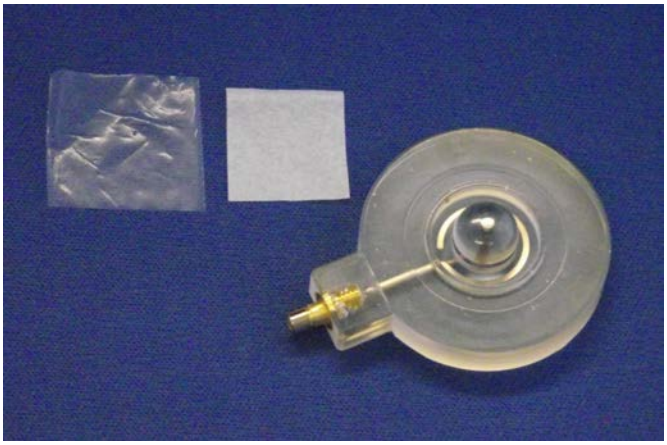
Stage 2

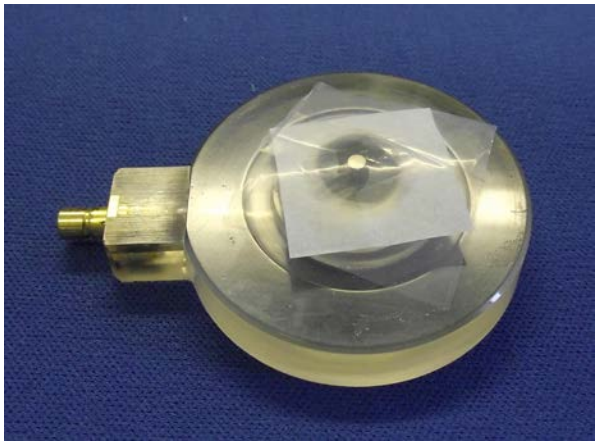
Stage 2 Using a Pasteur pipette, place a small drop of electrolyte on top of the dome of the electrode disc. Place a further 3 - 4 drops of electrolyte in the well of the electrode disc ensuring that the silver anode is completely covered



Stage 3

Place a 1.5 - 2cm2 paper spacer over the drop of electrolyte on the electrode dome. When complete, this paper spacer will act as a wick to form an electrolyte bridge between the anode and cathode. It is therefore critical that the spacer paper is of the correct size. From over 35 years of experience, Hansatech Instruments recommend using ultra-fine cigarette paper such as Rizla Blue. This product works particularly well as it is manufactured to a very tight thickness tolerance ensuring a good, even layer of electrolyte. When cutting spacer paper from Rizla Blue cigarette papers, take care not to use the gummed or folded section of the paper. If the folded section is used, ensure that this is at the very edge of the square spacer. Holding the PTFE membrane with a pair of forceps, use a sharp knife (or scissors) to cut a slightly larger section of membrane from the reel. Take care not to touch the membrane excessively as grease from finger prints can restrict the performance of the electrode. Carefully lay the section of membrane across the spacer paper on the dome of the electrode.





Stage 4

Stage 4 Place the small electrode disc O-ring over the end of the membrane applicator tool. Hold the applicator vertically so that the O-ring end of the applicator locates very gently over the dome

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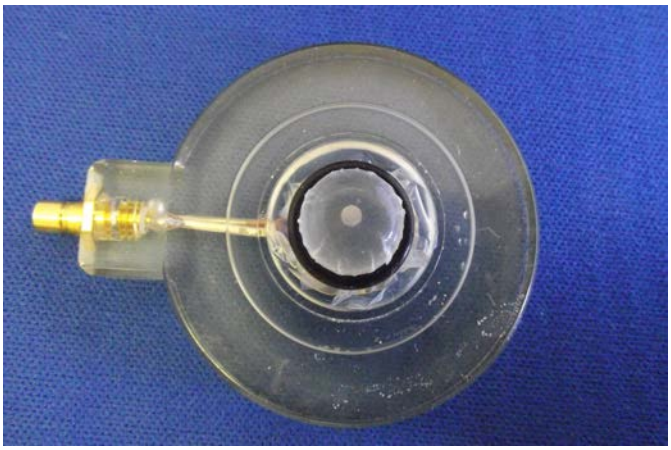
so as not to damage the PTFE membrane. In one smooth motion, push the applicator downwards so that the O-ring slides over the electrode dome and holds the spacer and membrane securely in place. Check that the membrane and spacer are smooth and free from trapped air bubbles, particularly over the platinum cathode itself. If the preparation is uneven, it will have an overall effect on the response time and sensitivity of the electrode. Also ensure that the spacer paper is in contact with the electrolyte in the well. If necessary, top the well up with a few drops of electrolyte ensuring that the silver anode is completely covered by the KCl.



Stage 5

Place the larger O-ring on the recess around the electrolyte well. If this second O-ring is not in place when the disc is installed into the electrode chamber, the silver anode will not be sealed from ambient air and measurements may be affected. Take care to ensure that the edges of the membrane and/or spacer paper do not contact the larger O-ring as this could affect the seal when the disc is installed into the electrode chamber. If the corners of the membrane and/or spacer paper protrude too much from the preparation, these may be trimmed carefully with a pair of sharp scissors or tucked carefully into the electrode well.

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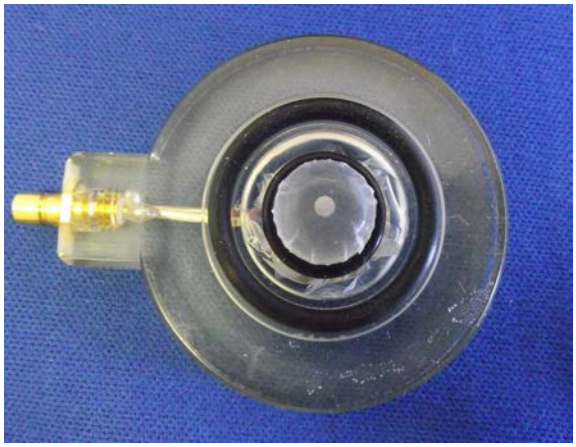

In Summary...

The finished, prepared oxygen electrode disc should have a smooth covering of spacer paper and PTFE membrane across the electrode dome. The electrolyte well should be topped up with KCl ensuring that some of the spacer paper is in contact with the electrolyte itself. Ensure everything is done correctly by cross referencing against the checklist below:

- The electrode disc is clean,
- The electrolyte is a 50% saturated potassium chloride
- 1 drop of electrolyte on the platinum cathode
- 3 - 4 drops of electrolyte in the well
- Spacer paper is 1.5 - 2cm2
- Spacer is not cut from the creased or gummed section
- Spacer paper placed centrally on the dome
- PTFE membrane cut slightly larger than spacer paper
- PTFE handled with forceps
- PTFE place centrally over spacer paper on the dome
- Smaller O-ring applied using membrane applicator tool
- Secured membrane is smooth and free from creases and bubbles across the platinum cathode
- There is sufficient electrolyte in the well to allow contact with the spacer paper
- Excess PTFE membrane is trimmed
- Larger O-ring placed in recess around the well
- Larger O-ring is not in contact with excess PTFE membrane

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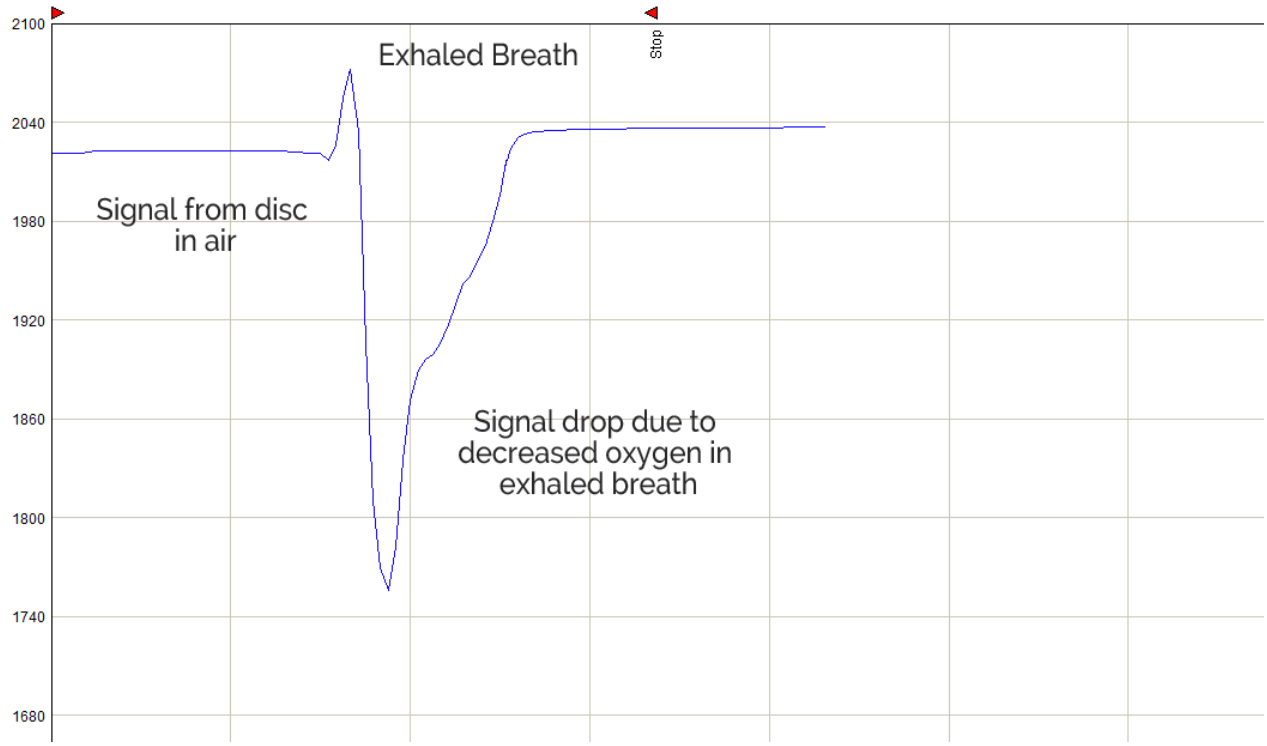
1.3.2.2 Testing the Response of the Electrode Disc

Once the electrode disc has been correctly prepared, it is advisable to check the response of the disc prior to mounting the disc within the electrode chamber.

Before proceeding, you will need to ensure that the electrode control unit you are using is correctly connected to the PC and that it is communicating with the OxyTrace+ software. If you are experiencing problems with communications between the control unit and software, please refer to the Software Initialisation section of this document.

To check the response, connect the electrode disc to the rear of the control unit at the electrode input using the S1/ADL electrode connection cable. Start a recording. The signal will initially be extremely high and displayed as nanoamps (nA) and will take a few minutes to stabilise at an air line level. A new or well maintained disc should read approx. 1600 nA in air but this may vary between ±240nA. Once the electrode signal is stable, breathe exhaled air across the disc and observe the reaction on the graph screen. The signal should be plotted as in the diagram below.



The first deviation in signal after breathing across the disc is induced due to a large increase in temperature of exhaled breath compared to ambient air temperature. Oxygen electrode discs are particularly sensitive to temperature and will show an increase in signal as a result. Since the temperature increase is only temporary, the oxygen signal will fall after a short time.

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After the temperature related signal increase, a steep drop in signal is observed due to decreased oxygen levels in exhaled breath (approx 17%). The signal should the begin to return to the original level as the ambient oxygen begins to equilibrate around the electrode disc.
If the observed signal is not as shown above, it may be caused by an inadequate electrode preparation or worst case, a problem with the disc itself. Try cleaning the disc and re-preparing before repeating the test described above. If problems still occur, please contact Hansatech Instruments.

Once this test has been completed satisfactorily, mount the disc into the base of the electrode chamber. An additional test may be carried out in order to test the response of the disc in situ. Please refer to the Operating the Magnetic Stirrer section for further details.

1.3.3 Electrode Disc Cleaning

1.3.3.1 Electrode Disc Maintenance

Although the oxygen electrode disc is one of the smallest components of an oxygen measurement system, it is the most crucial component of the system. Therefore, maintaining the disc to a high standard is extremely important. A significant proportion of technical support requests received by Hansatech Instruments Ltd relate to oxygen electrode disc performance. Of these requests, the vast majority are the result of failure to comply with recommended working practices and maintenance procedures. Some of the more common abuses of electrode discs include:

Failure to Maintain Electrode Disc

If the electrode disc is not maintained sufficiently, deposits of silver chloride (soft brown deposit) and/ or silver oxide (hard black deposit) will build up on the silver ring (anode) of the disc. In the image below, deposits of black silver oxide can clearly be seen on the silver anode of the left electrode disc. By contrast, the electrode disc on the right has a well maintained, clean anode. The formation of silver chloride is normal and desirable in small quantities as it is an electrical conductor and therefore improves the stability and sensitivity of the electrode disc. It is easily removed by cleaning and will reform when the electrode is next polarised. However, deposits of black silver oxide are to be avoided. Silver oxide is an electrical insulator and its formation will reduce the available surface area of the silver anode and result in a dramatic reduction or complete loss of electrode signal. The electrode disc should be maintained in accordance with our recommended procedure. Please refer to the Electrode Cleaning section.

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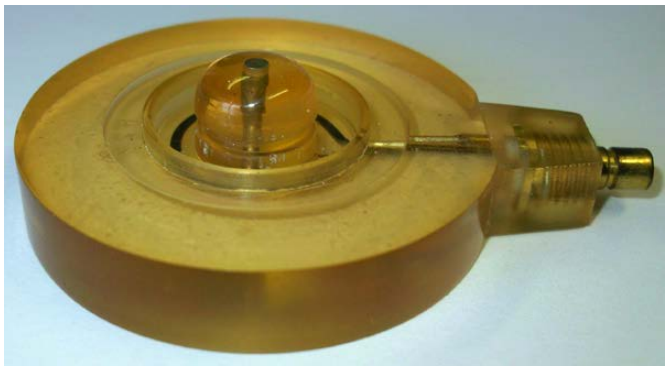
This image shows a typical case of poor maintenance. The disc has not been cleaned after use and was not stored in a sealed container with desiccant away from strong sunlight. As a consequence, the epoxy resin has become prematurely discoloured due to a combination of strong sunlight and absorption of moisture from the environment. The silver anode is almost entirely covered with a deposit of black silver oxide. The platinum stud and dome are both dirty and badly scratched. This will cause an uneven layer of electrolyte beneath the membrane during measurement leading to unstable and unresponsive signals. This disc is beyond economic repair.



In contrast, this disc has been maintained according to our recommendations and has been stored correctly in an air tight container with desiccant. The epoxy resin is clear, the silver anode is smooth and polished and the platinum cathode is scratch free. The area of the electrode dome surrounding the cathode is also free from abrasions and pits across the entire surface. An electrode disc, when kept in this kind of condition will provide years of high quality measurements

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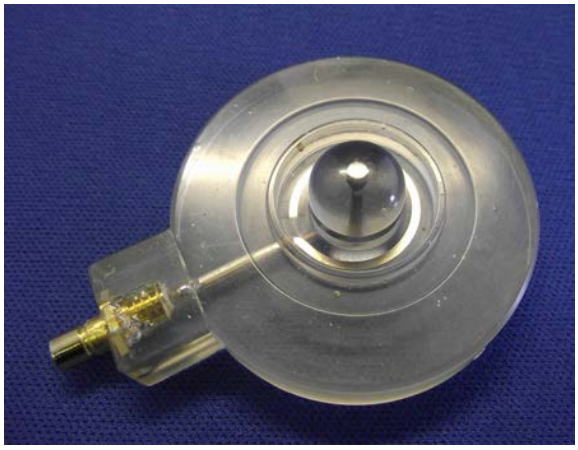
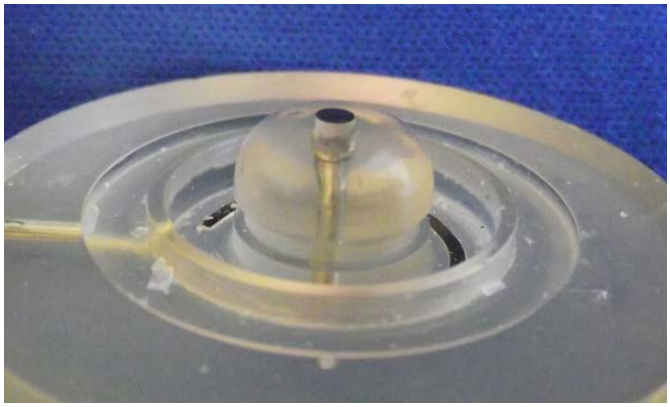


Disc is Left to Dry Out After Use

With the electrode disc un-polarised, the KCl electrolyte dries out quickly in air and begins to crystallise. KCl is quite aggressive and will, by capillary attraction, deposit in any minor crevice. If the electrode is left to dry out several times, it is possible that the KCl will finally break down the seal between either of the metal electrodes and the epoxy resin surround and seep into the fissure formed where it will crystallise to further expand the fissure.



This is most likely to occur between the platinum cathode and epoxy resin dome and regrettably will cause most damage at this point. Damage to the electrode can be observed under low power microscopy (x10 magnification) as a broad white collar around the neck of the platinum as shown in the image above. The effect of this will be to cause instability and drift of the electrode signal and a large increase in the level of electrical signal observed in zero oxygen (residual current). This will have the effect of reducing the ability of the electrode to measure small changes in oxygen flux and will eventually render the electrode unserviceable and beyond economic repair. The disc will require replacement.

After use, the electrode disc should be stripped down, cleaned carefully according to our recommended procedures and stored in a dry, air-tight container with added desiccant.

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It is possible to leave electrode discs in liquid-phase systems prepared over-night PROVIDED it remains polarised and is left mounted in the electrode chamber with stirred air-saturated water in the reaction vessel. In gas-phase measurements, the electrode is likely to dry out more rapidly and we would recommend stripping down, cleaning and storing the electrode after each measurement day.

Cleaning with Unsuitable Materials

Hansatech Instruments Ltd offer the S16 cleaning kit for maintenance of the oxygen electrode disc. The kit contains special cleaning pastes which are recommended for optimal electrode performance.

In the absence of this kit, users have resorted to cleaning the electrode with various abrasive compounds including commercial metal polishes or smoker's toothpastes etc. We would caution against the use of inappropriate materials as they may cause irreversible damage to the electrode. Some metal polishes are harmful to the disc as they can contain an ammonia base or solvent additive. These substances can cause irreversible poisoning to the platinum cathode and could also cause significant damage to the epoxy resin. Excessive cleaning with abrasive substances can badly scratch and alter the profile of the electrode dome which may cause an uneven layer of electrolyte across the cathode leading to instability, drift and lack of responsiveness of the oxygen signal. The S16 electrode cleaning kit is supplied with all Oxygraph and Oxytherm Systems and can also be obtained by contacting us directly. Details of the cleaning procedure can be found in the Cleaning Procedure section.

1.3.3.2 Cleaning the Electrode Disc

Periodic maintenance is required if you are to maintain your electrode in good condition. It should be cleaned after use and before prolonged storage. It is particularly important that the electrode is never left to dry out with electrolyte in place, as crystallisation of electrolyte may cause the platinum/epoxy resin seal to be breached and crystalline electrolyte to be deposited around the cathode. If this occurs, the electrode will become rapidly unserviceable and will require replacement. Cleaning is best achieved by following the procedure outlined below. (An electrode maintenance kit -ordering code S16- containing the items described is available. Please contact Hansatech Instruments for further details). It is important to use the correct polishing agent as

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harsh abrasive substances will damage the resin in which the electrodes are set. It should also be noted that many commercial metal polishes contain ammonia or solvents which may cause irreversible poisoning of the platinum cathode.

Cleaning Procedure

Silver Anode

The silver electrode (anode) is subject to electrochemical deposition of chloride and oxide salts during use. This typically manifests as a brown/black deposit which forms on the surface of the Silver. Whilst deposition of small amounts of brown Silver Chloride is normal and desirable, excessive deposits or deposits of black oxides will cause a rapid deterioration in electrode performance and may result in lower signal levels, signal drift and increased signal noise. Select a small cotton bud from the maintenance kit. Cotton buds vary a little in size, so check to see that the diameter of the cotton bud will allow it to be fully inserted into the well of the electrode, thus allowing contact with the silver electrode in the base of the well. If the bud is a little large, carefully reduce the size of the cotton bud by removing a layer of cotton wool from the tip until the bud will fit the well. Moisten the bud with a little distilled water. Apply a small amount of either the Rapid Hansatech Polishing paste to the tip of the bud. Insert the bud into the well of the electrode and gripping the bud just above the tip, apply moderate pressure and gently rotate the bud around the electrode well in a circular motion 6 - 10 times. Observe the Silver anode. Continue to gently polish the silver electrode until all brown or black deposits on the surface of the silver are removed and the surface of the silver is highly polished.

Platinum Cathode

The platinum cathode is not subject to such deposition and all that is necessary is to maintain a scratch-free highly polished finish using the procedure outlined below. Apply a small amount of the Rapid Hansatech Polishing paste (revised cleaning kits supplied from 01/10/01) or the No. 2 Fine Hansatech Polishing paste (earlier 2 part cleaning kits with Coarse and Fine pastes) to the unused end of the cotton bud moistened with a little distilled water , or if preferred to a small piece of moist cotton wool. Now, using a circular motion, gently polish the platinum cathode located in the centre of the electrode dome. It is important to restrict the polishing motion to the platinum and avoid as much as possible contact with the epoxy resin surrounding the platinum. Excessive polishing of the cathode area is to be avoided as this will eventually lead to a change of curvature of the electrode dome, which will result in deterioration of electrode performance. When finished, the platinum cathode should have a “mirror finish”. If the surface of the platinum has become scratched, repeat the above procedure using the Rapid Hansatech Polishing paste (revised cleaning kits supplied from 01/10/01) or first using the No. 1 Coarse Hansatech Polishing paste, followed by the No. 2 Fine Polishing paste (earlier 2 part cleaning kits with Coarse and Fine pastes).

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1.3.3.3 Electrode Disc Rinsing, Drying & Storage

Rinsing & Drying

Once the electrodes have been satisfactorily cleaned, it is important to remove all traces of the polishing paste from the electrode surfaces by rinsing the electrode with a small volume of distilled water whilst gently scrubbing these areas with a soft bristled brush. (A toothbrush is ideal). Avoid wetting the electrical connector of the disc during this procedure. Dry the electrode thoroughly using paper towel.

Storage

Store the electrode, when not in use, in an air-tight container along with a suitable desiccant such as Silica gel. Periodically replace or rejuvenate the desiccant.

1.3.3.4 Leaving the Electrode Disc Polarised Overnight

At the end of a day's experiments, care of the used equipment not only constitutes good laboratory practice but is also crucial to the longevity of the equipment, particularly the electrode disc. Once the experiments are finished, there are 2 procedures that may be followed to ensure that the disc is kept in good working condition:

Apparatus is dismantled

The reaction mixture and magnetic follower (or leaf disc and supports in gas phase measurements) should be removed from the electrode chamber. Any hazardous substances should be disposed of according to any COSHH guidelines in practice in the lab. The disc should be removed from the chamber base and disconnected from the electronics.

Carefully remove the smaller O-ring, membrane and spacer paper with a pair of forceps paying particular attention to not scratching the electrode dome. An uneven surface across the dome results in an uneven layer of electrolyte leading to unstable oxygen signals in future measurements.

The disc should be rinsed thoroughly with deionised water and dried (avoiding contact with the electrode cable connection in the case of Oxygraph+ users. Using the S16 cleaning kit (supplied) and following the guidelines set out in the Electrode Cleaning and Maintenance section of this document, clean the silver anode of the disc in order to remove any silver oxide (black deposit) or silver chloride (brown deposit) which may have tarnished the silver during the electrochemical reactions that occur during polarisation.

Once thoroughly cleaned and rinsed, the disc should be placed in an air tight, desiccated container and stored away from direct sunlight until needed.

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Apparatus remains polarised

It is feasible to leave the disc set up over night if measurements are to continue the following day. However, strict guidelines must be observed in order to prevent irreversible damage to the electrode disc. Please note that the following procedure is only suitable for liquid-phase measurements.

Once the final measurement of the day is complete, remove any reaction mixture from the chamber and replace with approx 1.5ml of deionised water.

Replace the magnetic follower and set the stirrer to On. It is important to leave the disc polarised in stirred, deionised water if the disc is to be left overnight in order to prevent the disc from drying out. The effects of drying out are described in the Electrode Cleaning and Maintenance section of this document.

When starting measurements again the following day, it is advisable to re-check the disc calibration by first adding a sample of air-saturated deionised water and ensuring that the oxygen signal compares to the Oxygen Calibration Table published by Truesdale & Downing (Nature 173:1236, 1954), followed by adding sodium dithionite or bubbling nitrogen gas into the reaction chamber in order to check the zero oxygen line.

If the steps described above indicate the disc preparation is still in a good condition, measurements may continue. However, if the measured signals show deviations in the expected signals, it is advisable to either recalibrate or in some circumstances, it may be necessary to dismantle the disc for cleaning purposes and re-preparation.

1.4 LD2/3 Oxygen Electrode Chamber

1.4.1 Introduction

1.4.1.1 Introduction to LD2/3

The LD2/3 leaf-disc electrode chamber allows oxygen uptake / evolution measurements to be performed from leaf-discs, excised needles, algae, mosses, lichens etc with a surface area of up to 10cm2. A prepared S1 oxygen electrode disc is mounted directly below the sample chamber with the dome of the electrode forming the chamber floor. LD2/3 features an upper and lower water jacket ensuring superior temperature control of the sample and S1 electrode disc when linked to a temperature controlled circulating water bath. The leaf chamber section has 2 gas ports providing both a calibration and flow-through capability for rapid changes in the gaseous environment above the sample. An additional tapped and stoppered hole is provided for the introduction of an optional temperature sensor or similar auxiliary sensor.

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A clear cast acrylic top window allows illumination of the sample via the LH36/2R LED light source with 1 additional optical port provided for either additional illumination or insertion of a quantum sensor, etc. A further port is orientated more vertically towards the sample and allows the fibre optic cable from the FMS 1 and FMS 2 modulated fluorometers to be positioned close to the sample allowing simultaneous measurement of chlorophyll fluorescence.

Maximum chamber volume: 9.5cm^3

1.4.1.2 Features of the LD2/3

A - Cast Acrylic Top Window
B - Upper Water Jacket
C - Upper Water Jacket Connectors
D - Securing Clasps
E - Gas Port
F - Leaf Chamber
G - Lower Water Jacket Connectors
H - Lower Water Jacket
I - Optical Port
(Not Pictured) FMS Fibre-Optic Adapter

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1.4.2 Setting up the LD2/3

1.4.2.1 Installing the S1 Electrode Disc in an Electrode Chamber

Before installing the electrode disc into the chamber, it should be correctly prepared according to the guidelines set out in the Electrode Preparation section.
Once the disc has been prepared and suitably tested, it is now ready to be installed in the electrode chamber.

Remove the base ring from the electrode chamber and place the prepared electrode disc onto the top.

Offer the base ring and disc assembly up into the underside of the main body of the electrode chamber so that the electrode cable connection fits with the key way on the chamber base thread. Gently hold the base in position while threading the base ring onto the electrode chamber so that it is slightly more than finger tight.

It is important not to over-tighten the base ring as this will cause the membrane to stretch over the electrode dome. During measurement, when the membrane starts to relax, the measured signal will drift.

Ensure that the electrode remains polarised in order to prevent the electrode from drying out whilst other preparations are made prior to measurement.

1.4.2.2 Checking for Leaks

Once an electrode disc is giving a stable signal, the electrode response may be used to test the system for leaks. Fit the electrode disc into the base of the electrode chamber as described above. Insert a gas-tight syringe into one of the gas taps whilst ensuring that the second gas-tap is in the closed position. Inject or remove air (typically 1 ml) from the chamber using the syringe. Either way, a rapid positive or negative response should be observed and no further drift should be detected. It should be noted that the electrode signal should be very stable if the tap is open. If excessive drift occurs when the tap is open then the electrode has not yet stabilised (some downward drift always occurs when the potentiating voltage is first applied but if this has not fallen to negligible proportions within half an hour then the electrode itself may require attention).

Removing 1 ml of air from the chamber will induce a rapid negative response in electrode signal. Once the signal re-stabilises, no further drift should occur (A in diagram below). A gradual upward drift (B in diagram below) may indicate a leak in the chamber allowing ambient air entry.

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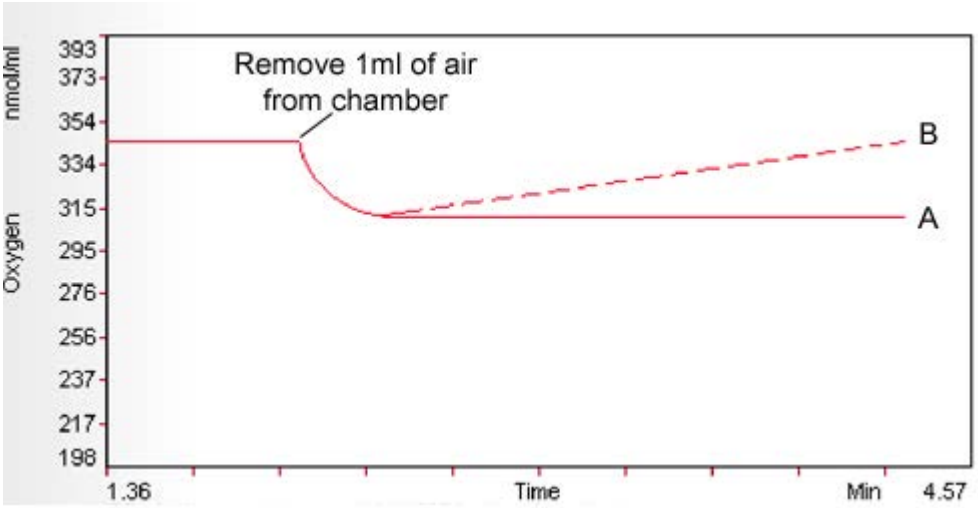

When the tap is closed, some downward drift will always be observed at higher gain but, in normal use and normal gain, it should be negligible. It is caused by oxygen consumption at the cathode. It can be diminished by using less amplification but not without loss of sensitivity (see e.g. Delieu & Walker, New Phytol. 71, 201-225 (1972)).

If the electrode chamber leaks then the only likely possibilities are that one or more of the /-rings involved in sealing the chamber are insufficiently compressed, incorrectly seated or worn. The O-rings are located as shown in the diagram below:

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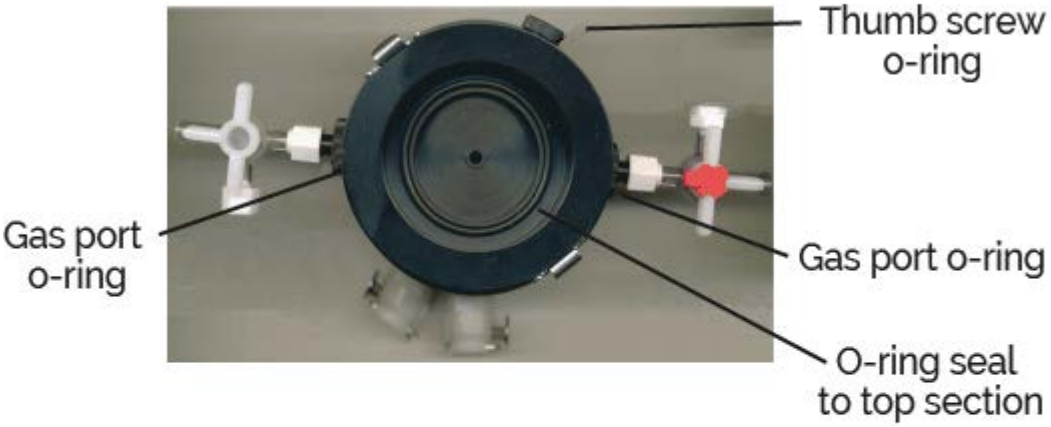
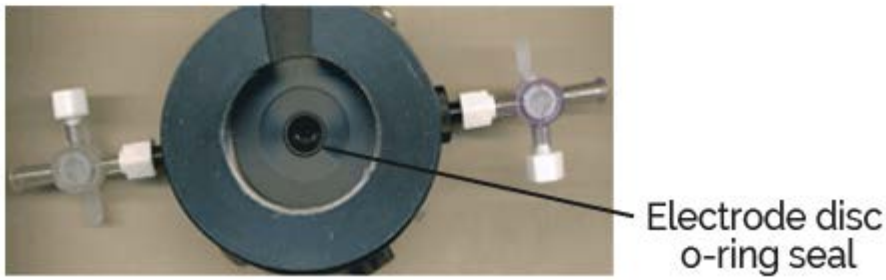
Middle section of LD2/3 electrode chamber. Top view



Middle section of LD2/3 electrode chamber. Bottom view



Also, check that the luer fit between the gas tap and port is properly made and gas tight. This can be checked by removing the suspect tap and repeating the leak test with the syringe fitted directly to the gas-port.
It is also important to ensure that the 2 O-rings on the prepared electrode disc are in place. If the leak is still not detected once all the O-rings have been checked / replaced, please follow these guidelines carefully however it should be noted that this practice should be used as a last resort.

Inject several ml of air into the chamber and close the gas tap. Disconnect the electrode connection cable and submerge the entire electrode chamber in a bowl of water. Rising bubbles will show the location of the leak.

IT IS CRITICAL THAT THE CONNECTOR OF THE ELECTRODE DISC IS DRIED THOROUGHLY BEFORE MEASUREMENTS PROCEED AS LIQUID IN THE CONNECTOR WILL CAUSE SHORT CIRCUITS.

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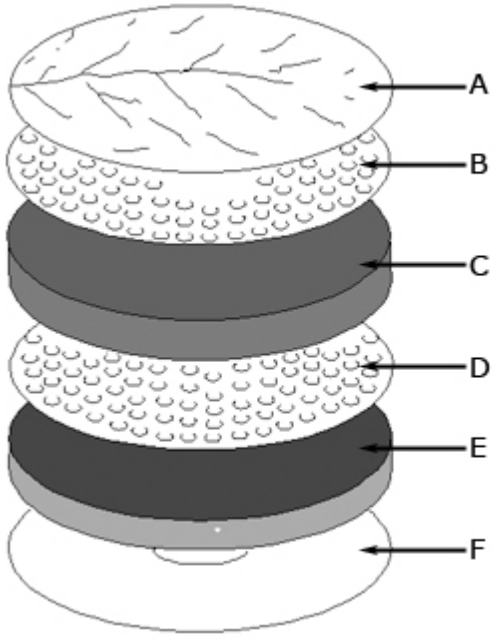
1.4.2.3 Preparing the Sample

Typically a 10 cm^2 leaf disc is cut from the sample material using the circular cutting tool provided and placed on a sandwich of materials which is inserted into the reaction chamber as shown in the diagram below.



*In the LD2/3 electrode chamber the leaf disc (**A**) is supported by a sandwich of materials. Beneath the leaf is a stainless steel grid an unperforated centre (**B**), layer of capillary matting (**C**), stainless steel grid with open centre (**D**), sponge spacer (**E**) and a stainless steel washer (**F**) which locates nearest the electrode disc.*

The purpose of the sandwich of materials is to support the leaf sample above the cathode whilst still permitting adequate diffusion of evolved oxygen through the floor of the reaction chamber to the cathode of the electrode disc.
A stainless steel washer with a hole in the centre locates around the cathode to provide a foundation for the other layers. These consist of a sponge spacer and a layer of bicarbonate treated capillary matting (please refer to the Maintaining Carbon Dioxide Inside the Reaction Chamber section) separated by a stainless steel grid. A further stainless steel grid with a closed centre supports the leaf sample above the capillary matting to allow gas exchange by the leaf. The closed centre in the steel grid prevents actinic light transmitted through the leaf interfering with the electrochemical reaction which occur at the cathode.

The sandwich of materials is loaded into the reaction chamber using the loading tool to align the layers. The loading tool should be seated around the edge of the chamber whilst the layers are gently lowered in sequence through the centre of the tool.

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A stainless-steel leaf disc cutter is provided. This should be used in conjunction with a soft rubber pad held behind the leaf during cutting and should not be allowed to become blunt. The cut disc has an area of 10cm2. The size of the disc ensures that the proportion of cut cells to intact cells is negligible. Once the sample is in place, the loading tool should be removed carefully to avoid disturbing the alignment. The O-ring which seals the reaction chamber is then added around the edge of the sandwich and the top of the chamber lowered and sealed using the clips on the side of the chamber. If a 10 cm^2 leaf disc has been used the upper surface of the leaf sample should fill the visible area of the window of the chamber. If this is not the case the alignment of the sandwich has been disturbed which will result in failure to illuminate all of the leaf area properly. The chamber should be opened and the sandwich re-aligned with the loading tool.

SMALLER SAMPLES WITH AN AREA LESS THAN 10 CM2 CAN BE MEASURED IN THE ELECTRODE CHAMBER. FOR PROPER ILLUMINATION THEY SHOULD BE PLACED IN THE CENTRE OF THE REACTION CHAMBER AT THE TOP OF THE SANDWICH. IT IS ADVISABLE TO MEASURE THE AREA OF SAMPLE PRIOR TO INSERTION SO THAT MEASUREMENTS OF OXYGEN FLUX CAN BE NORMALISED ON A UNIT AREA BASIS. IT SHOULD BE REMEMBERED THAT SMALL SAMPLES WILL GENERALLY YIELD SMALLER OXYGEN SIGNALS WHICH COULD BE HARDER TO RESOLVE. A COMMON TECHNIQUE TO OVERCOME THIS WITH LONG THIN LEAFED SAMPLES SUCH AS GRASS SPECIES IS TO FILL THE 10 CM2 AREA WITH A NUMBER OF LEAVES ADJACENT TO EACH OTHER.

1.4.2.4 Maintaining Carbon Dioxide Inside the Leaf Chamber

During photosynthesis carbon dioxide is removed from the environment and reduced to carbohydrate within the leaf. In a sealed or "closed" system such as the gas phase oxygen electrode, the concentration of carbon dioxide will gradually decline as photosynthesis proceeds. Eventually the availability of carbon dioxide will limit the rate of photosynthesis rather than any inherent physiology of the sample.

The time before such artefacts appear is determined by several factors, namely the rate of carbon dioxide consumption by photosynthesis and the quantity of carbon dioxide sealed in the reaction chamber at the beginning of the experiment. Under standard atmospheric conditions air contains approximately 0.035% carbon dioxide by volume. Hence the reaction chamber of the empty electrode unit contains approximately 245 µl of carbon dioxide (0.035 % x 1000 x the volume of the reaction chamber 7 ml). A typical healthy 10 cm2 C3 leaf disc can easily consume 50 µl carbon dioxide min-1 which could cause the leaf disc chamber to become depleted within 4 - 5 minutes (Walker 1987). It is also possible that desiccation of the sample over a period of time could cause leaf stomata to close, restricting entry of carbon dioxide to the interior of the leaf where assimilation occurs. An elevated ambient carbon dioxide concentration reduces the risk of such problems by increasing the total amount of carbon dioxide in the reaction chamber and the diffusion gradient for carbon dioxide entering the leaf. Normally carbon dioxide concentrations in the order of 1% are used to reduce the possibility of excessive carbon dioxide affecting other cellular processes (Walker 1987).

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Several techniques can be used to increase the carbon dioxide concentration of air in the reaction chamber to around 1%. The most common method is the addition of a small amount of bicarbonate buffer to the capillary matting the reaction chamber. If a bicarbonate solution left to equilibrate with an equal volume of air at around 25 °C , the solution will dissociate to [HCO3-] and [CO2] in a ratio of nearly 90 : 1. Using this information it is possible to calculate the quantity of bicarbonate solution which is required to yield a 1% carbon dioxide atmosphere in the volume of the reaction chamber. The calculation is as follows:

A 1% carbon dioxide atmosphere contains 10 ml carbon dioxide per litre of gas. Since 1 mole of an ideal gas occupies a volume of 24.45 L at 25°C, 10 ml of an ideal gas at 25°C will contain 1 / (24.45 x 1000 x 10) or 0.409 mmoles i.e. the concentration of carbon dioxide in a 1% atmosphere is 0.409 mM. As bicarbonate dissociates in ratio of 90 :1 a bicarbonate solution 90 times as concentrated will dissociate to yield a 1% carbon dioxide atmosphere. The concentration of such a solution will be 90 x 0.409 of 36.81 mM.

In the reaction chamber of the LD2/3 electrode the capillary matting is usually treated with 2 or 3 drops of bicarbonate solution (0.2 ml) which is left to equilibrate with a greater volume of air (7 ml). Hence the concentration of the bicarbonate solution must be scaled up by a suitable factor (7 / 0.2 or 35 times). As a result 2 or 3 drops of a 1M bicarbonate solution is usually sufficient to yield a 1% carbon dioxide atmosphere in the reaction chamber.

SODIUM OR POTASSIUM BICARBONATE IS FREQUENTLY USED. A MOLAR SOLUTION CONTAINS 84.01 GL-1 SODIUM BICARBONATE OR 100.12 GL-1 POTASSIUM BICARBONATE.

The carbon dioxide concentration can be controlled more precisely by mixing pure cylinder gases in defined concentrations. Premixed "special" gases can be purchased to order or gas from cylinders of pure oxygen, nitrogen and carbon dioxide can be mixed in defined ratios using mass flow control valves or Wöstoff pumps. The carbon dioxide concentration of such a blend can be verified using an infra red gas analyser (IRGA) and adjusted to precise requirements. The gas mixture can then be flushed through the LD2/3 chamber to displace atmospheric air by opening both taps.

If a number of experimental protocols are performed consecutively on the same sample or if experiments involving high rates of photosynthesis are carried out, it is possible that the carbon dioxide content of the 1% carbon dioxide atmosphere in the reaction chamber will be insufficient. The use of carbon dioxide concentrations in excess of 1% is not advisable due to the risk of artefacts induced by elevated carbon dioxide effects on other cellular processes. Hence it may be necessary to re-charge the chamber with carbon dioxide during a protocol. This is done by unsealing the reaction chamber by opening both taps and flowing air containing elevated carbon dioxide through the chamber to replace the carbon dioxide depleted air. Ideally this should be done with a known gas composition which has been warmed and humidified to the same extent as the chamber air. However if gas mixing apparatus is unavailable enhanced levels of carbon dioxide can be achieved by passing exhaled air through the chamber.

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ANY AIR WHICH IS FLOWED THROUGH THE CHAMBER SHOULD BE PRE-EQUILIBRATED TO THE TEMPERATURE OF THE CHAMBER. THIS IS DONE BY BREATHING EXHALED AIR INTO A SEALED CHAMBER STORED IN THE CIRCULATING WATER-BATH WHICH CONTROLS THE CHAMBER TEMPERATURE. AIR CAN BE TRANSFERRED AND FORCED THROUGH THE OPEN LD CHAMBER VIA SYRINGE. AFTER REMOVAL OF THE SYRINGE FROM THE TAP IT IS IMPORTANT TO LEAVE THE CHAMBER OPEN FOR A FEW SECONDS TO ENSURE THAT THE AIR PRESSURE INSIDE THE CHAMBER HAS NOT INCREASED.

In the presence of carbon dioxide, Clark-type oxygen electrodes normally develop bicarbonate in the electrolyte. This is because hydroxyl ions are generated at the cathode as oxygen is reduced and these react with carbon dioxide diffusing through the membrane. In some circumstances (e.g. if the carbon dioxide in the chamber is at a high partial pressure and if the oxygen consumption at the cathode is low, as it will be if no oxygen is being evolved within the chamber) the entry of carbon dioxide will exceed the formation of hydroxyl ions and the electrolyte will become acid. In consequence, the residual current will rise and, particularly if the electrode is being used at its maximum sensitivity, an appreciable artificial signal may be recorded. Under extreme conditions this can make the apparatus unusable because the spurious signal increases with time and, therefore, varies with the immediate past history of the electrode.

1.4.2.5 Temperature Control

Introduction to Temperature Control

All experimental assays will have an optimum temperature at which the sample will be most efficient in either evolving or consuming oxygen and it is important that this experimental temperature is maintained at all times throughout the duration of the experiment. This includes ensuring that any additions to be made to the reaction vessel during the experiment are pre-equilibrated to the assay temperature before being added.

According to the studies of G.A. Truesdale and A.L. Downing (The solubility of oxygen in water, 1954, Nature 173: 1236), at any given temperature and atmospheric pressure, air saturated, de-ionised water contains a known concentration of dissolved oxygen which may be calculated mathematically.

Oxygen concentration in a liquid decreases as temperature increases which is why it is important that the assay temperature is established for both the sample and any additions prior to the commencement of the experiment. However, in addition to this effect, the oxygen electrode disc itself is also sensitive to temperature.

The electrode disc signal increases with an increase in temperature. In assays where higher temperatures are used (such as mitochondrial respiration studies), it is recommended that the disc is fully equilibrated to the assay temperature before any calibration or measurement takes place. This can take up to 15 minutes.

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Therefore, it is not just liquid-phase measurement systems that are temperature critical; gas-phase systems are equally susceptible to fluctuations in temperature.

The assay temperature needs to be decided as one of the initial considerations of the experiment as the apparatus is calibrated based on a specified temperature.

With the exception of the Oxytherm Peltier chamber, all other liquid and gas-phase electrode chambers provide temperature control to the sample and disc via a water jacket which encapsulates the reaction / sample vessel. In liquid-phase systems, the vessel is constructed from borosilicate glass allowing an efficient heat transfer between the circulating water and the sample, thus maintaining effective temperature control.

In gas-phase electrode chambers, the sample is separated from the circulating water by a quartz window which also provides an effective thermal transfer.

The water jacket should be connected, using the plastic connectors provided, to the flow and return tubes from a thermoregulated circulating water bath.

In DW1, DW1/AD and DW3 electrode chambers, connect the flow tube to the lower water jacket connector and the return tube to the upper water jacket connector. This ensures that a fresh supply of water is constantly flowing around the DW1 water jacket and assists in dispersing any air bubbles which will rise naturally to the upper outlet port. In all other electrode chambers, the water jacket connectors are on the same plane so flow/return tubes may be connected to either connector.

The tubing lengths between the circulator and the electrode chamber should be kept minimal in order to maintain good temperature control. A circulator providing a flow rate of 6 litres/min or more is recommended.

Although the water jacket gives more positive control of sample temperature than many systems in which control is effected entirely by circulated air, it is, like them, entirely incapable of rapidly dissipating heat generated within the chamber by light containing a large infra-red component.

Appropriate filters (water, copper sulphate solution, interference filters, heat-reflecting mirrors etc) should therefore be employed when using a light source. Light saturation without heating is, of course, extremely difficult to achieve but the fact that the electrode is itself very responsive to small changes in temperature provides an easy method of checking the effectiveness of any light filtering system which is employed.

Halogen white light sources such as the Hansatech LS2 and the discontinued FLS1 light source produce a significant amount of heat. These light sources were fitted with an infra-red reducing hot-mirror filter in order to reduce the heating effect of the light source.

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All current Hansatech light sources contain LED emitters which produce significantly less heat output.

Temperature Effects on Liquid-Phase Samples

According to the studies of G.A. Truesdale and A.L. Downing (The solubility of oxygen in water, 1954, Nature 173: 1236), at any given temperature and atmospheric pressure, air saturated, de-ionised water contains a known concentration of dissolved oxygen which may be calculated mathematically.

This data is based on measurements of dissolved oxygen in water at the given temperature and standard atmospheric pressure published by Truesdale & Downing (Nature 173:1236, 1954).

Temperature (˚C)	Oxygen (PPM)	Oxygen (nmol/ml)
0	14.16	442.5
5	12.37	386.6
10	10.92	341.3
15	9.76	305
20	8.84	276.3
25	8.11	253.4
30	7.52	235
35	7.02	219.4

The formula used in calculating the oxygen values in the table is as follows:

Cs=14.16 - (0.394 * T) + (0.007714 * T^2) - (0.0000646 * T^3)

(Where Cs is the oxygen saturated concentration in ppm and T is temperature in °C)

1 ppm is equivalent to **1µg/ml or (1µg/32g/mol) = 0.03125 µmol/ml or 31.25 nmol/ml**

Temperature Effects on the Electrode Disc

The electrode disc signal increases with an increase in temperature. Therefore, the response from the disc opposes that of the oxygen signal at different temperatures. In assays where higher

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temperatures are used (such as mitochondrial respiration studies), it is recommended that the disc is fully equilibrated to the assay temperature before any calibration or measurement takes place. This can take up to 15 minutes and can be observed as follows:

- Prepare the electrode disc and mount correctly into the base of the electrode chamber. For liquid-phase chambers, add a 1ml, stirred sample of air-saturated, de-ionised water that is pre-equilibrated to the experimental temperature to the chamber.
- Connect the water jacket(s) of the electrode chamber to the supply from the circulating water bath (Refer to Setting Up Temperature Control section for more information).
- Begin a recording from the electrode disc and observe the oxygen signal. If the experimental temperature is high (e.g. 32°C), the signal from the electrode disc will initially rise due to the temperature sensitivity of the disc.
- Once the disc begins to equilibrate to the temperature, the signal will begin to decline as the higher temperature effect on the sample is opposite.
- Once the signal from the disc has re-stabilised, the disc is equilibrated to the assay temperature and calibration may be performed.

1.4.3 Care and Maintenance

1.4.3.1 LD2/3 Disassebly

The cast acrylic top window of the LD2/3 simply unscrews allowing the inside of the water jacket to be cleaned. Take care not to expose the Perspex (Plexiglas) top window to undiluted ethanol, acetone etc. Please note that the large O-ring which seals the leaf chamber to the quartz glass window of the top section should be checked for its condition from time to time and replaced if necessary. If it becomes perished it is possible that the leaf chamber will not seal correctly leading to signal drift.

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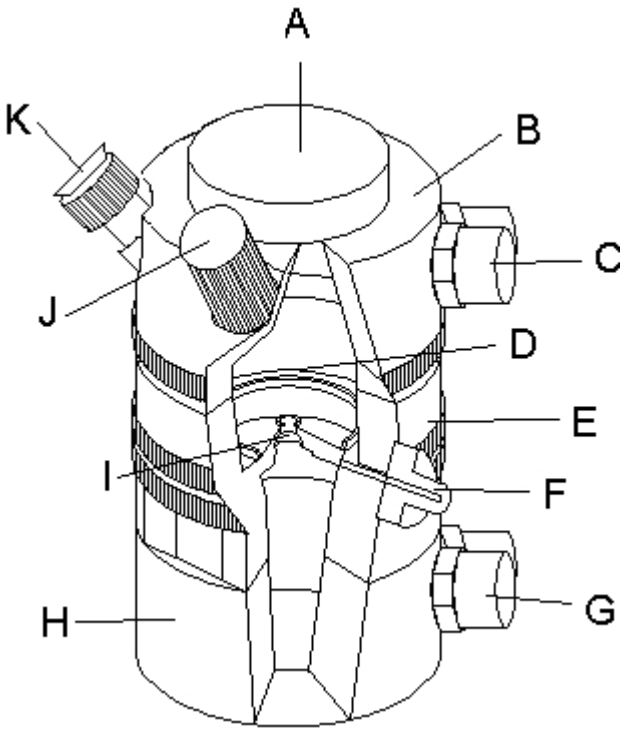
1.4.3.2 LD2/3 Spare Parts



- A: Top window
  Part No. 820051
- B: Upper water jacket
  Part No. 820184
- C: Water jacket connector socket & plug
  Part Nos. 890123 and 890124
- D: Inner window
  Part No. 885201
- E: Leaf chamber section
  Part No. 820062
- F: Gas tap port
  Part No. 820055
- G: Water jacket connector socket & plug
  Part Nos. 890123 and 890124
- H: Lower water jacket
  Part No. 820063
- I: Electrode disc
  Part No. S1
- J: Optical port
  Part No. 941007
- K: Fibre optic port
  Part No. 980261
- Clip and latch
  Part Nos. 728020 & 728021
- Spare O-rings
  Part No. S9B

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1.5 Oxylab + Electrode Control Unit

1.5.1 Introduction to Oxylab + Electrode Control Unit

The Oxylab+ oxygen electrode control unit is designed to provide PC control of oxygen uptake or evolution measurements across a broad range of applications from studies of mitochondrial and cellular respiration to measurements of isolated chloroplast suspensions or solid state leaf samples in photosynthesis research. In conjunction with user-friendly OxyTrace+ data acquisition and system configuration software, the Oxylab+ control unit provides an effective tool for the measurement of oxygen signals from the S1 Clark type oxygen electrode disc mounted in one of a range of liquid and gas-phase electrode chambers with quick and easy system calibration and configuration. Simultaneous recording of an optional auxiliary input signal (e.g. temperature, pH, chlorophyll fluorescence, TPP+ or other specific ion electrodes etc) is also possible using the appropriate apparatus.

A system comprises a maximum of 8 Oxylab+ control units linked to the USB port of a Windows® PC. The control unit features a built-in magnetic stirrer (for liquid-phase measurements) and all the electronics required to control and measure the signal from the oxygen electrode disc.

A maximum of 2 Oxylab+ systems can be used at any one time to perform quantum yield analysis.

Oxylab+ is compatible with all existing liquid and gas-phase Hansatech oxygen electrode chambers and accessories. In addition, Oxylab+ features light source control capabilities providing automated intensity changes of Hansatech Instruments LED light sources LH11/2R and LH36/2R. Additional tools within the OxyTrace+ software package allow editing of PFD (photon flux density) tables providing complete automation of complex light intensity changes during light response curves.

OxyTrace+ software controls all major hardware and data acquisition functions with simple calibration routines for both liquid and gas-phase measurements. Data from the electrode disc, optional auxiliary input signal and temperature signal from the QTP1 PAR/Temperature probe sensor are plotted as a chart recorder emulations in "real-time" with post-measurement data analysis tools included within the program. Completed experiments are saved in Comma Separated Values (*.csv) format which then may be opened directly in other Windows® data analysis applications such as Excel®.

1.5.2 Setting up Oxylab +

1.5.2.1 Connections to the Oxylab +

Before making any measurements, it is important that the system is set up correctly. The diagram below shows the various connections that must be made to the control units.

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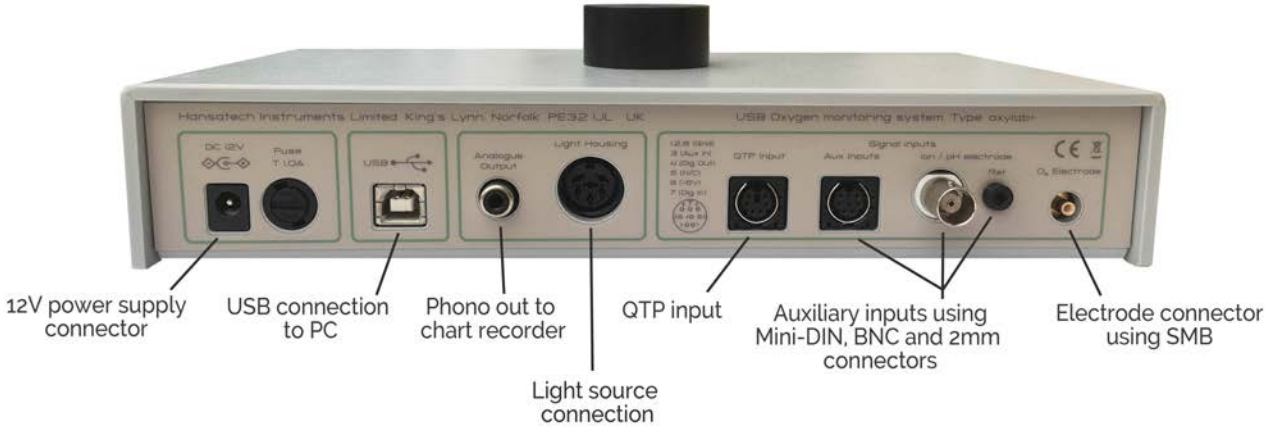

- Ensure that the 12V power supply is connected to the 12V input socket on the control unit. Confirmation that this connection has been made is shown by an illuminated green LED on the front panel of the control unit.
- Connect the USB cable to an available port on the PC. Connect the other end of the USB cable to the USB connection on the rear of the control unit. If more than one control unit is to be used, a second unit can be connected directly to the PC in the same way. More details of multi-channel systems can be found in the Multi-channel systems section..
- Connect the prepared electrode disc to the control unit via the electrode connection cable.
- Connect the light source to the light housing connection socket.
- If an auxiliary signal is to be recorded, users will require an OXY/AUX connection cable. This cable connects to the Electrode Disc Input on the rear of the Oxygraph and has a flying lead to which the prepared electrode disc and connection cable should be connected.
- If the QTP1 PAR/temperature probe is to be used, connect this to the QTP input socket.

1.5.3 OxyTrace + Software

1.5.3.1 Startup

OxyTrace + Software Initialisation

This section assumes that the OxyTrace+ software has been successfully installed on your PC and that all the necessary connections have been made between the electrode control unit and the PC with the control unit powered up.
Run the OxyTrace+ software by double clicking the desktop icon that was created during installation or by selecting OxyTrace+ from the Hansatech Instruments Ltd program group in the Windows Start Menu. If no control unit is located, the following dialogue box will be generated by the OxyTrace+ software.

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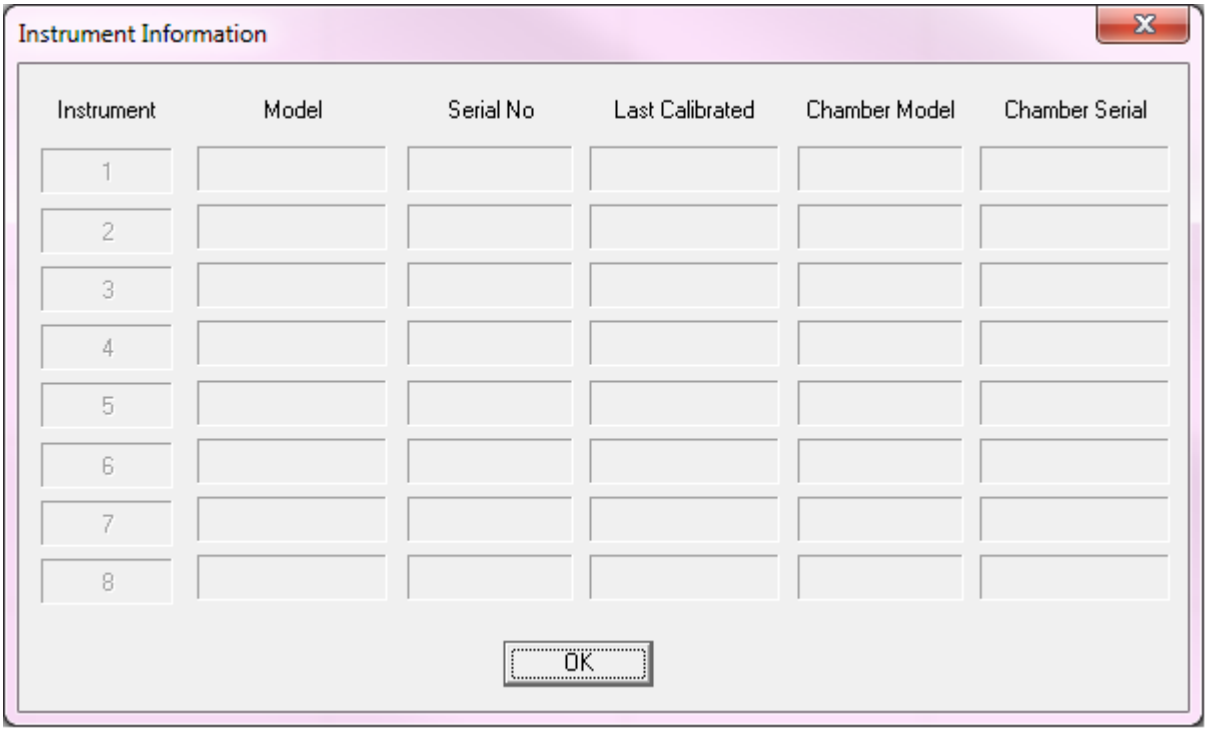

If you see this dialogue box displayed with empty values for Model, Serial No etc, please ensure that you have correctly connected the control unit to the PC and that it is powered up with a blue status indication light on the front panel of the control unit. When this been confirmed, press OK to close this dialogue and then select Hardware > Scan for Instruments. You will then see the following dialogue compete with information regarding the connected control unit such as serial number and calibration status. Press OK to continue.

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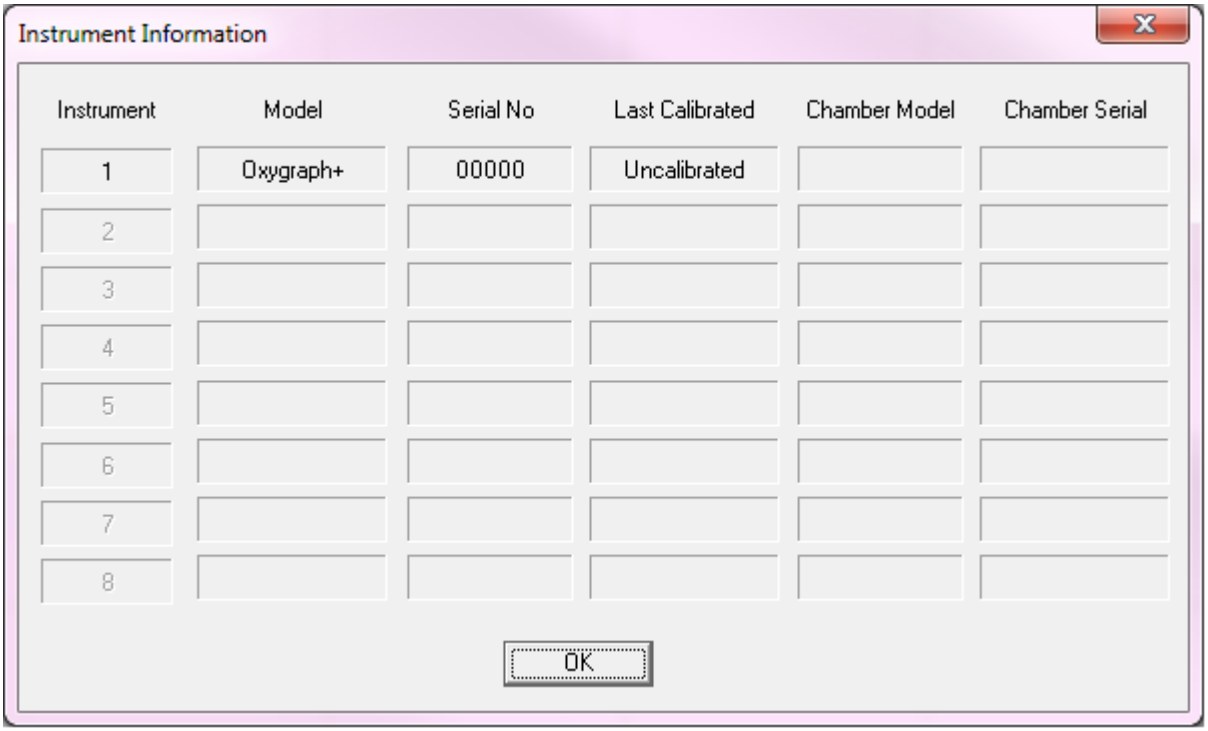

If communications with the control unit cannot be established, please refer to the Communication Problems between Control Unit and the PC section for further information

Testing the Electrode Disc Response

Once the electrode disc has been correctly prepared, it is advisable to check the response of the disc prior to mounting the disc within the electrode chamber.

Before proceeding, you will need to ensure that the electrode control unit you are using is correctly connected to the PC and that it is communicating with the OxyTrace+ software. If you are experiencing problems with communications between the control unit and software, please refer to the Software Initialisation section of this document.

To check the response, connect the electrode disc to the rear of the control unit at the electrode input using the S1/ADL electrode connection cable. Start a recording. The signal will initially be extremely high and displayed as nanoamps (nA) and will take a few minutes to stabilise at an air line level. A new or well maintained disc should read approx. 1600 nA in air but this may vary between ±240nA.

Once the electrode signal is stable, breathe exhaled air across the disc and observe the reaction on the graph screen. The signal should be plotted as in the diagram below.

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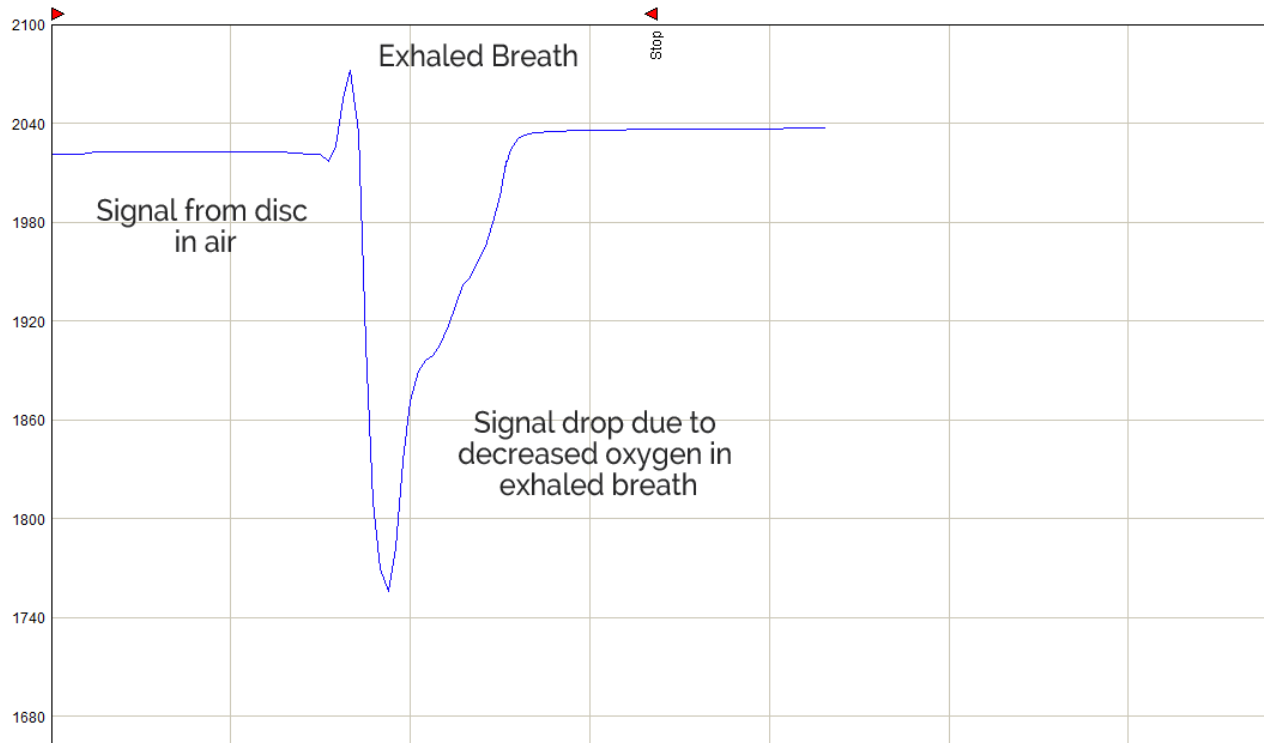

The first deviation in signal after breathing across the disc is induced due to a large increase in temperature of exhaled breath compared to ambient air temperature. Oxygen electrode discs are particularly sensitive to temperature and will show an increase in signal as a result. Since the temperature increase is only temporary, the oxygen signal will fall after a short time.

After the temperature related signal increase, a steep drop in signal is observed due to decreased oxygen levels in exhaled breath (approx 17%). The signal should the begin to return to the original level as the ambient oxygen begins to equilibrate around the electrode disc.

If the observed signal is not as shown above, it may be caused by an inadequate electrode preparation or worst case, a problem with the disc itself. Try cleaning the disc and re-preparing before repeating the test described above. If problems still occur, please contact Hansatech Instruments.

Once this test has been completed satisfactorily, mount the disc into the base of the electrode chamber. An additional test may be carried out in order to test the response of the disc in situ. Please refer to the Operating the Magnetic Stirrer section for further details.

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1.5.3.2 OxyTrace + Workspace

Menu Bar

File Menu

- New

Creates a new document

- Open

Opens an existing file

- Overlay

Overlay's an existing file on to the same axes as the file currently loaded

- Save

Saves the current file

- Save As

Specify a new filename for the current file

- Print

Prints the current file. The printout will reflect whichever method of data display is currently active. For example, if the tabulated data display mode is selected, this is what will be printed

- Print Preview

Shows a preview of how the current file view will be printed

- Print Setup

Opens the Windows printer settings dialogue

- Recently Opened Files

Displays a list of the most recently opened files

- Exit

Closes the program

Hardware Menu

When an Oxygraph+ control unit is connected to OxyTrace+ , the Hardware menu is presented as follows:

- Start Recording
  Begins recording the oxygen signal from the S1 electrode and any auxiliary input that is configured.
- Stop Recording
  Stops the current recording.
- Channel Configuration
  Opens the Instrument Summary dialogue
- Acquisition Rate
  Opens the Data Acquisition Rate dialogue.
- Stirrer Speed
  Opens the Sample Stirring Dialogue

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- Setup NEW PFD Table
  Opens the PFD (Photon Flux Density) table to allow experimental light intensity steps to be defined and executed. Please refer to the Configuration of PFD table section for further information
- Scan for Instruments
  This tool is prompts OxyTrace+ to scan the PC com ports for a connected Oxygraph control unit. Use this tool when adding additional Oxygraph control units to a multi-channel chain or when requiring a connected control unit after having selected View Files at the Software Initialisation dialogue.
- Instrument Test
  Opens the Instrument Test dialogue for control unit diagnostic features.

When an Oxytherm+ control unit is connected to OxyTrace+ , the Hardware menu is presented as follows:

- Start Recording
  Begins recording the oxygen signal from the S1/MINI electrode and any auxiliary input that is configured.
- Stop Recording
  Stops the current recording.
- Channel Configuration
  Opens the Instrument Summary dialogue
- Acquisition Rate
  Opens the Instrument Summary dialogue allowing the rate that the control unit measures oxygen from the electrode disc to be configured.
- Stirrer Speed
  Opens the Sample Stirring Dialogue.
- Temperature Control
  Opens the Peltier Chamber Temperature dialogue to set Oxytherm temperature control.
- Temperature Record
  Logs the measured temperature of the Peltier Chamber. Temperature values are presented in the Tabulated Data view.
- Actinic Light Control
  Allows the configuration of the actinic light within the Oxytherm+
- Chamber Viewing Light
  Presents to option to enable or disable the Oxytherm+ viewing light
- Setup NEW PFD Table
  Opens the PFD (Photon Flux Density) table to allow experimental light intensity steps to be defined and executed. Please refer to the Configuration of PFD table section for further information

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- Scan for Instruments
  This tool is prompts OxyTrace+ to scan the PC com ports for a connected Oxytherm+ control unit. Use this tool when adding additional Oxytherm+ control units to a multi-channel chain.
- Instrument Test
  Opens the Instrument Test dialogue for control unit diagnostic features.

When an Oxylab+ control unit is connected to OxyTrace+ , the Hardware menu is presented as follows:

- Start Recording
  Begins recording the oxygen signal from the S1 electrode and any auxiliary input that is configured.
- Stop Recording
  Stops the current recording.
- Channel Configuration
  Opens the Instrument Summary dialogue.
- Acquisition Rate
  Opens the Instrument Summary dialogue.
- Stirrer Speed
  Opens the Sample Stirring Dialogue.
- Temperature Record
  Logs the measured temperature of the Peltier Chamber. Temperature values are presented in the Tabulated Data view.
- Light Control
  Opens the Light Source Control dialogue which allows the attached light source to be set to a given intensity. Please refer to the Manual Control of Light Source section for further information.
- Setup NEW PFD Table
  Opens the PFD (Photon Flux Density) table to allow experimental light intensity steps to be defined and executed. Please refer to the Configuration of PFD table section for further information.
- Scan for Instruments
  This tool is prompts OxyTrace+ to scan the PC com ports for a connected Oxylab control unit. Use this tool when adding an additional Oxylab control unit to a dual-channel setup or when requiring a connected control unit after having selected View Files at the Software Initialisation dialogue.
- Instrument Test
  Opens the Instrument Test dialogue for control unit diagnostic features.

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Calibrate Menu

Liquid Phase Calibration:

- Air Saturated Water

  Begins a standard liquid phase calibration routine using air saturated water as the known constant.
- Manual

  In assays which do not use air saturated water, use this calibration option. You will be prompted to enter a known oxygen value to reference against the airline stage of the calibration routine. Please refer to the Manual Calibration Routine for further information.

- Gas Phase Calibration:

When using a gas-phase electrode chamber, use this option to initiate the gas-phase calibration routine. Please refer to the Gas-Phase Calibration section for further information

- Oxygen Calibration Details

Views the details of any current oxygen calibration. Please refer to the Viewing Oxygen Calibration Details section for further information

- Set Oxygen Warning Interval

An oxygen calibration warning interval may be set so that after a given number of days post calibration, the user is prompted that the calibration has expired when the software is initially run. Please refer to the Set Oxygen Calibration Warning section for further information.

- Calibrate Light - Automatic

When using an Oxylab+ control unit, this begins the automatic calibration routine for connected light source. Please refer to the Light Source Calibration section for further information.

- Calibrate Light - Manual

This allows for a light source to be manually calibrated. Please refer to the Light Source Calibration section for further information.

- Light Calibration Details

Views the details of any current light calibration. Please refer to the Light Source Calibration section for further information.

- Set Light Warning Level

An light calibration warning interval may be set so that after a given number of days post calibration, the user is prompted that the calibration has expired when the software is initially run. Please refer to the Light Source Calibration section for further information.

- Calibrate Auxiliary/pH ISE/Other ISE

Displays the auxiliary channel details and Initialises the Auxiliary Input Calibration routine. Please refer to the Auxiliary Device Calibration section for further information.

- Aux Calibration Details

Displays the calibration details of connected auxiliary devices. Please refer to the Auxiliary Device Calibration Details section for further information

- Set Auxiliary warning interval

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An auxiliary calibration warning interval may be set so that after a given number of days post calibration, the user is prompted that the calibration has expired when the software is initially run. Please refer to the Set Auxiliary Calibration Warning section for further information.

View Menu

- File Information

Displays general information that saved along with the current file. Please refer to the File Information section for further information

- Toolbar

Toggles the Toolbar between show/hide

- Status Bar

Toggles the Status Bar at the foot of the screen between show/hide

Graph Menu

- Setup Trace Colours

Allows trace colour and data point marker colours to be customised. Please refer to the Trace Settings section for further information

- Display Overlaid Traces

Toggles any overlaid traces between show/hide

- Zoom Window

Graph zoom control allowing a selected portion of the trace to be enlarged. Clicking this icon changes the mouse cursor to cross hairs. Clicking and drag the window to highlight the required section and click the mouse again to apply the zoom. Please refer to the Axes Settings section for further information

- Zoom Auto

Graph zoom control which automatically re-sizes the axes to efficiently present all recorded oxygen and any auxiliary data. Please refer to the Axes Settings section for further information

- Zoom XY

Graph zoom control. Selecting this option opens the Set Axes window allowing oxygen and auxiliary axes to be scaled by entering in the required from and to values for oxygen, time and optional auxiliary axes. Please refer to the Axes Settings section for further information

- Zoom Plus

Graph zoom control which increases the zoom based at the center of the graph. Please refer to the Axes Settings section for further information

- Zoom Minus

Graph zoom control which decreases the zoom based at the center of the graph. Please refer to the Axes Settings section for further information

- Zoom Undo

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Graph zoom control used to undo the previous zoom command. Please refer to the Axes Settings section for further information.

Data Bar Menu

When an Oxygraph+ control unit is connected to OxyTrace+ , the Hardware menu is presented as follows:

- Output

Toggles output level in the data bar between show/hide

- Rate Data

Toggles Live Rate data in the data bar between show/hide.

- Aux Data

Toggles auxiliary device data in the data bar between show/hide

- Small Font

Toggles the font used in the data bar between small and large size

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When an Oxytherm+ control unit is connected to OxyTrace+ , the Hardware menu is presented as follows:

- Output

Toggles output level in the data bar between show/hide

- Rate Data

Toggles Live Rate data in the data bar between show/hide.

- Aux Data

Toggles auxiliary device data in the data bar between show/hide

- Set Temperature

Toggles the set chamber temperature display in the data bar between show/hide

- Actual Temperature

Toggles actual chamber temperature display in the data bar between show/hide

- Small Font

Toggles the font used in the data bar between small and large size

- Flurescence Data

Toggles fluorescence level in the data bar between show/hide

When an Oxylab+ control unit is connected to OxyTrace+ , the Hardware menu is presented as follows:

- O2 signal mV

Toggles output level in the data bar between show/hide

- Rate Data

Toggles Live Rate data in the data bar between show/hide.

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- Aux Data

Toggles auxiliary device data in the data bar between show/hide

- Light Setting

Toggles the current set light source intensity display in the data bar between show/hide

- Actual PAR Light

Toggles actual current light source intensity reading from the QTP1 sensor in the data bar between show/hide

- Actual Temperature

Toggles actual chamber temperature reading from the QTP1 sensor in the data bar between show/hide

- Small Font

Toggles the font used in the data bar between small and large size

Tools Menu

When an Oxygraph+ control unit is connected to OxyTrace+ , the Hardware menu is presented as follows:

- PFD Setup & Go

Opens the PFD (Photon Flux Density) table to allow experimental light intensity steps to be defined and executed. Please refer to the Configuration of PFD table section for further information

- PFD Go

Executes the currently defined PFD table. Please refer to the Configuration of PFD table section for further information

- QY Graph Setup

Displays the Quantum Yield graph screen configuration options. Please refer to the Quantum Yield Graph Setup section for further information

- Oxygen Graph

Displays the graph screen in the data display area. Please refer to the Graph Screen section for further information

- Oxygen+ Rate Graph

Displays a split screen with the oxygen graph at the lower section and the Live Rate data in the upper section. Please refer to the Plotting Live Rate Data section for further information.

- Tabulated Data

Displays the tabulated data screen in the data display area. Please refer to the Tabulated Data section for further information

- Add Event Mark

Adds an event marker to the current recording. Please refer to the Adding Event Marks section for further information

- Edit Event Marks

Allows event marks on the current file to be edited via the Edit Event Marks dialogue. Please refer to the Editing Event Marks section for further information

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- Delete Event Marks

Allows event marks on the current file to be deleted via the Delete Event Marks dialogue. Please refer to the Deleting Event Marks section for further information

- Options

Opens the Options dialogue allowing general program options to be configured. Please refer to the Program Options section for further information

---

When an Oxytherm+ control unit is connected to OxyTrace+ , the Hardware menu is presented as follows:

- PFD Setup & Go

Opens the PFD (Photon Flux Density) table to allow experimental light intensity steps to be defined and executed. Please refer to the Configuration of PFD table section for further information

- PFD Go

Executes the currently defined PFD table. Please refer to the Configuration of PFD table section for further information

- QY Graph Setup

Displays the Quantum Yield graph screen configuration options. Please refer to the Quantum Yield Graph Setup section for further information

- Oxygen Graph

Displays the graph screen in the data display area. Please refer to the Graph Screen section for further information

- Oxygen+ Rate Graph

Displays a split screen with the oxygen graph at the lower section and the Live Rate data in the upper section. Please refer to the Plotting Live Rate Data section for further information.

- Tabulated Data

Displays the tabulated data screen in the data display area. Please refer to the Tabulated Data section for further information

- Add Event Mark

Adds an event marker to the current recording. Please refer to the Adding Event Marks section for further information

- Edit Event Marks

Allows event marks on the current file to be edited via the Edit Event Marks dialogue. Please refer to the Editing Event Marks section for further information

- Delete Event Marks

Allows event marks on the current file to be deleted via the Delete Event Marks dialogue. Please refer to the Deleting Event Marks section for further information

- Options

Opens the Options dialogue allowing general program options to be configured. Please refer to the Program Options section for further information

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When an Oxylab+control unit is connected to OxyTrace+ , the Hardware menu is presented as follows:

- PFD Setup & Go

Opens the PFD (Photon Flux Density) table to allow experimental light intensity steps to be defined and executed. Please refer to the Configuration of PFD table section for further information

- PFD Go

Executes the currently defined PFD table. Please refer to the Configuration of PFD table table section for further information

- QY Graph Setup

Displays the Quantum Yield graph screen configuration options. Please refer to the Quantum Yield Graph Setup section for further information

- Oxygen Graph

Displays the graph screen in the data display area. Please refer to the Graph Screen section for further information

- Oxygen+ Rate Graph

Displays a split screen with the oxygen graph at the lower section and the Live Rate data in the upper section. Please refer to the Plotting Live Rate Data section for further information.

- Tabulated Data

Displays the tabulated data screen in the data display area. Please refer to the Tabulated Data section for further information

- PFD Data

Displays the currently defined PFD table in the data display area. Please refer to the Configuration of PFD table section for further information.

- QY Data

Displays the tabulated data screen in the data display area. Please refer to the Tabulated Data section for further information

- QY+ Graph Data

Displays a split screen with tabulated data screen plus Graph Data. Please refer to the Tabulated Data section for further information

- Add Event Mark

Adds an event marker to the current recording. Please refer to the Adding Event Marks section for further information

- Edit Event Marks

Allows event marks on the current file to be edited via the Edit Event Marks dialogue. Please refer to the Editing Event Marks section for further information

- Delete Event Marks

Allows event marks on the current file to be deleted via the Delete Event Marks dialogue. Please refer to the Deleting Event Marks section for further information

- Options

Opens the Options dialogue allowing general program options to be configured. Please refer to the Program Options section for further information

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Rate menu

- Setup Live Rate Display

Opens the Setup Live Rate Display dialogue allowing a time interval over which the Live Rate is calculated in real-time during a measurement. Please refer to the Setup Live Rate Display section for further information.

- Rate Cursors

Toggles the rate cursors between show/hide. When displayed, 2 rate cursors are presented as vertical lines on the oxygen axes representing the beginning and end of the rate interval. Please refer to the Manual Rate Measurements section for further information

- Enter Rate Cursor Times

Allows the start and end points of the rate interval to be defined as an actual time point during the current file. Please refer to the Manual Rate Measurements section for further information.

- Display Rate Table

Opens the Rates of Change table. This window displays all rates that have been calculated to date for the current file. Please refer to the Manual Rate Measurements section for further information

- Add Rate to Table

When the rate interval has been defined, use this tool to add the rate calculation to the Rates of Change table. Please refer to the Manual Rate Measurements section for further information.

- Select Channels

Opens a dialogue which controls which channel signals the rate cursors will take into account when calculating a rate of change. Options are any of the channels individually or all channels together. If only 1 channel is present, this selection is obsolete. Please refer to theManual Rate Measurements section for further information.

- Setup 'Line of Best Fit'

When enabled, the Line of Best Fit automatically draws the best fit slope against the oxygen signal between the rate interval cursors. This option maybe preferentially enabled to act on individual channel signals or across all connected channel signals. If the Line of Best Fit is disabled, the calculation of the rate still happens but the line is not drawn on the graph. Please refer to the Manual Rate Measurements section for further information.

Help Menu

- Help

Opens this help document

- About OxyTrace+

Displays version and copyright information.

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OxyTrace + Toolbar

The following tables show the different icons found on the OxyTrace+ toolbar.

Opens an existing file

Saves the current file.

Prints the current file. The printout will reflect whichever method of data display is currently active. For example, if the tabulated data display mode is selected, this is what will be printed

Begins recording the oxygen signal from the electrode disc and any auxiliary input that is configured

Stops the current recording

Opens the Instrument Summary dialogue allowing Oxygen signal to be seen in both nA and mV as well as displaying legacy units

Opens the Data Acquisition Rate dialogue allowing the rate that the control unit measures oxygen from the electrode disc to be configured

Magnetic stirrer control. This button has dual functionality depending on options that are set within the Stirrer Speeds dialogue. If the option "Enable Individual Controls" IS NOT checked in the stirrer speeds dialogue, this button will simply switch the stirrers or any connected electrode control units on and off. If the "Enable Individual Controls" options IS checked, this button will open the Stirrer Speeds dialogue to allow individual control of each control unit stirrer

This tool is prompts OxyTrace+ to scan the USB ports for a connected control unit. Use this tool when adding additional control units to a multi-channel chain

Begins a standard liquid phase calibration routine using air saturated water as the known constant

When using a gas-phase electrode chamber, use this option to initiate the gas-phase calibration routine. Please refer to the Gas-Phase Calibration section for further information

Displays general information that saved along with the current file. Please refer to the File Information section for further information

Display the data from the overlaid file as well as the normal file

Toggles the rate cursors between show/hide. When displayed, 2 rate cursors are presented as vertical lines on the oxygen axes representing the beginning and end of the rate interval. Please refer to the Manual Rate Measurements section for further information

Graph zoom window control allowing a selected portion of the trace to be enlarged. Clicking this icon changes the mouse cursor to cross hairs. Clicking and drag the window to highlight the required section and click the mouse again to apply the zoom. Please refer to the Axes Settings section for further information

Graph zoom control which automatically re-sizes the axes to efficiently present all recorded oxygen and any auxiliary data. Please refer to the Axes Settings section for further

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information

Graph zoom control. Selecting this option opens the Set Axes window allowing oxygen and auxiliary axes to be scaled by entering in the required from and to values for oxygen, time and optional auxiliary axes. Please refer to the Axes Settings section for further information

Graph zoom control which increases the zoom based at the center of the graph. Please refer to the Axes Settings section for further information

Graph zoom control which decreases the zoom based at the center of the graph. Please refer to the Axes Settings section for further information

Graph zoom control used to undo the previous zoom command. Please refer to the Axes Settings section for further information

Displays the graph screen in the data display area. Please refer to the Graph Screen section for further information

Displays a split screen with the oxygen graph at the lower section and the Live Rate data in the upper section. Please refer to the Plotting Live Rate Data section for further information

Displays the tabulated data screen in the data display area. Please refer to the tabulated data section for further information

Adds an event marker to the current recording. Please refer to the Adding Event Marks section for further information

Displays version and copyright information

Oxytherm+ Specific Icons (additional to the common icons)

Opens the Peltier Chamber Temperature dialogue to set Oxytherm+ temperature control

Oxylab+ Specific Icons (additional to the common icons)

Opens the Light Source Control dialogue which allows the attached light source to be set to a given intensity. Please refer to the Manual Control of Light Source section for further information.

Opens the Setup New PFD table dialogue. This function allows complex light regimes to be configured for execution within an experiment. Please refer to the Configuration of PFD Table section for further information.

Executes the currently defined PFD table. Please refer to the Configuration of PFD Table table section for further information.

Display the existing photon flux density table

Display the quantum yield data.

Display both the quantum yield data and oxygen trace

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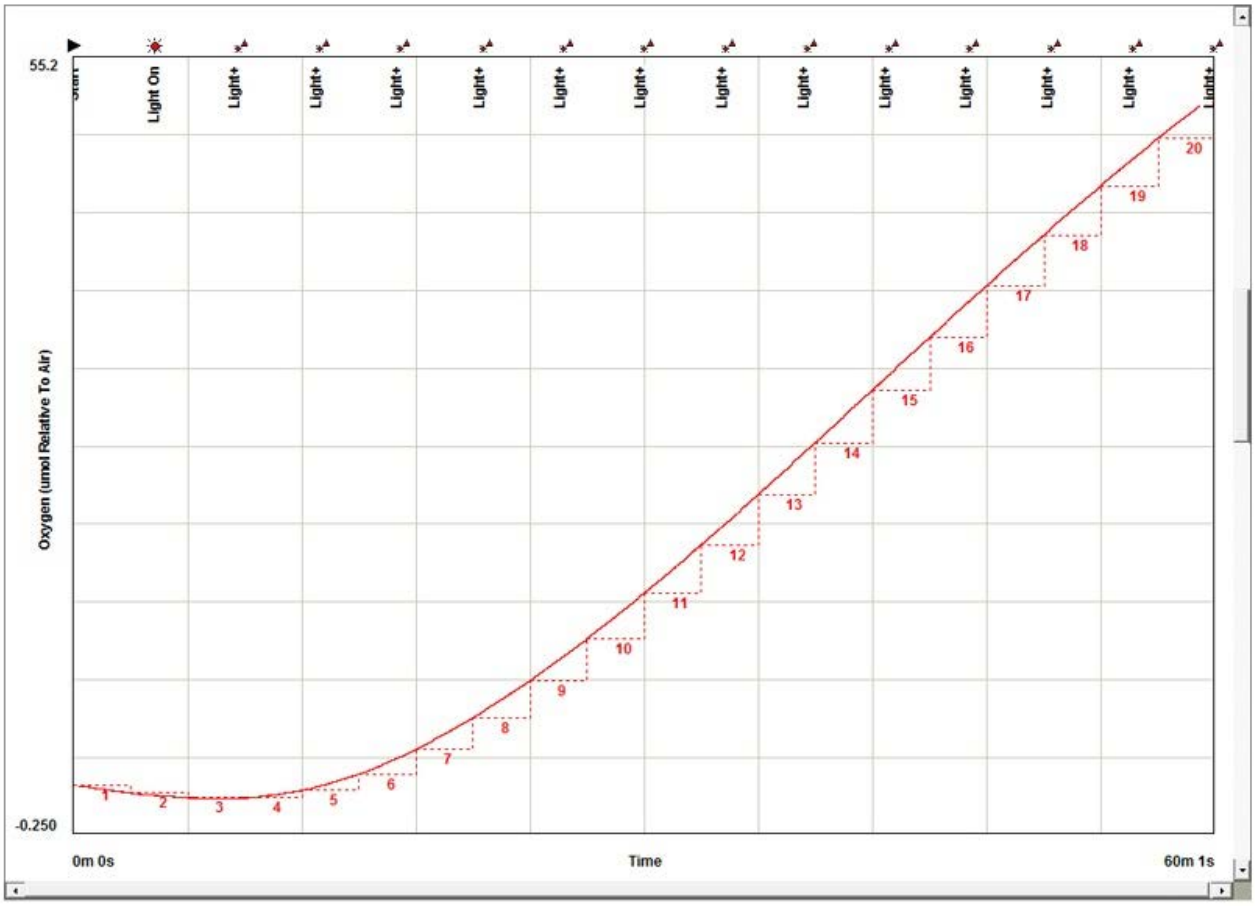
Graph Area

Introduction to the Graph Area

The graph area of OxyTrace+ allows the signal from the electrode disc (and optional auxiliary input) to be plotted on the PC screen as a real-time chart recorder emulation. Various controls and functions exist to facilitate the way in which part or all of an experiment is displayed on screen.



The recorded signal is plotted to the graph at the predefined data acquisition rate. Once the plotted traces reach to right side of the graph, the screen begins to scroll. The graph may be scrolled in 3 different methods:

- Smooth Scrolling - the graph scrolls in the same fashion as a chart recorder.
- Half Screen - the graph shifts by half a screen.
- Full Screen - the graph shifts by a full screen.

These options may be selected from the Options menu.

During the measurement, OxyTrace+ can display up to 5 minutes worth of recorded data on screen at any one time. Data recorded that is not in view may be easily reviewed by moving the horizontal scroll bar at the bottom of the graph. This does not affect normal measurement.

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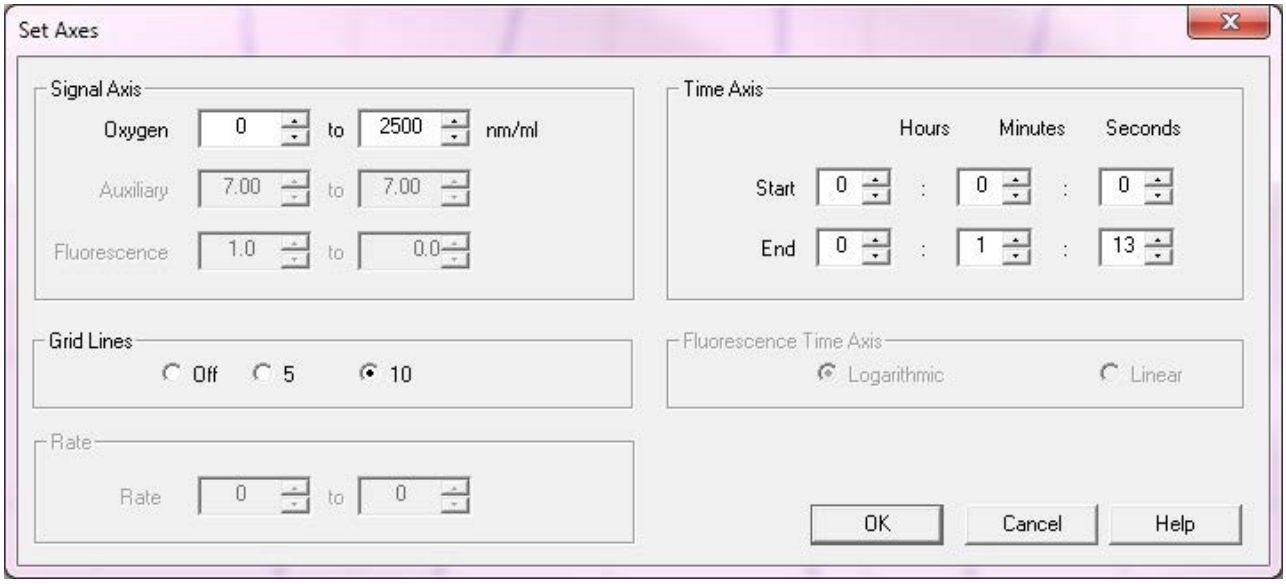
Axes Settings

OxyTrace+ features a comprehensive range of zoom controls in order that the recorded data may be presented most efficiently on screen. They can be accessed either by selecting one of the options in the Graph menu bar option or by clicking one of the following icons:

Zoom Window - Clicking this icon allows a selected portion of the trace to be enlarged. Clicking this icon changes the mouse cursor to cross hairs. Clicking on the graph activates the selection window. Drag the window to highlight the required section and click the mouse again to zoom.

Auto Zoom - Clicking this icon automatically re-sizes the axes to efficiently present all recorded data. This function may also be performed by pressing the F3 function key on the keyboard.

Zoom XY - Clicking this icon opens the manual Set Axes window as shown below.



Axes are adjusted by entering in the required from and to values for oxygen, time and optional auxiliary/fluorescence axes. If the auxiliary or fluorescence signals are disabled, the settings for the associated axes are inaccessible. The grid lines that appear by default in the graph area can also be modified in this dialogue.

Increase Zoom - Clicking this function zooms in on the centre section of the screen

Decrease Zoom - Clicking this function zooms out of the screen

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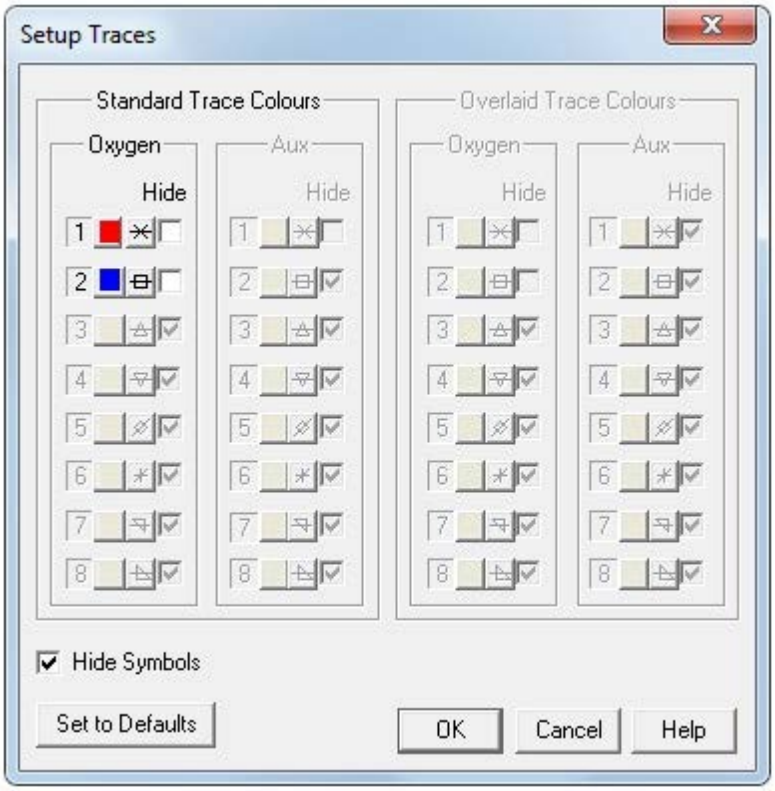
Undo Zoom - Click this icon to undo the previous zoom command. This function may also be performed by pressing the **F4** key on the keyboard.

Trace Settings

OxyTrace+ allows all individual signal traces to be assigned colours and data point symbols for ease of identification. This is particularly important when using Oxygraph+ and Oxytherm+ multi-channel systems at a full capacity of 8 individual channels with auxiliary channels enabled.



Clicking on the colour block adjacent to the channel numbers opens the standard Operating System colour picker dialogue allowing a colour to be selected by assigning RGB values or by selecting a predefined colour from the default Operating System colour palette. Clicking on the symbol adjacent to the channel number opens a dialogue allowing the individual data point marker to be selected from a set of 10 different options. The default selection is to hide these data point markers. Unchecking the Hide Symbols check box at the bottom of the dialogue will display all individual data point markers in the graph area.

The "Hide" check box column allows individual channel traces to be toggled in and out of view in the graph area.

Colours and symbols of additional systems may also be defined, these settings are enabled once OxyTrace+ detects multiple channels.

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Data Bar

The OxyTrace+ data bar presents a digital read out of the signals from the control unit. Data from multi-channel systems will be displayed on individual lines. The information displayed is as follows:

- Trace colours - Both oxygen and auxiliary trace colours are represented in the data bar so that information can easily be matched to traces on screen in multi-channel systems.
- Oxygen electrode signal - Presented in nA when uncalibrated and nmols/ml (liquid-phase) or µmol/RTA (gas-phase) when calibrated.
- Calibration indicator - Indicates whether the channel is:

  ✓ = Calibrated

  ✓ = Calibration has expired (see Calibration Details for information on Warning Intervals)

  ! = Uncalibrated

- Rate Data - Displays the Live Rate according to the options selected (please refer to the Rate section for further details)
- Output - Analogue output signal presented in mV
- Aux. Data - Displays the signal from an auxiliary input in arbitrary units if enabled (please see Auxiliary Signals for further details)
- Set and actual chamber temperature - Oxytherm+ Only

Clicking the right mouse button on the data bar brings up an additional dialogue of options as shown in the diagram above. This dialogue allows any of the columns (except for the oxygen signal which is always visible) to be hidden or shown simply by clicking the appropriate column heading. The default setting shows the live rate along with the oxygen signal, trace colour and calibration status which are displayed as standard and are not selectable.

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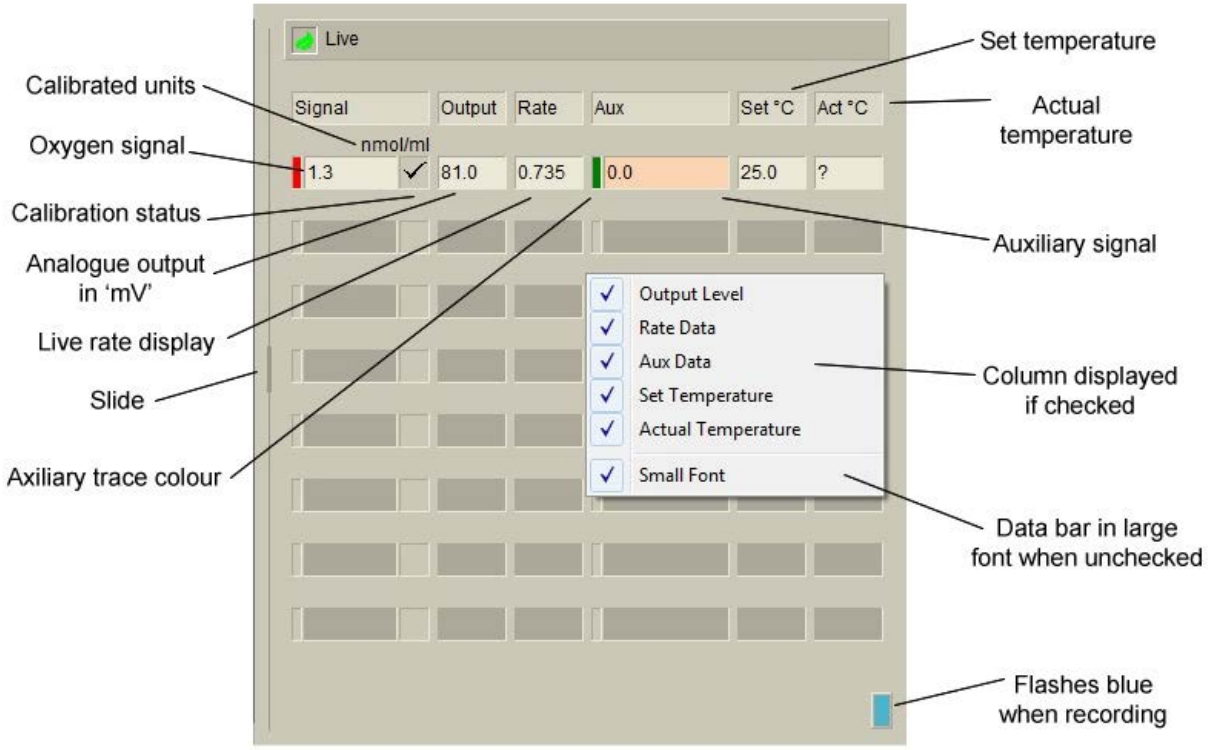

PLEASE NOTE THAT THESE OPTIONS WILL BE DIFFERENT ACCORDING TO THE CONTROL UNIT IN USE. THIS DIAGRAM SHOWS THE OPTIONS AVAILABLE FOR THE OXYTHERM+ CONTROL UNIT. IN OXYGRAPH+ SYSTEMS, THE TEMPERATURE OPTIONS WILL BE HIDDEN. IN OXYLAB+ BASED SYSTEMS, THE SET TEMPERATURE OPTION IS REMOVED AND FURTHER OPTIONS DISPLAYED ARE LIGHT SETTING AND ACTUAL PAR READING IN ADDITION TO THE ACTUAL TEMPERATURE OPTION. THESE SETTINGS REFLECT THE DIFFERENT OPTIONS AVAILABLE IN THE DATA BAR MENU.

The data bar may also be adjusted to show all enabled columns, just oxygen signal or no columns by using the slide feature. By clicking and holding down on the slide you may drag the data bar to the right to adjust between which columns will be shown.
Other features of the data bar are selectable font size (either small or large fonts) and a flashing recording indicator in the bottom left corner of the data bar.
All of the controls for the Data Bar features may be accessed from the Data Bar menu.

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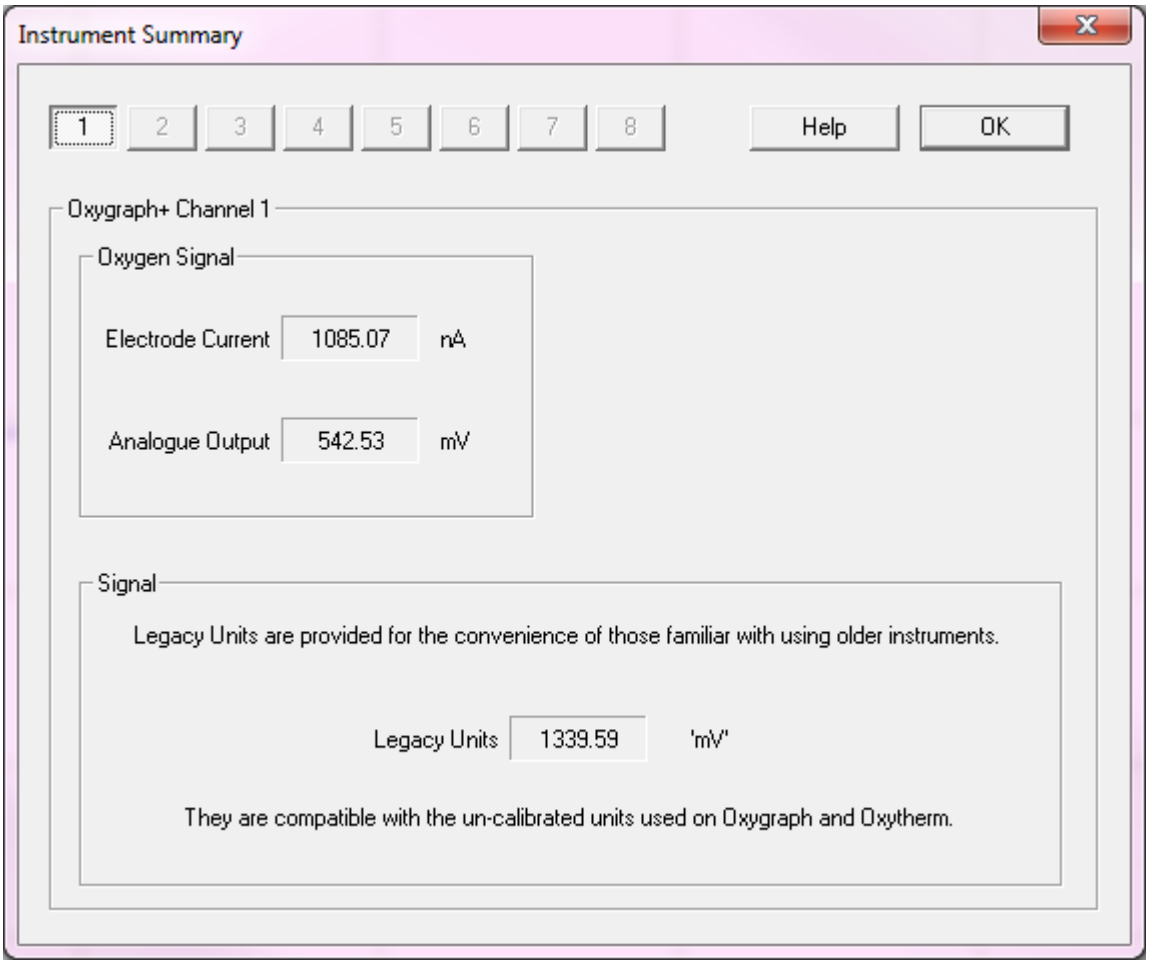
1.5.3.3 System Setup and Configuration

Instrument Summary

The Instrument Configuration window can be accessed either by selecting Hardware > Instrument Summary from the menu bar, by clicking the icon on the tool bar or by pressing the **F8** function key on the keyboard.



Channel selection

The buttons at the top of the window correspond to the possible 8 channels of a multi-channel Oxygraph+ /Oxytherm+ system or possible 2 in Oxylab+ based systems. Channels which are actively connected to the system are clickable whereas inactive channels are greyed out.

Oxygen Signal : Digital display of electrode signal

These meters show the readout from the electrode disc as a digital value. The calibrated units are displayed in either nmol/ml for liquid-phase or µmol RTA for gas-phase calibrations, when uncalibrated the units displayed are nA.

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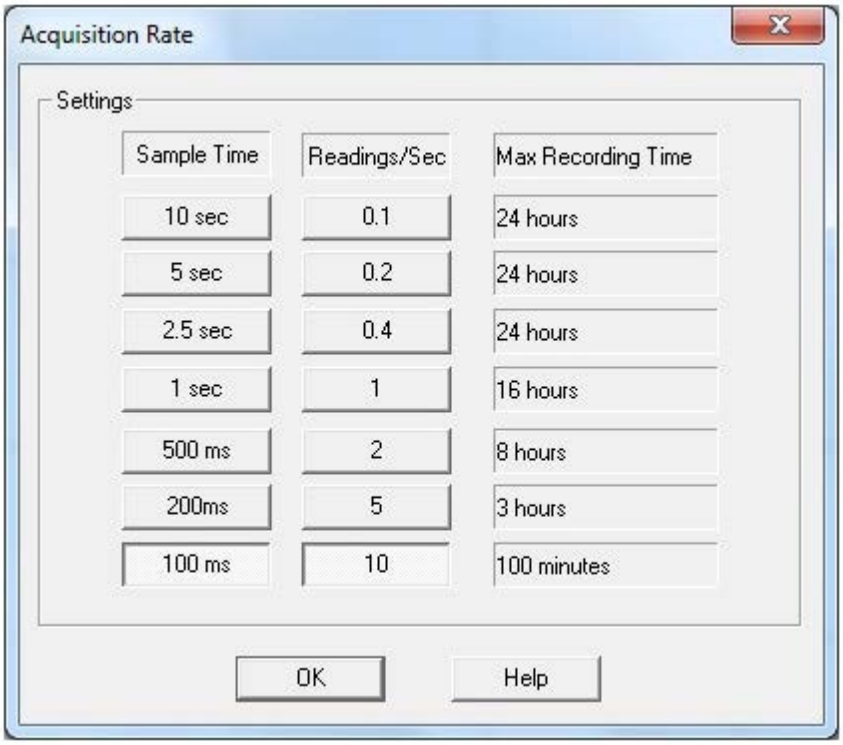
Signal: Legacy Units

Legacy Units are provided for the convenience of those familiar with using older instrments. They are compatible with the un-calibrated units used on Oxygraph or Oxytherm.

Data Aquisition Rate

OxyTrace+ is able to log data from the oxygen electrode and also from an optional auxiliary input at several different user-defined acquisition intervals. The intervals available for selection between each recording are:

100 msec	1 sec	10 sec
200 msec	2.5 sec	
500 msec	5 sec	



The acquisition rate settings are accessed by either selecting Hardware > Acquisition Rate from the menu bar or by clicking the icon from the tool bar. The window above shows information regarding the available data acquisition intervals. The table below shows further information for data acquisition rates that is not given in the Acquisition rate window:

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Acquisition Rate	Max Recording Time	Max Samples per Channel	Max Samples Total (8 Oxygen, 8 Auxiliary)	Estimated File Size
100 msec	100 mins	60,000	960,000	62 MB
200 msec	3 hrs	54,000	864,000	56 MB
500 msec	8 hrs	57,600	920,000	60 MB
1 sec	16 hrs	57,600	920,000	60 MB
2.5 sec	24 hrs	34,560	552,960	35.9 MB
5 sec	24 hrs	17,280	276,480	17.9 MB
10 sec	24 hrs	8,640	138,240	8.9 MB

The acquisition rate should be defined based upon the type of experiment to be performed. Assays where rapid response in oxygen tension is to be studied should use the fastest acquisition rate whereas experiments such as light response curves can afford a slower rate of data acquisition.

Once the acquisition rate has been selected, the OxyTrace+ software opens the Setup Live Rate Display window

Sample Stirring

Sample Stirring

Electrochemical reactions taking place at the cathode of the electrode disc require the consumption of oxygen in order to produce an electric current. This current is then digitised by an electrode control unit and presented as a chart recorder emulation in the OxyTrace+ software.

The nature of this process means that as oxygen is continuously consumed at the cathode, an oxygen depleted layer is produced immediately above the cathode. As the rate of consumption by the disc is greater than the rate of oxygen diffusion through liquid, the measured signal from the electrode disc will continuously decrease during measurement. This problem is overcome by continuously stirring the sample in order to replenish the depleted layer at the cathode and to ensure that the dissolved oxygen within the sample is kept evenly distributed throughout the reaction vessel.

The magnetic stirrer is an integral feature of the electrode control unit and is located on the top of the system. Stirring of the sample is achieved by placing a magnetic follower or "Flea" into the reaction vessel itself. Activating the stirrer rotates the magnet beneath the stirrer cap at user definable speeds of between 150 and 900 RPM. The flea tracks the magnetic stirrer, rotating just above the cathode of the electrode so as not to cause any damage to the membrane preparation. 2 types of flea are available:

PTFE coated magnet

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This is the standard flea supplied with the systems. It consists of a small magnetic bar coated in PTFE.

Glass flea

Glass fleas consist of several fine strands of iron sealed into a thin glass capillary tube.
Each type of flea has advantages and disadvantages of use. For example, for experiments where very small changes in oxygen tension are to be measured, some users prefer the glass fleas as PTFE is known to absorb a very minute amount of oxygen. In the large majority of applications however, this absorption is not an issue. Glass fleas are not magnetic themselves so they are not as reliable at tracking the rotating magnet in more viscous samples Therefore, the maximum speed of stirring would be lower than that of the PTFE coated flea. The effects of the oxygen depleted layer can be clearly seen by performing a short experiment:
Prepare the electrode disc and mount it into the base of the electrode chamber.

- Place the chamber on top of the magnetic stirrer.
- Add 2ml of air-saturated, de-ionised water into the chamber and drop a flea into the sample.
- Open the stirrer control window as described above and set the stirrer to a speed of 75%.
- Start a recording and allow the trace to stabilise.
- Once the trace has stabilised, stop the stirrer. The signal will start to decline steadily.
- Re-start the stirrer. The signal will return to it's original level.

The screenshot below shows how the trace should respond to the procedure outlined above.

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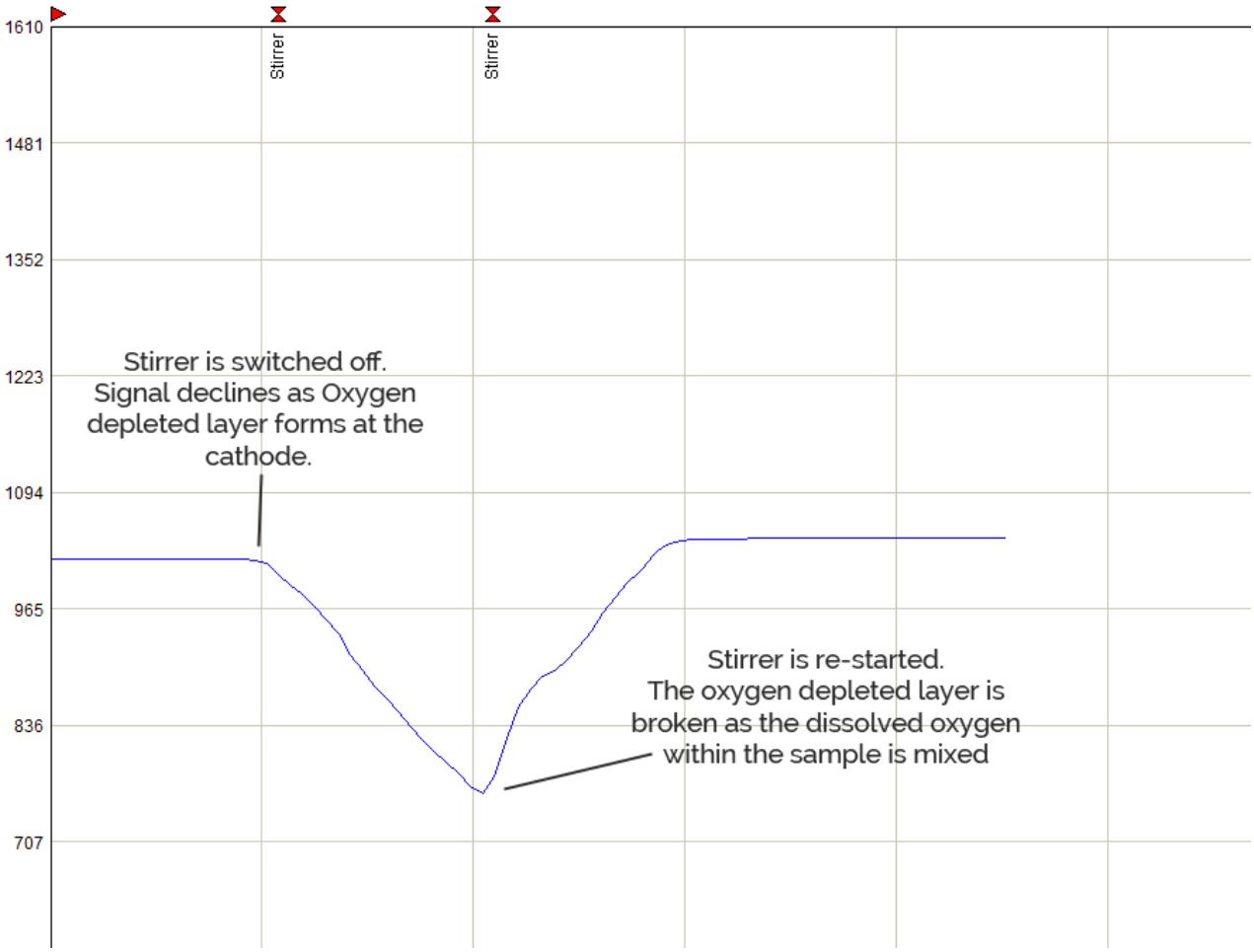

Operating the Magnetic Stirrer

The stirrer is controlled directly from the OxyTrace+ software. It can be accessed by selecting Hardware > Stirrer Speed from the menu or by pressing the icon on the toolbar.

Single Unit

If using a single unit system the following dialogue is generated.

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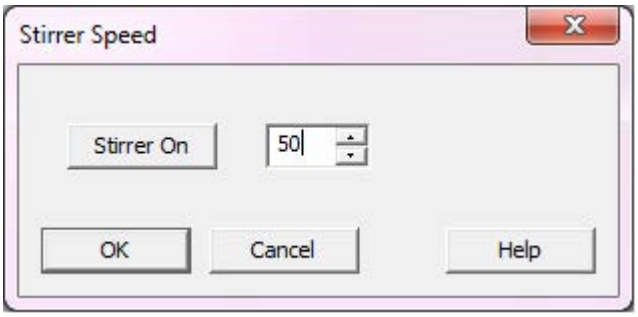
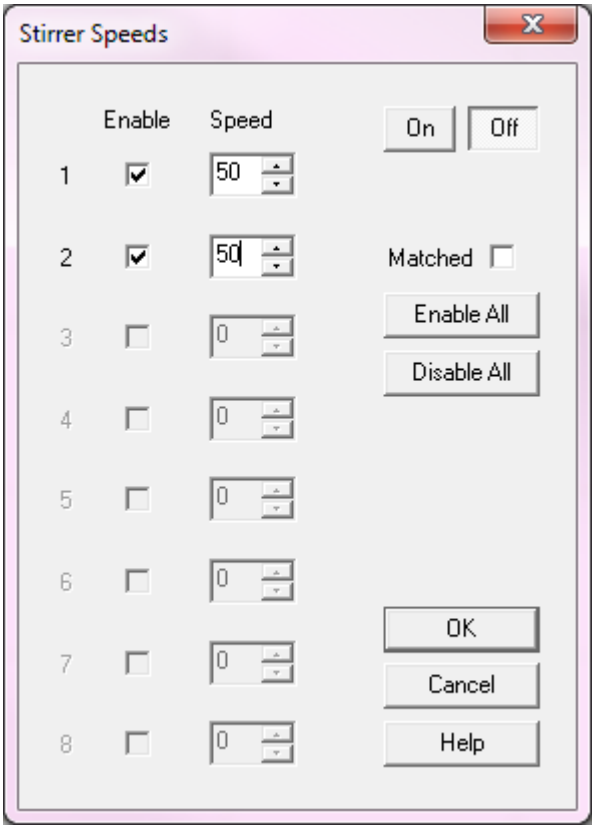


A stirrer speed is entered into the dialogue and can be adjusted with the up and down arrows. The 'Stirrer On' button will switch on the stirrer to the set speed.

Multi Channel

If using a multi-channel system the following dialogue is generated:



The 'Enable' check boxes allow the required stirrers to be selected individually. Alternatively 'Enable All' or 'Disable all' can be used to quickly select which stirrers are used.

The 'Matched' functions allows all stirrers to be quickly set to the same value. Pressing this will grey out the 'Speed' section for all other channels except for the first channel, this box is then

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used as the master speed. Any speed entered into this box will be updated for all available channels.

Once the stirrer speeds have been configured, the 'On' or 'Off' buttons will toggle the stirrers on and off. Stirrers can then also quickly be turned off/on bu pressing the icon from the toolbar.

Programme Options

The Options dialogue is accessed by selecting Tools > Options from the software menu. The following dialogue is displayed.

The functions of the options within this dialogue are explained below:

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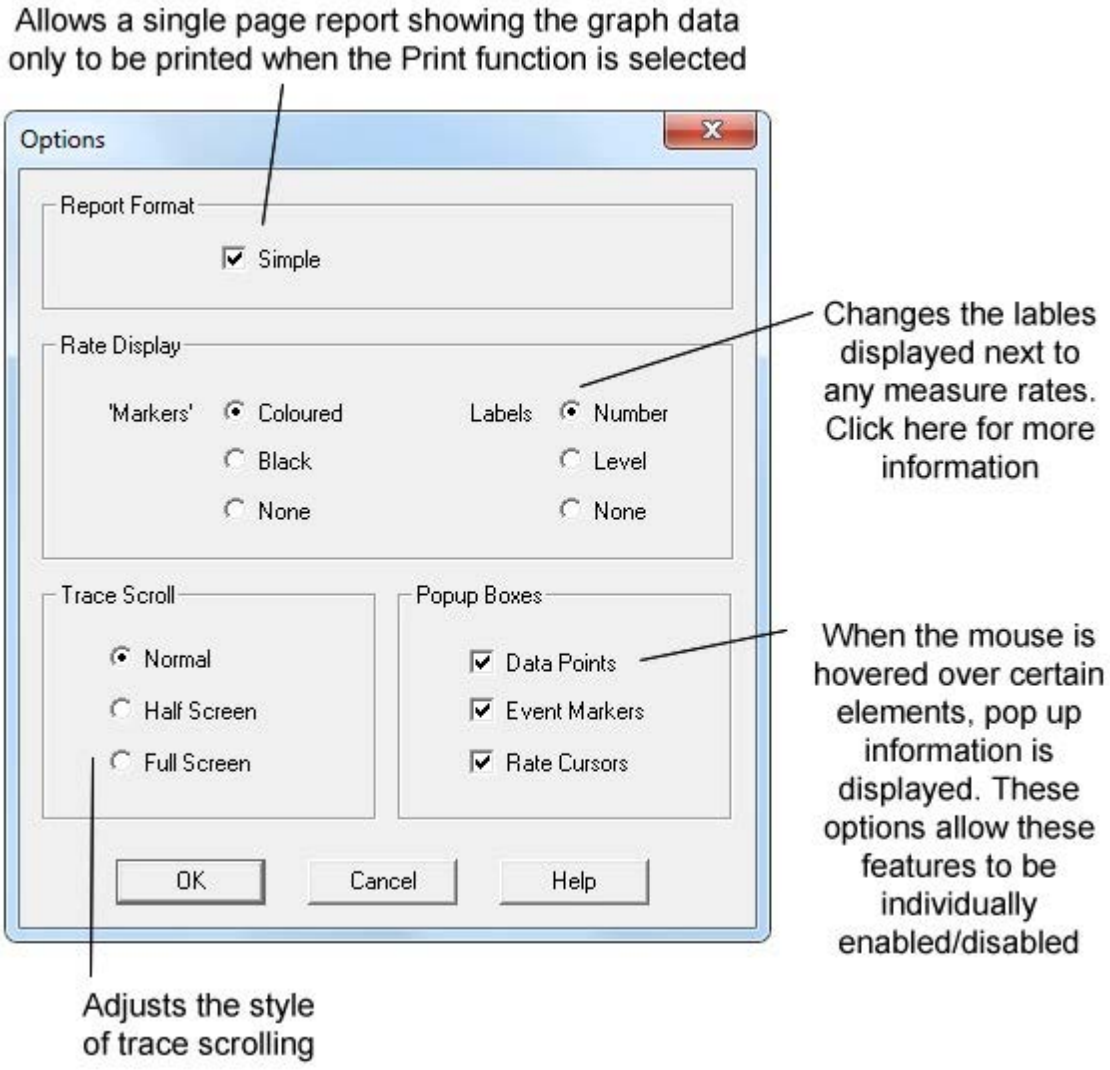

Quantum Yield Measurements

Quantum Yield Graph Options

The Quantum Yield Graph Setup window is displayed automatically at the end of a light response experiment. The image and points below describe the different areas of the Quantum Yield Graph Setup window and explain the functionality of each element.

The diagram below shows the dialogue that is presented when the system is calibrated in liquid-phase. There are several additional options that are displayed on the dialogue when the system is calibrated in gas-phase.

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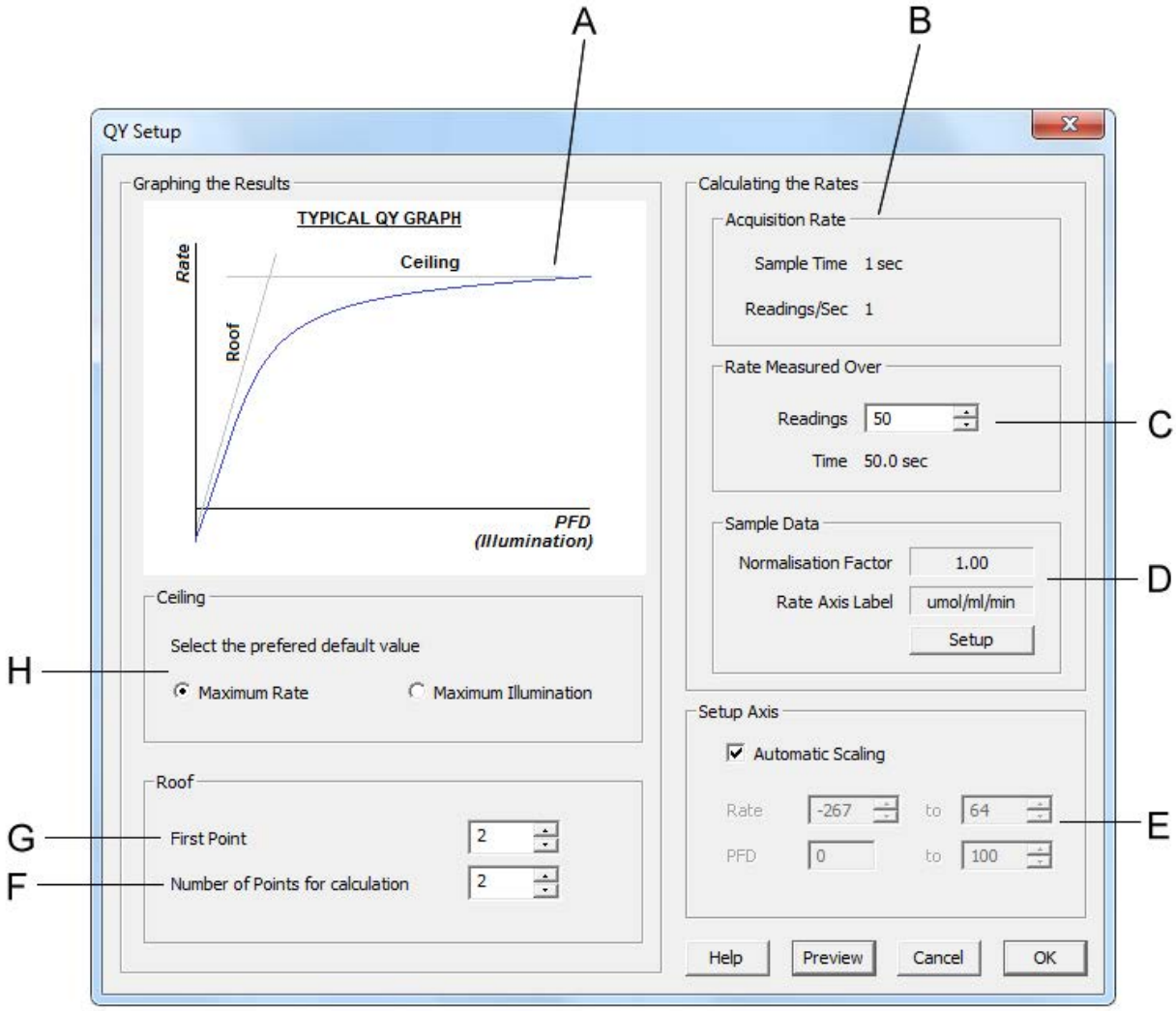

- A: Typical QY Graph

This diagram represents an ideal Quantum Yield graph and shows the roof and ceiling values.

The intercept on the vertical axis is a measure of dark respiration, that on the horizontal axis is the light compensation point. The initial slope is a measure of quantum yield and its reciprocal, quantum requirement. The curve lies within two constraints, a sloping "roof" and a "horizontal ceiling" (the "true ceiling" or maximum rate).

The roof is a thermodynamic constant pitched at an angle dictated by the maximal photosynthetic efficiency (in this instance a nominal quantum yield value of 0.111). The ceiling represents the absolute maximum rate of carbon assimilation. For purposes of comparison, a lower or "nominal" ceiling, imposed by the rate of carbon assimilation at 800 $\mu molsm^{-2}s^{-1}$ is employed.

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- B: Displays the data acquisition rate that was used in the light response experiment.
- C: The number of individual readings per light step that are used to calculate the rate of oxygen change.
- D: The default area of sample material taken into account when making the Quantum Yield calculation. Any deviation in this figure will need to be accounted for and an area correction should be made.
- E: Quantum Yield graph axes settings. Can be set to auto scale or can be scaled manually.
- F: The number of points on the graph that are taken into account when calculating the Roof or Slope of the data.
- G: The first point on the graph that is to be taken into account when calculating the Roof or Slope of the data.
- H: This option allows either the maximum rate measurement or the maximum level of illumination recorded to be used as the Ceiling value of the data.

1.5.3.4 System Calibration

Liquid Phase Calibration

Principles of Liquid-Phase Calibration

Before any measurements can take place, the electrode disc must be calibrated so that the electrical signal received from the disc can be presented as actual calibrated units (nmol/ml). Calibrating the disc for liquid-phase measurements involves a two step procedure in which the signal from the oxygen electrode is referenced to 2 known oxygen concentrations in order to derive an Offset and Calibration Factor.
The 2 calibration steps are:

Air Line

According to the studies of G.A. Truesdale and A.L. Downing (The solubility of oxygen in water, 1954, Nature 173: 1236), at any given temperature and atmospheric pressure, air saturated, deionised water contains a known concentration of dissolved oxygen which may be calculated mathematically. The following information is used by the OxyTrace+ software in order to accurately reference the electrical signal from the electrode for the air line stage of calibration: This data is based on measurements of dissolved oxygen in water at the given temperature and standard atmospheric pressure published by Truesdale & Downing (Nature 173:1236, 1954).

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Temperature (°C)	Oxygen (PPM)	Oxygen (nmol/ml)
0	14.16	442.5
5	12.37	386.6
10	10.92	341.3
15	9.76	305
20	8.84	276.3
25	8.11	253.4
30	7.52	235
35	7.02	219.4

The formula used in calculating the oxygen values in the table is as follows:

$$Cs=14.16 - (0.394 * T) + (0.007714 * T^2) - (0.0000646 * T^3)$$

{Where Cs is the oxygen saturated concentration in ppm and T is temperature in °C}

1 ppm is equivalent to 1µg/ml or (1µg/32g/mol) = 0.03125 µmol/ml

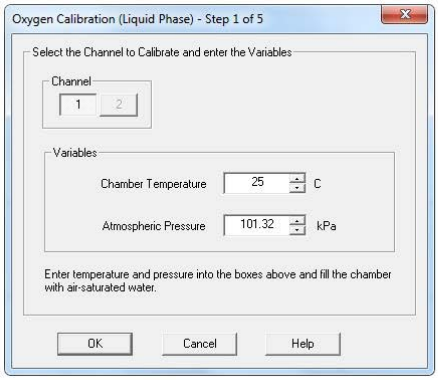
or

31.25 nmol/ml

At any atmospheric pressure other than standard atmospheric (760mm Hg or 101.32 KPa), the value obtained is adjusted by a factor of:

$$\frac{\text{Current atmospheric pressure}}{\text{standard atmospheric pressure}}$$

The calibration factor is therefore calculated as is shown in the diagram below:



Temperature and pressure variables cross referenced

Temperature (°C)	Oxygen (ppm)	Oxygen (nmol/ml)
0	14.16	442.5
5	12.37	386.6
10	10.92	341.3
15	9.76	305
20	8.84	276.3
25	8.11	253.4
30	7.52	235
35	7.02	219.4

1600 nA is equal to 253.4 nmol/ml

$$\frac{253.4}{1600}$$

Calibration factor 0.158

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In the example calibration above, the signal measured from the electrode in a stirred sample of air-saturated, de-ionised water was 1600 nA. In the first step of the calibration, the user is prompted to enter assay temperature and pressure variables. These entries are referenced against the look-up tables and formula of Truesdale and Downing (see above) in order to establish the amount of oxygen present within the chamber. This value, in this example, is 253.4 nmol/ml. The measured 1600 nA must therefore, be married to the nmol/ml value in order to obtain the calibration factor.

Zero Oxygen Line

In a perfect world, an oxygen concentration of zero would produce an electrical signal of zero from the electrode disc. However, the electrode does have what is known as residual current which does give a small signal in zero oxygen concentration.
It is necessary to subtract the value of signal caused by residual current from all recorded data points in order to give accurate results. This is known as the Offset. Zero oxygen within the reaction chamber can be achieved in 2 ways:

- Sodium Dithionite. - The addition of a strong oxygen reducing agent such as sodium dithionite provides a good method of achieving zero oxygen. However, great care must be taken to remove all traces of the dithionite before commencing measurement as even a minute amount of remaining dithionite will have a serious effect on the oxygen concentration of the sample. Once removed, the chamber should be thoroughly rinsed with distilled water several times.

IT IS CRUCIAL THAT CARE IS TAKEN, WHEN REMOVING THE SODIUM DITHIONITE AND WATER FROM THE REACTION VESSEL, THAT THE MEMBRANE OF THE ELECTRODE IS NOT DAMAGED. THE IDEAL METHOD OF REMOVING LIQUID FROM THE CHAMBER IS TO USE AN ASPIRATOR WITH A SOFT, RUBBER TIP (MIND THE FLEA!!!). IF AN ASPIRATOR IS NOT AVAILABLE, CAREFULLY USE A PASTEUR PIPETTE TO REMOVE THE LIQUID.

- Nitrogen. - A more convenient method of achieving zero oxygen is to bubble nitrogen gas through the liquid in the reaction vessel in order to displace all the oxygen. This method is in some ways safer than the use of sodium dithionite as there is no risk of contaminating the actual sample during the measurement and there is less risk of damaging the membrane of the disc. However, the use of nitrogen is a slower process and to achieve zero line is more difficult than dithionite.

Liquid Phase Calibration Process

Once the liquid-phase calibration routine has been selected, OxyTrace + will generate a series of prompts to guide you through the calibration process. Please follow the guidelines below to complete the liquid-phase calibration process.

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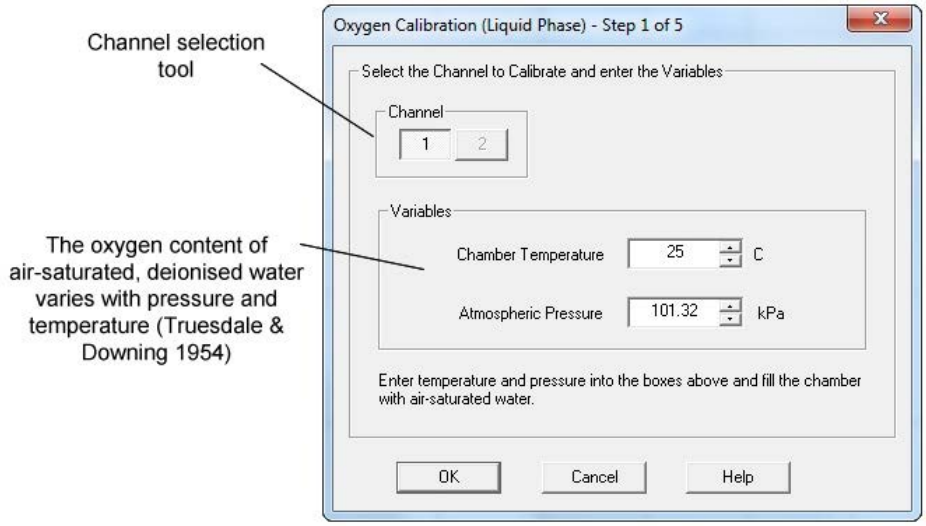
Air Line

Prepare the disc as described in the Electrode Preparation section, install into the base of the electrode chamber and connect the electrode disc to the rear of the control unit using the electrode connection cable. Place 2ml of air saturated, deionised water into the reaction vessel. Air saturated water is obtained by vigorously shaking a small quantity of deionised water (approx. 50ml) in a large conical flask (approx. 1L). In all systems (with the exception of the Oxytherm electrode control unit), connect the water jacket of the electrode chamber to the circulating water bath for temperature control purposes and ensure that the sample and electrode disc equilibrate to the temperature required by the assay before commencing calibration. Oxytherm users should set the chamber temperature directly from the OxyTrace + software temperature settings.

IT IS IMPORTANT TO REMEMBER THAT THE SAMPLE OF AIR SATURATED WATER SHOULD BE EQUILIBRATED TO THE ASSAY TEMPERATURE BEFORE THE CALIBRATION PROCEDURE BEGINS. IT CAN BE PRE-EQUILIBRATED IN A WATER BATH OR SIMPLY ALLOWED SUFFICIENT TIME TO REACH TEMPERATURE ONCE ADDED TO THE REACTION VESSEL.

The liquid-phase calibration sequence is activated either from the Calibrate > Liquid-Phase Calibration menu option or directly from the icon on the toolbar. Once activated, the following window is generated.



Entering the appropriate temperature and atmospheric pressure into this window allows the OxyTrace + software to accurately reference the air line. Whatever signal is recorded from the electrode disc during the air line phase will be subjected to the formula of Truesdale and Downing (see Principles of Liquid-Phase Calibration page) in order to derive the calibration factor. Once the correct variables have been entered into this window, proceed by pressing OK.
The following window is generated.

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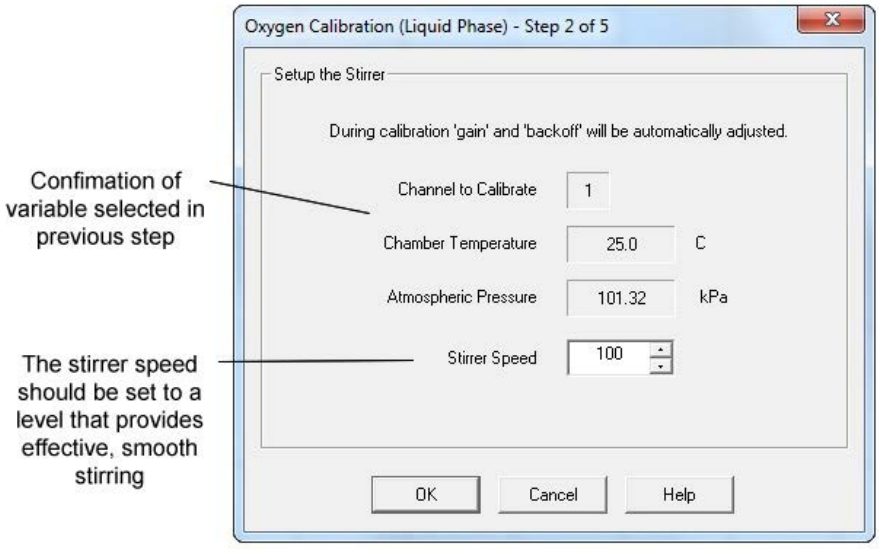
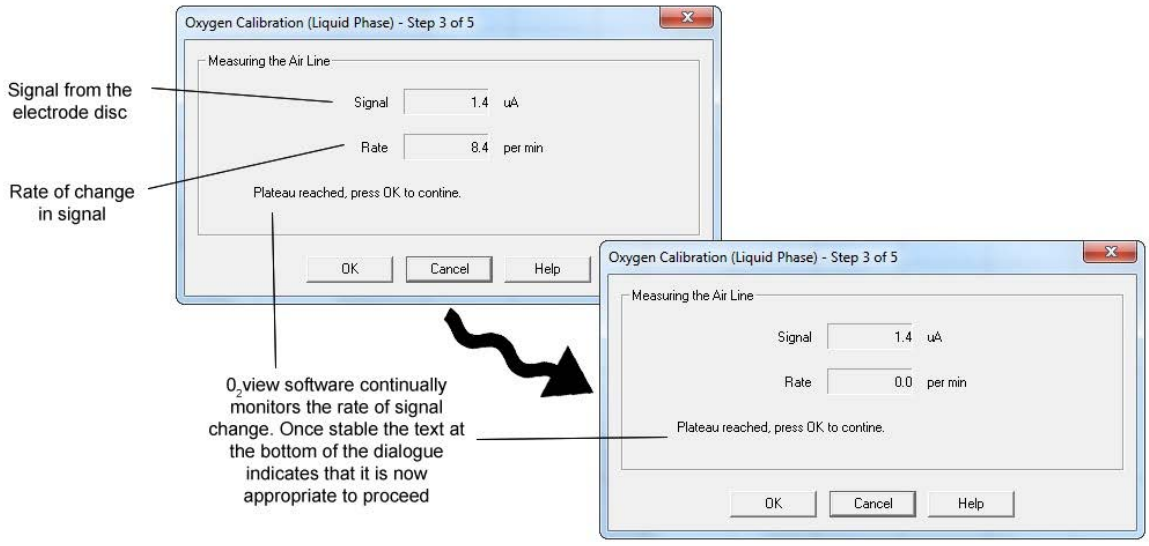


Assay temperature and ambient pressure variable defined in the previous window are displayed as confirmation. The stirrer setting should provide efficient, smooth stirring of the sample without causing a noisy signal. Please refer to the Stirrers section for details on selecting an appropriate stirring rate. Once the stirrer setting has been selected, proceed by pressing OK. The following window is generated.



Observe the mV signal in the window above and wait until the prompt that a plateau has been reached. Once the signal is deemed to be stable, the OK button should be pressed in order to proceed.

IN THE CASE OF A NEW OR WELL MAINTAINED ELECTRODE DISC, YOU WOULD IDEALLY EXPECT TO SEE A SIGNAL OF APPROX. 1600NA (± 240NA) WHEN MEASURING AIR LINE IN AIR SATURATED

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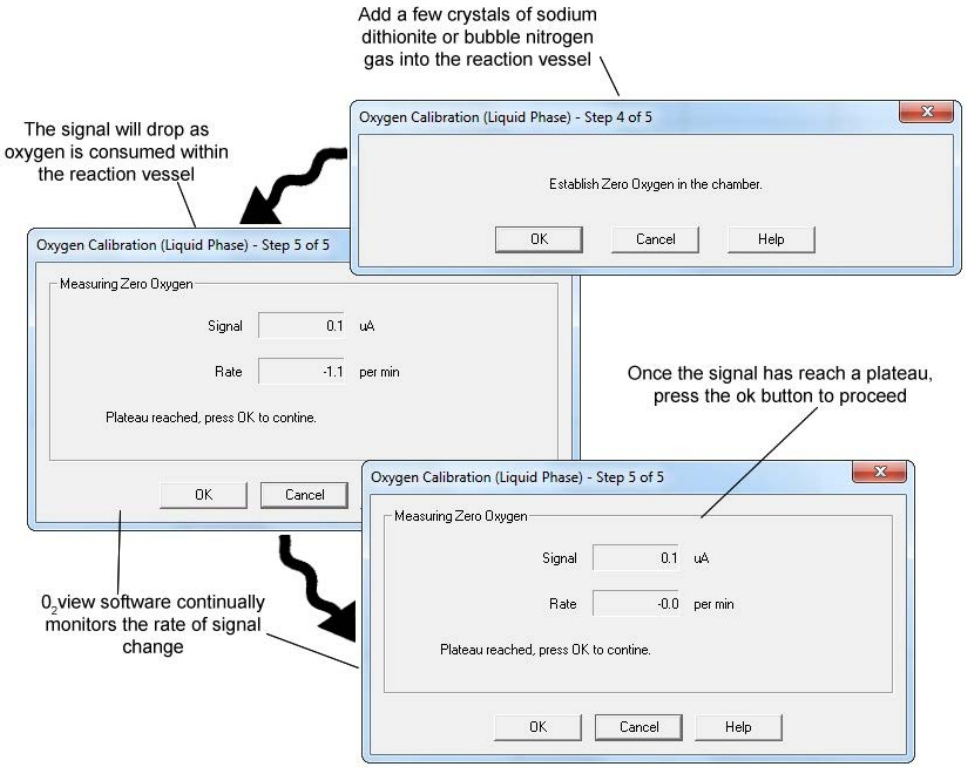
WATER. THE ACTUAL SIGNAL LEVEL AT THIS POINT IS NOT CRITICAL; IT IS MORE IMPORTANT THAT THE SIGNAL IS STABLE WITH MINIMAL NOISE. HOWEVER, IF THE SIGNAL LEVEL IS LOWER THAN 1300NA, IT COULD INDICATE A PROBLEM WITH THE ELECTRODE ITSELF.

If the signal from the electrode disc does not become stable after several minutes, this could be indicative of either one or a combination of several problems. For details on how to isolate the fault to either the electrode disc, the control unit or the electrode connection cable, please refer to the Electrode Disc Diagnostics section for further information.

Zero Oxygen Line

The second stage of the calibration procedure is to establish the zero oxygen line in order to determine the Offset (see see Principles of Liquid-Phase Calibration page). To perform this stage, use either of the following methods:

- Nitrogen gas - bubble nitrogen gas into the reaction vessel in order to displace all the oxygen in the sample.
- Sodium Dithionite - a few crystals of this strong reducing agent is sufficient to reduce all the oxygen dissolved in the sample.
-



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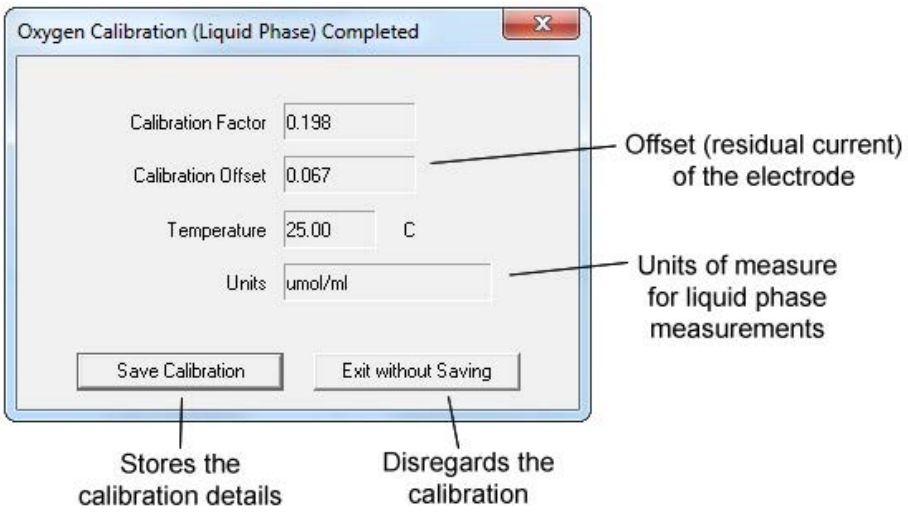
IN THE CASE OF A NEW OR WELL MAINTAINED ELECTRODE DISC, YOU WOULD NORMALLY EXPECT TO SEE A SIGNAL OF LESS THAN 1% OF THE SIGNAL OBSERVED DURING THE AIR LINE STAGE. FOR EXAMPLE, IF THE AIR LINE SIGNAL WAS 1506 NA, YOU WOULD EXPECT THE ZERO LINE TO MEASURE LESS THAN 15.06 NA.

If the zero oxygen line recorded from the disc seems to be too high, this could be indicative of one of several things:

- Insufficient oxygen reduction/removal from sample - sodium dithionite does have a finite shelf-life. If old dithionite is used, there is a possibility that the oxygen reducing capabilities will be significantly reduced or absent altogether. Always ensure that sodium dithionite is kept in small quantities in a clearly labelled air-tight container. Bubbling nitrogen into the chamber must be performed carefully in order to ensure that all oxygen has been displaced.
- Damaged electrode disc - unfortunately, most cases of high residual current reported are due to insufficient electrode maintenance. If the electrode is left to dry out after measurements have been completed, KCl crystallisation can irreversibly damage the disc by penetrating any minute fissures in the seal between the platinum cathode and the epoxy electrode dome and forcing the seal apart. This problem has a "snowball" effect as every time the disc is prepared from now on, KCl will enter the fissures causing them to widen. Please refer to the Electrode Maintenance section for further information.
- Once the zero oxygen stage has been completed, press OK to proceed. The following window is generated.



Details of the completed electrode calibration factor are presented in this window. Click Save Calibration to store the information or click Exit without saving to discard this calibration factor. If the calibration has been unsuccessful, OxyTrace + will display a warning message and an indication of which parameters have not been successful.

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Manual Calibration Process

Systems are used for measurement of samples in a wide range of different mediums and in some cases, it is desirable to calibrate the system using the medium in question in place of the air-saturated distilled water used in the standard, documented calibration routine.

O2view has a built calibration routine that allows a known concentration of oxygen to be assigned to a signal from the electrode disc.
If an alternative medium is to be used, it is important to remember that the solubility of oxygen in the medium will more than likely be different to that of distilled water.
The following example shows a manual calibration routine using seawater in place of air-saturated distilled water.

The oxygen content will depend upon both the salinity and temperature of the water. The oxygen content of sea water over a range of salinity and temperature has previously been determined by Green and Carritt (1967) and is presented in the table below.

Chloride Concentration				
	5000mgL^{-1}	10000 mgL^{-1}	15000 mgL^{-1}	20000 mgL^{-1}
Temperature (˚C)	Oxygen Concentration(µmolml-1)			
0	0.43	0.406	0.378	0.353
5	0.378	0.356	0.334	0.312
10	0.334	0.315	0.3	0.281
15	0.303	0.284	0.268	0.253
20	0.272	0.259	0.247	0.231
25	0.25	0.237	0.225	0.209

ALTHOUGH THIS CALIBRATION ROUTINE HAS BEEN REFERRED TO AS SEAWATER CALIBRATION, IT IS POSSIBLE TO USE THIS ROUTINE TO CALIBRATE THE SYSTEM USING ANY OTHER MEDIUM PROVIDED THE OXYGEN CONCENTRATION OF THE MEDIUM IS KNOWN

Performing the Manual Calibration

Prepare the electrode disc and mount it in the electrode chamber. Ensure the disc is responding correctly before adding the seawater that is to be used for the calibration routine to the electrode chamber ensuring that the seawater is at the correct temperature for the experiment. Allow the electrode disc signal to stabilise. From the Menu bar, select Calibrate > Liquid Phase Calibration > Manual . The following dialogue opens.

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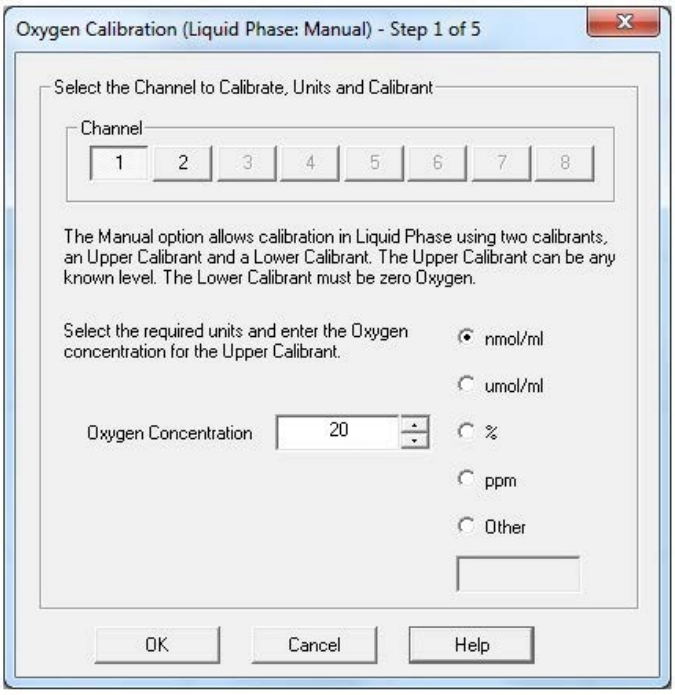

Enter the known oxygen concentration into the Oxygen concentration input. This value will be referenced to the initial, stable oxygen electrode signal level in place of the standard airline measurement. Then select the choice of units you wish to use by either selecting one of the given units or by using 'Other' and entering any units you desire.

Once this step of the calibration routine is complete, the calibration routine is identical to the standard liquid-phase calibration routine. Please refer to the Liquid-Phase Calibration section for further information.

Viewing Calibration Details

Once the system has been successfully calibrated in either liquid or gas-phase, details of the calibration may be viewed in a separate window. This window is accessed from the Calibrate > Oxygen Calibration Details option from the menu bar.
All aspects of the calibration are shown in the window as shown in the diagram below:

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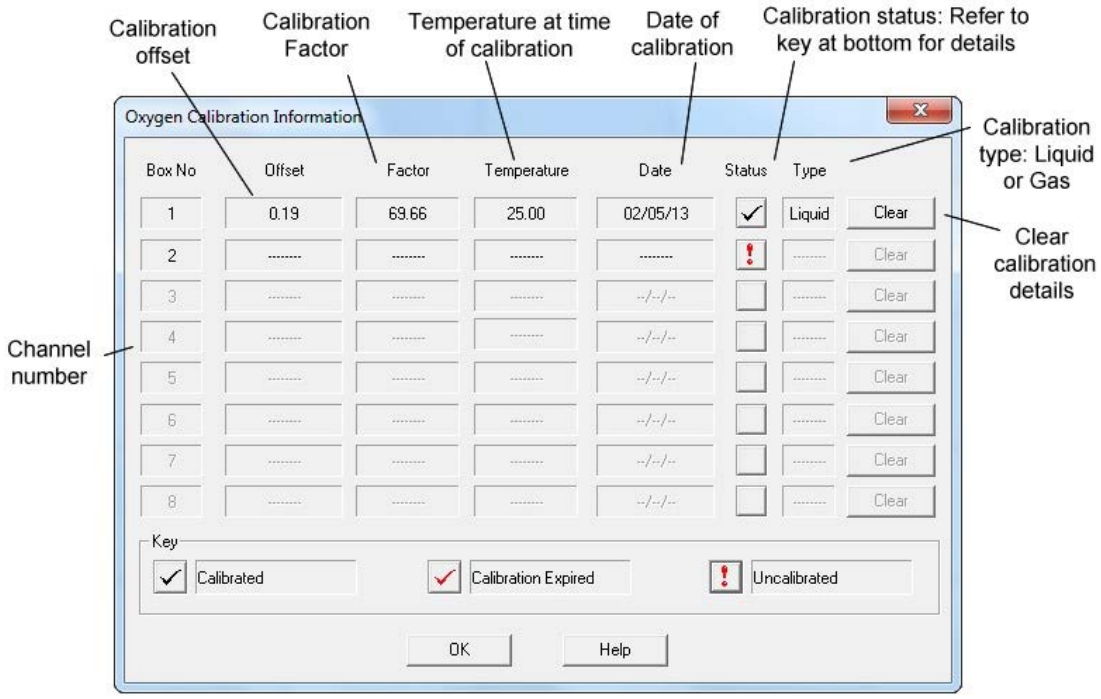

The Date column is relevant should the apparatus be intended for long measurement assays lasting more than a day or if the electrode is to be left prepared and in situ (please note that this should only be performed in liquid-phase electrode chambers as electrodes in gas-phase "dry out"). A calibration warning interval may be set so that after a given number of days post calibration, the user is prompted that the calibration has expired when the software is initially run. Please refer to the Set Oxygen Calibration Warning section for further information.

Set Calibration Warning

If the apparatus is intended for long measurement assays lasting more than a day or if the electrode is to be left prepared and in situ (please note that this should only be performed in liquid-phase electrode chambers as electrodes in gas-phase "dry out"), a calibration warning interval may be set so that after a given number of days post calibration, the user is prompted that the calibration has expired when the software is initially run. A calibration warning can also be set in the same manner for any auxiliary devices connected to the instrument.

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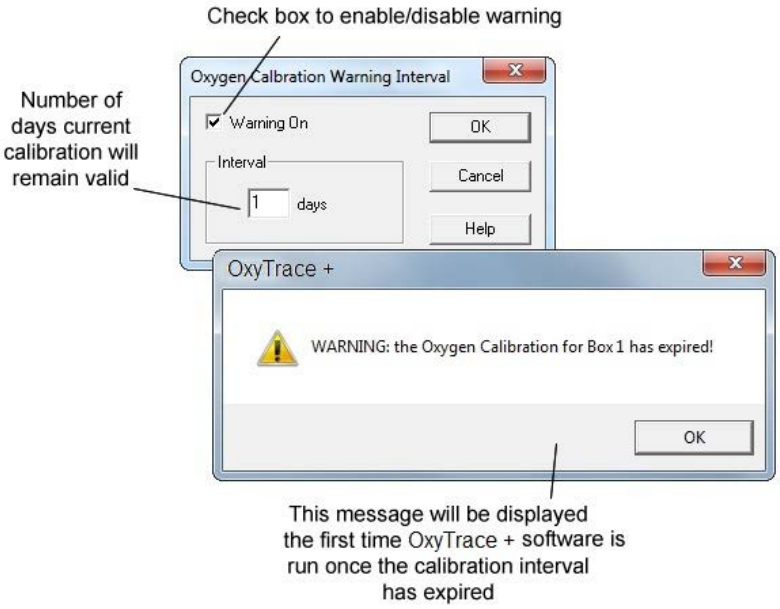

IT IS IMPORTANT TO REMEMBER THAT OVER TIME, A POLARISED DISC BUILDS UP A DEPOSIT OF BLACK SILVER OXIDE DUE TO THE NATURE OF THE ELECTROCHEMICAL REACTIONS TAKING PLACE. A DISC THAT IS LEFT MEASURING FOR MORE THAN A DAY MAY BEHAVE DIFFERENTLY AT THE END OF THE ASSAY THAN AT THE BEGINNING. PLEASE REFER TO THE SECTION LEAVING THE ELECTRODE DISC POLARISED OVERNIGHT.

Gas Phase Calibration

Principles of Gas-Phase Calibration

Before any measurements can take place, the electrode disc must be calibrated so that the electrical signal received from the disc can be presented as actual calibrated units (µmol Relative To Air). Calibrating the disc for gas-phase measurements involves a three step procedure in which the signal from the oxygen electrode is referenced to known oxygen concentrations in order to derive an Offset and Calibration Factor. The following steps are used in order to calibrate the electrode:

Air Line

The air-line refers to the signal recorded by the electrode disc in ambient air. It is this value that is used as the calibration offset.

Injection / Removal Line

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In order to obtain a calibration factor, the software requires the addition of a known oxygen concentration to reference against a level of change in the bit signal from the disc. This is performed by adding or removing 1ml of air from the electrode chamber using a syringe. Either addition or removal of air results in the same calibration factor. The user is given both options depending on the type of assay performed. For example, oxygen evolution measurements should use an injection of 1ml, oxygen consumption measurements should use a removal of 1ml of air as good laboratory practice.

At given temperature and pressure, gas laws dictate that the amount of oxygen present in 1ml of air can be easily calculated. At standard temperature (273.15 degrees Kelvin or 0 °C) and pressure (101.32 kPa or 1 atmosphere), the amount of oxygen in 1ml of air (containing 21% by volume) is 210µl. Since 1 mole of gas occupies 22.414µl, this is equivalent to:

$$\frac{210}{22.414}$$

OR

9.37Ì MOLES

At any other temperature, the amount can be derived by multiplying by a factor of:

$$\frac{273}{(273+T°C)}$$

(where T is the experimental temperature) so that, for example, at 25°C :

$$\frac{273}{(273+25°C)} = 0.9161$$

9.37 Ì MOL X 0.9161 = 8.58

THEREFORE, 1ML OF AIR AT 25°C CONTAINS 8.58 Ì MOL OF OXYGEN.

At any other pressure the amount derived above should be adjusted by a factor of:

$$\frac{\text{CURRENT ATMOSPHERIC PRESSURE}}{\text{101.32 KPA (STANDARD ATMOSPHERIC PRESSURE)}}$$

This factor is then married with the change in bit level during the injection / removal of air in order to obtain a calibration factor. The following diagram shows how this factor is obtained.

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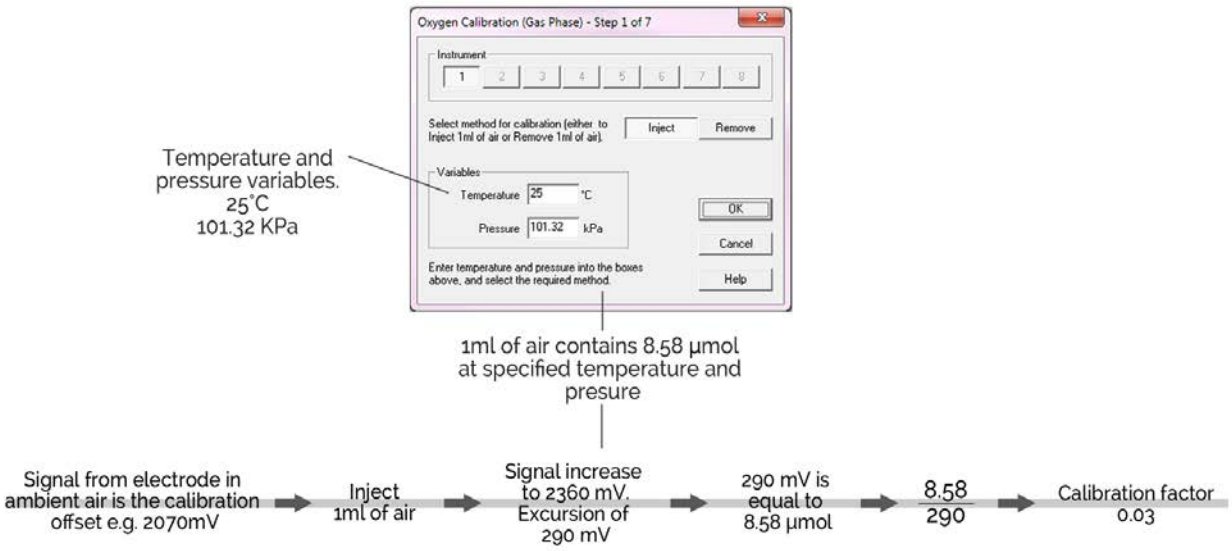

Gas Phase Calibration Process

Please follow the guidelines below for the gas-phase calibration sequence:

- Prepare the disc as described in the Electrode Preparation and Maintenance manual (supplied with the S1 electrode) and install into the base of the electrode chamber.
- Connect the electrode disc to the rear of the control unit using the electrode connection cable.
- Ensure that the appropriate sample mounts and gas-taps are fitted to the electrode chamber (refer to either of the Gas-Phase Chambers manuals).
- Connect the water jacket of the electrode chamber to the circulating water bath for temperature control purposes and ensure that the leaf chamber and electrode disc equilibrate to the temperature required by the assay before commencing calibration.

Step 1

The gas-phase calibration sequence is initiated by selecting Calibrate > Gas Phase Calibration Once selected, the following window is generated.

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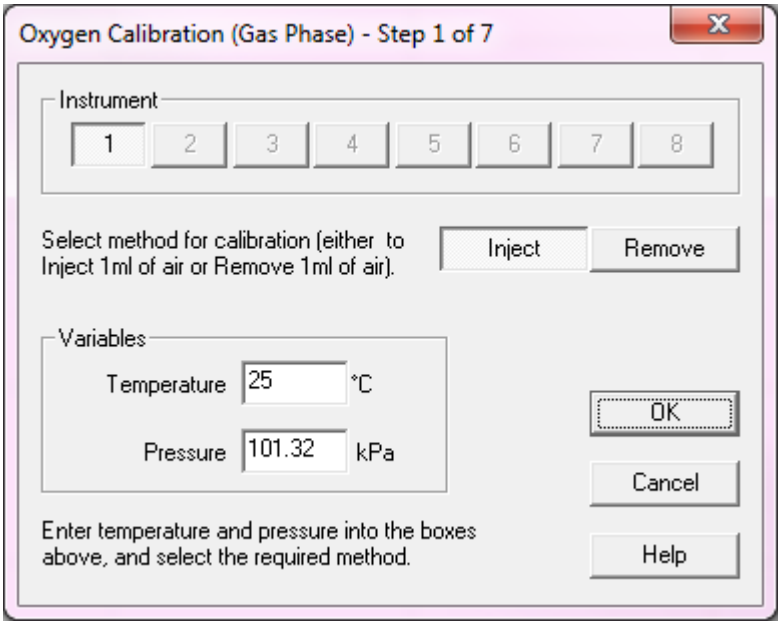
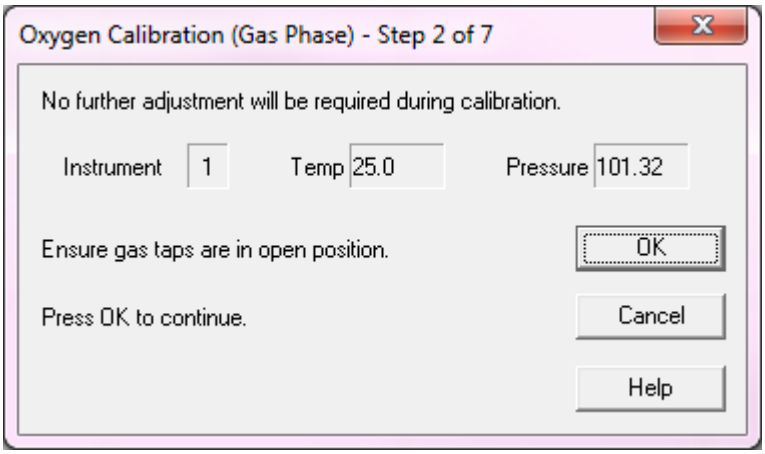


Select method for calibration to either inject 1ml of air or remove 1ml of air. Then enter the relevant temperature and air pressure variables into this dialogue and press OK.

Step 2

Ensure that the gas taps are in the open position and press OK to continue



Step 3

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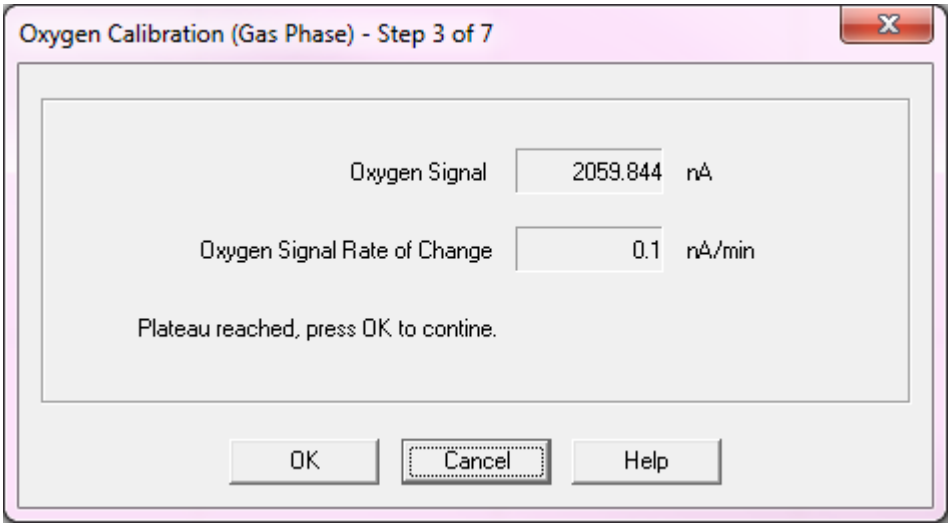

Here both the oxygen signal and rate of change are displayed. The software will wait until a plateau has been reached before moving onto the next step "Please wait for the signal to plateau " is displayed at this time as well as an override button which allow you to bypass this wait time if you are happy with the signal level (BEWARE THIS MAY AFFECT THE ACCURACY OF THE CALIBRATION IF THE SIGNAL IS NOT FULLY STABILISED). The value that has been recorded for the air line will be used as the Calibration Offset.

During measurement, the signal recorded from the disc in ambient air will be displayed as the origin of the axes (0). Any oxygen evolution / consumption by a sample will be displayed in the calibrated unit µmol relative to the air line.

IN THE CASE OF A NEW OR WELL MAINTAINED ELECTRODE DISC, YOU WOULD NORMALLY EXPECT TO SEE A SIGNAL OF APPROX. 2050MV (HALF THE RANGE OF THE CONTROL UNIT RESOLUTION) WHEN MEASURING AIR LINE IN A GAS-PHASE ELECTRODE CHAMBER.

If the signal from the electrode disc does not become stable after several minutes, this could be indicative of either one or a combination of several problems. For details on how to isolate the fault to either the electrode disc, chamber, the control unit or the electrode connection cable, please refer to the System Diagnostics section.

Once the signal has stabilized OK will be displayed

Step 4

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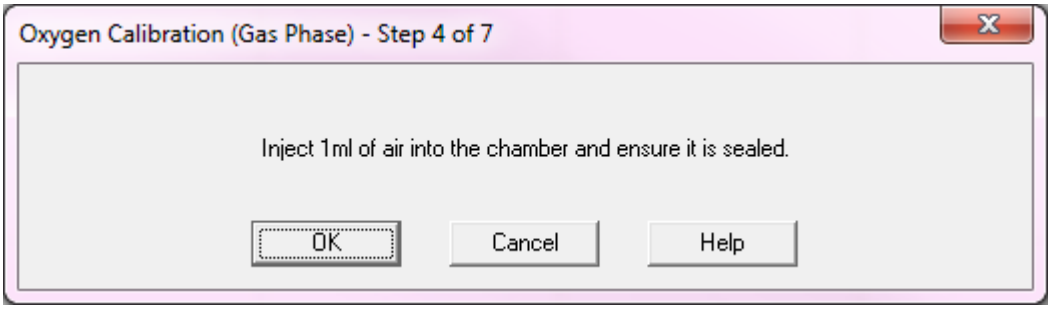

This stage of the calibration procedure is to establish the Injection / Removal line in order to determine the Calibration Factor. By injecting / removing 1ml or air to / from the electrode chamber, the bit signal recorded by the disc will change. The final amount of bits excursion in signal will then be referenced to a calibrated unit (µmol) taking into account any temperature and pressure variables entered in step 1. Please refer to the previous section for a full description of the mathematics involved in this calibration step.

Before selecting OK at this dialogue, use a syringe fitted securely into the gas port and either remove or add 1ml of air into the chamber (depending on the option selected in step 1). LD2/3 electrode chamber users should ensure that the
second gas port is closed.

Quickly close the gas port once the air has been injected/removed and press OK.

Step 5

At this point an increase/decrease of oxygen signal should be observed depending on option to inject/remove air.

IN THE CASE OF A NEW OR WELL MAINTAINED ELECTRODE DISC, YOU WOULD NORMALLY EXPECT TO SEE A SIGNAL CHANGE OF APPROX. 9% OF THE MEASURED AIR LINE AFTER A 1 ML INJECTION / REMOVAL.

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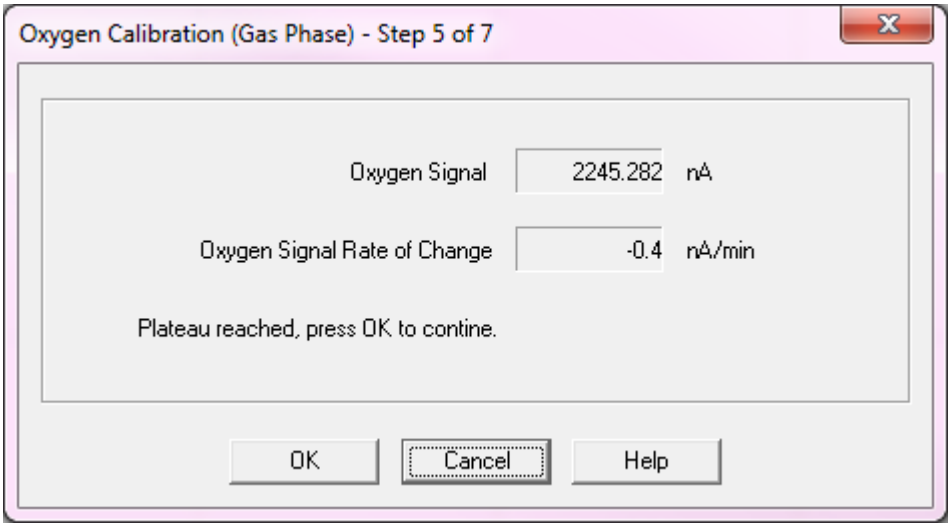
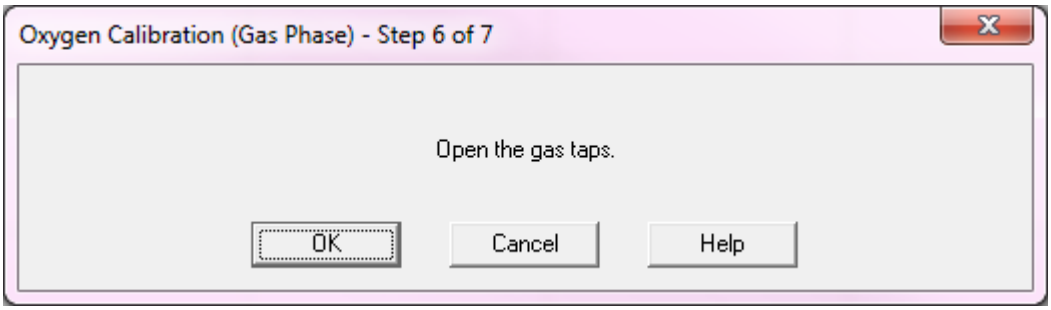


Allow the signal to stabilise at the new level just as with step 3. Once the oxygen signal has reached a plateua OK will be displayed.

If the signal does not stabilise and begins to drift downwards/upwards, this is indicative of a leak in the chamber. Please refer to the System Diagnostics section for details on isolating and fixing leaks in gas-phase electrode chambers.

Step 6

In this step of the calibration, the air-line is re-checked by opening the gas taps and measuring the signal from ambient air in exactly the same way as in calibration step 3. Press OK to continue



Step 7

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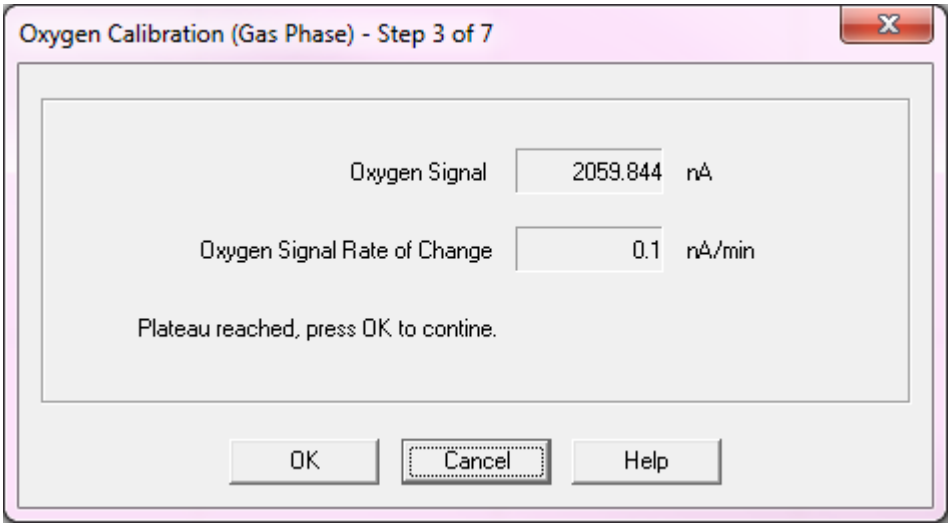
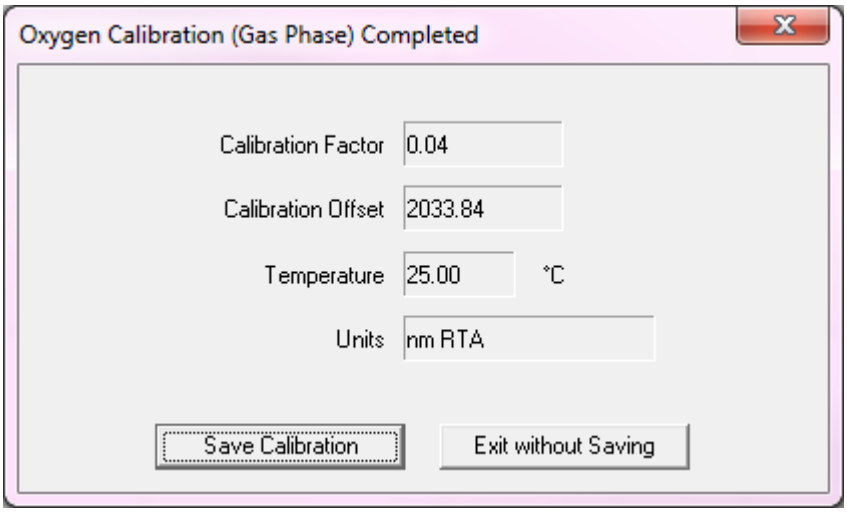


Follow the same procedure as step 3.

Step 8



Once the calibration is complete, details of the factor and offset applied during the calibration will be displayed. The calibrated units and temperature are also displayed.

The Save Calibration option will update the instrument with the calibration factor and offset ready for measurements to be taken.

The exit without saving option will disregard this calibration and will revert the instrument to its previous state, either uncalibrated or using a previously saved calibration.

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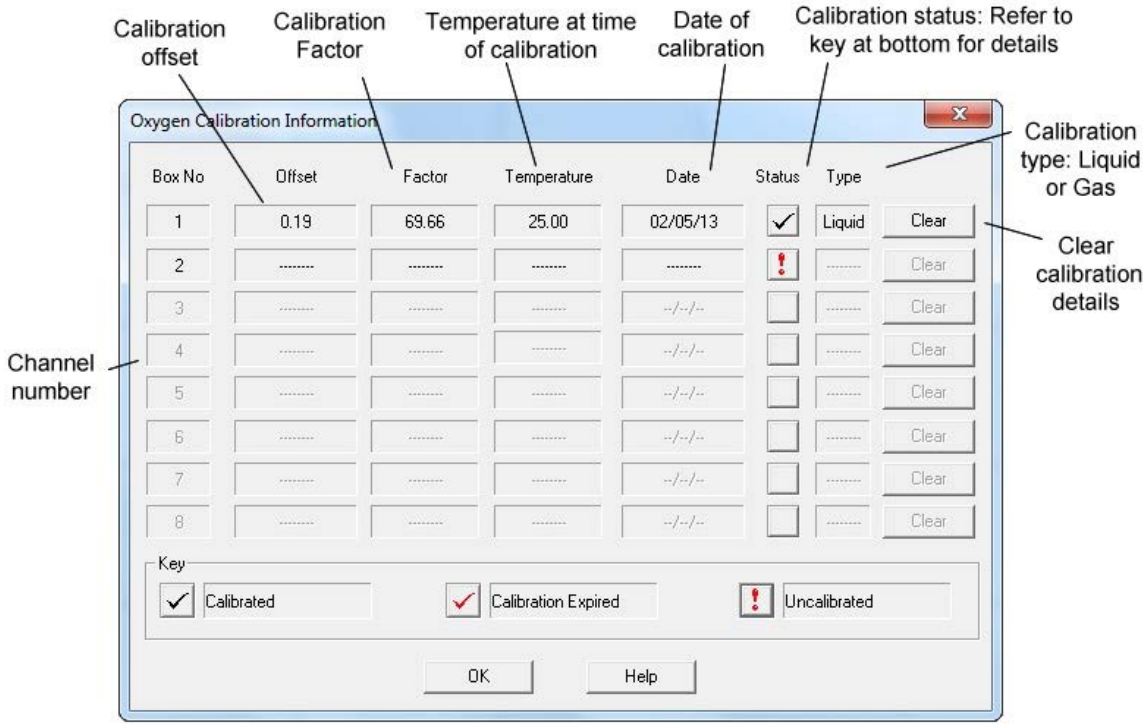
Viewing Calibration Details

Once the system has been successfully calibrated in either liquid or gas-phase, details of the calibration may be viewed in a separate window. This window is accessed from the Calibrate > Oxygen Calibration Details option from the menu bar.
All aspects of the calibration are shown in the window as shown in the diagram below:



The Date column is relevant should the apparatus be intended for long measurement assays lasting more than a day or if the electrode is to be left prepared and in situ (please note that this should only be performed in liquid-phase electrode chambers as electrodes in gas-phase "dry out"). A calibration warning interval may be set so that after a given number of days post calibration, the user is prompted that the calibration has expired when the software is initially run. Please refer to the Set Oxygen Calibration Warning section for further information.

Set Calibration Warning

If the apparatus is intended for long measurement assays lasting more than a day or if the electrode is to be left prepared and in situ (please note that this should only be performed in liquid-phase electrode chambers as electrodes in gas-phase "dry out"), a calibration warning interval may be set so that after a given number of days post calibration, the user is prompted that the calibration has expired when the software is initially run. A calibration warning can also be set in the same manner for any auxiliary devices connected to the instrument.

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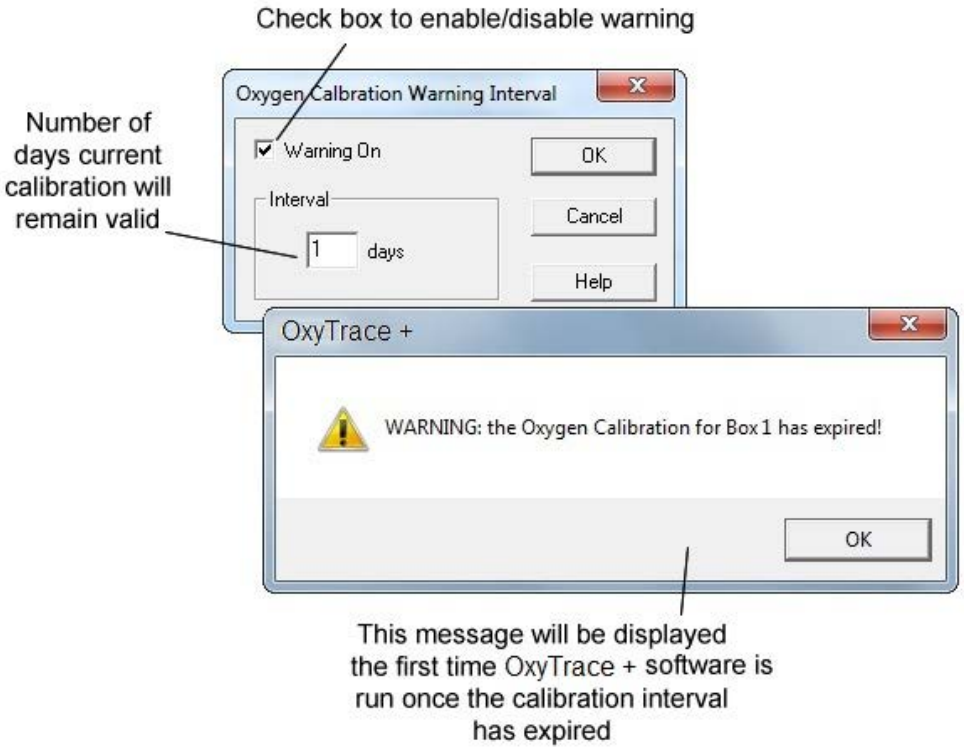

IT IS IMPORTANT TO REMEMBER THAT OVER TIME, A POLARISED DISC BUILDS UP A DEPOSIT OF BLACK SILVER OXIDE DUE TO THE NATURE OF THE ELECTROCHEMICAL REACTIONS TAKING PLACE. A DISC THAT IS LEFT MEASURING FOR MORE THAN A DAY MAY BEHAVE DIFFERENTLY AT THE END OF THE ASSAY THAN AT THE BEGINNING. PLEASE REFER TO THE SECTION LEAVING THE ELECTRODE DISC POLARISED OVERNIGHT.

Light Source Calibration

Light Source Calibration_2_2

Before any light response measurements are made, the light source requires calibration so that the Oxylab+ electrode control unit can set the light intensities at calibrated units of µmolsm-2s-1. Regardless of the light source in use, the calibration routine is identical. The light source is set at 10 increasing intensities between minimum and maximum output and a calibration factor is derived from the measured values at each step.

For Chlorolab 2+ and 3+ electrode systems, the QTP1 PAR/Temperature probe may be connected to the rear of the Oxylab+ control unit allowing an automatic light calibration routine to be used. Please refer to the Mounting QTP1 in the Electrode Chamber section for details on how to setup the electrode chamber, light source and QTP1 probe for calibration.

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For the Leaflab 2+ electrode system, the QSRED cannot be directly interfaced with the Oxylab control unit so the manual light calibration routine should be used.

To setup the QSRED, LD2/3 electrode chamber and LH36/2R light source ready for calibration, please follow the steps below referring to the diagram below:

- Place the LH36/2R on the bench, fan side down.
- Remove the upper section of the LD2/3 electrode chamber and place this on to the LH36/2R light source with the upper window interfacing with the light source.
- Place the large area sensor of the QSRED onto the lower window of the LD2/3 upper section. This should be facing upwards.
- Follow the instructions below for manual calibration of the light source using QSRED.

Both Automatic and Manual light calibration routines can be initiated from directly within the PFD Setup Table or from the Calibrate menu.

Before selecting the calibration type, ensure that the PAR sensor is positioned optimally for the light source in use.

Automatic Calibration

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The Oxylab+ control unit will automatically step through the 10 light settings and record the measured PAR value for each step. At the end of the 10 steps, OxyTrace+ will calculate the calibration factor.

IN ORDER TO CALIBRATE TWO LED1 LIGHT SOURCES IT IS NOT POSSIBLE TO USE THE AUTOMATIC CALIBRATION ROUTINE. TO CALIBRATE TWO LED1 LIGHT SOURCES A MANUAL CALIBRATION ROUTINE MUST BE USED; AT EACH LIGHT STEP THE QTP1 MUST BE TURNED TO FACE EACH OF THE LIGHT SOURCES IN TURN AND THE INTENSITY ADDED BEFORE ENTERING THE VALUES IN THE SOFTWARE.

Manual Calibration

Oxylab will set the light source intensity at the lowest level and prompt for a PAR value from an external PAR sensor. When this value has been entered, the light source will be set to the next intensity step. This process repeats until 10 light intensity steps between minimum and maximum intensity have been recorded. At the end of the 10 steps, OxyTrace+ will calculate the calibration factor.

Viewing Light Calibration Details

Light source calibration details may be viewed by selecting Calibrate > Light Calibration Details from the menu bar. The dialogue that is generated indicates calibration status (calibrated, calibration expired or uncalibrated), maximum light intensity available and date of calibration.

Set Light Calibration Warning Interval

This function allows the light source calibration to be assigned a validity period in weeks. Once the validity period expires, the light source calibration status changes to calibration expired and a warning message will be generated by OxyTrace+ prompting a light source re-calibration recommendation.

1.5.3.5 Data Handling

Event Marking

Adding Event Marks

During an experiment, certain events may occur that require documentation. For example, in a study of mitochondrial response to the addition of ADP would require a marker to indicate at what point the ADP was added so that the speed at which the mitochondria responded to the addition can be assessed.

OxyTrace+ makes provisions for this by allowing the addition of event marks with user-defined labels and comments. At any point during the measurement, an event mark can be added either by selecting Tools > Add Event Mark from the menu bar, by clicking the icon on the tool bar or by pressing the F5 function key on the keyboard. The following window is generated:

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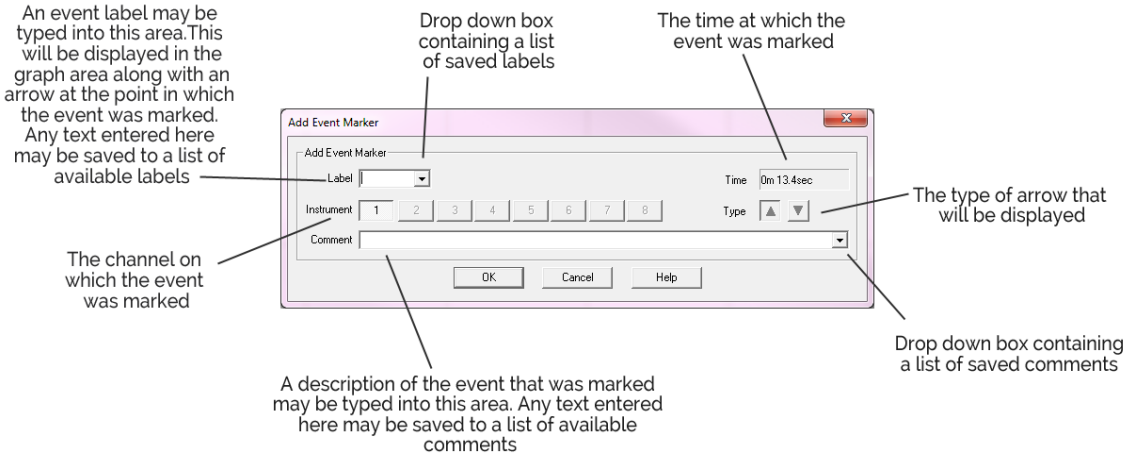
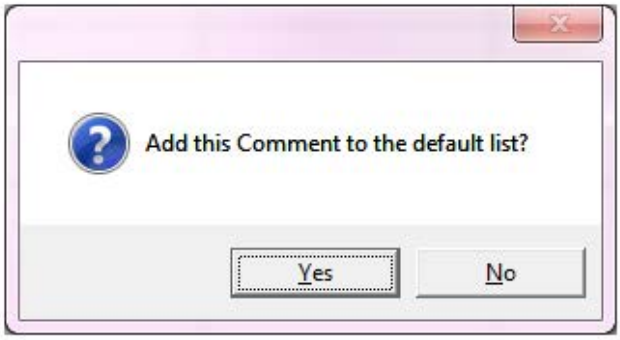
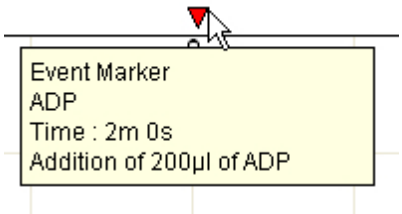


Any text typed into the label or comment fields of the Add Event Marker window may be saved to the OxyTrace+ configuration file on exiting. The following dialogues are generated allowing the text to be saved or not:



Saved labels and comments will then be available by selecting from the drop down boxes.

PLEASE NOTE THAT IF THE CONFIGURATION FILE IS DELETED FOR ANY REASON, THE LABELS AND COMMENTS THAT HAVE BEEN PREVIOUSLY DEFINED WILL BE LOST.



Event markers are displayed at the top of the graph area along with any label that has been assigned. Any comment that has been added to the event mark is displayed when the mouse

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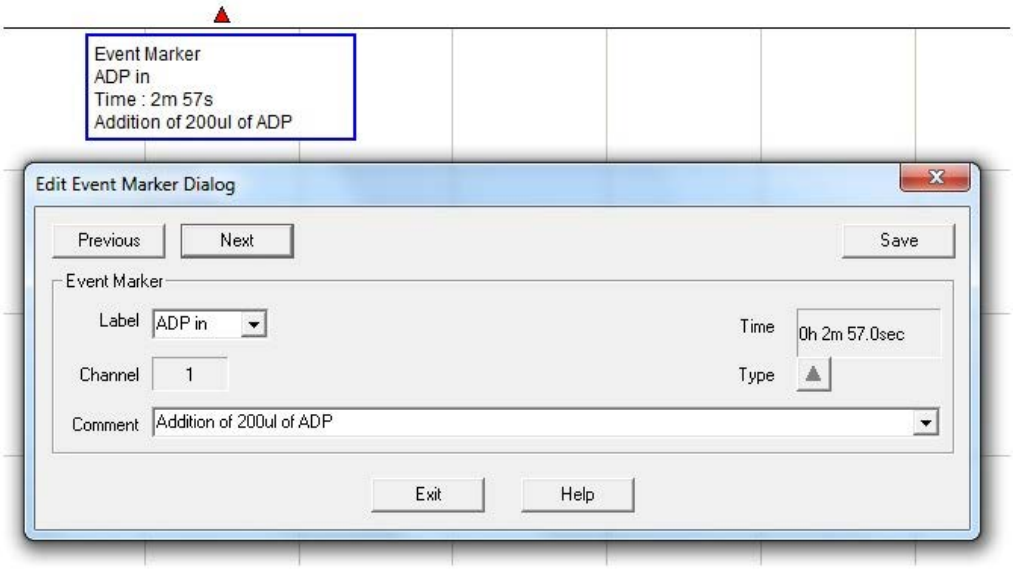
cursor is hovered over the event mark on the graph as shown in the diagram above. This feature may also be disabled from the Options menu. Event markers may be reviewed and edited or deleted at any point during and after the measurement. Events are also saved with the experiment and are printed.

For some functions of the software, event markers are automatically added to the graph and cannot be edited or deleted. These features include:

- Start recording - indicated by a right-facing arrow and the label "Start"
- Stop recording - indicated by a left-facing arrow and the label "Stop"
- Stirrer speed adjustment - indicated by 2 downwards arrows with the label "Stirrers". The first mark is added when the stirrer window is first opened, the second when the stirrer window is closed. This indicates that between these 2 points, stirrer settings were being adjusted
- Light on - indicated by a light symbol and the label Light on
- Light intensity change - indicated by a light symbol with either an up or down pointing arrow to indicate an increase or decrease in intensity.

Editing Event Marks

Event marks can be reviewed and edited or deleted at any point during and after the experiment. Selecting Tools > Edit Event Marks from the menu bar opens the following window.



The event to be edited is displayed in a blue box along with the information associated with it. If any changes are made, the Save button should be pressed before either exiting the window or moving to another mark using the previous or next buttons. Please note that only the text associated with the event mark may be changed. The type of event (up or down) cannot be amended.

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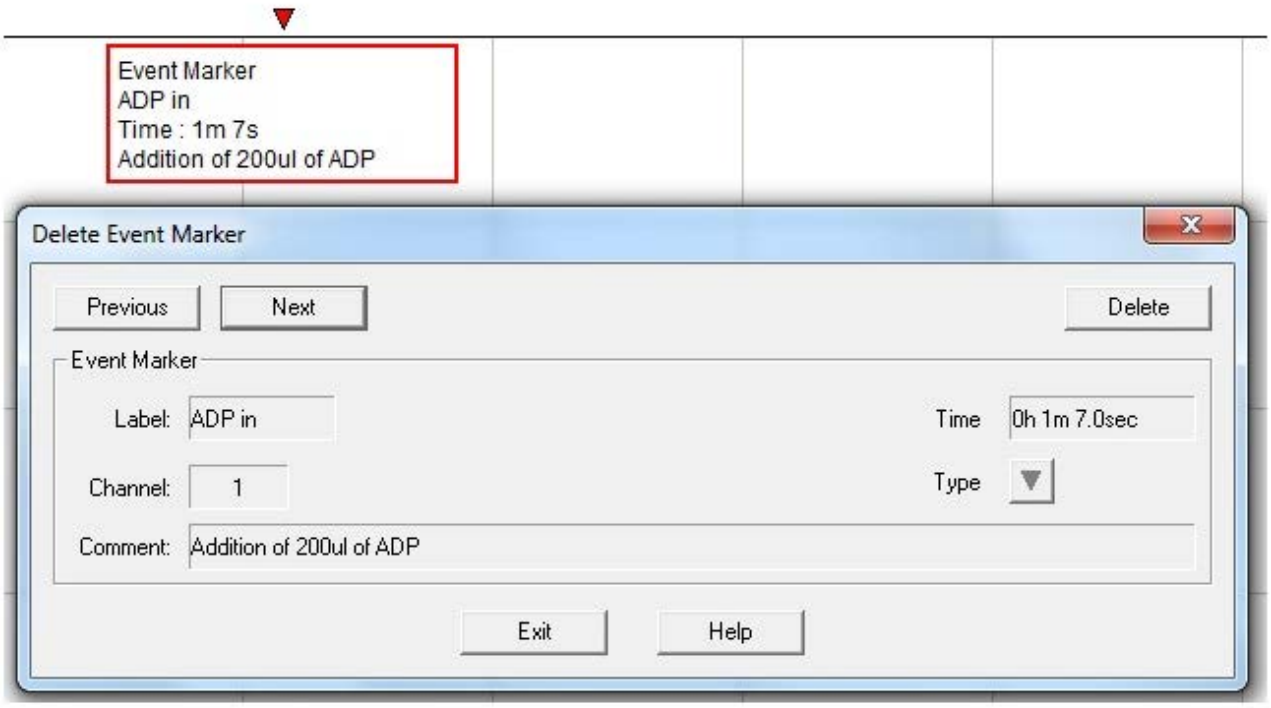
Deleting Event Marks

To delete an event mark, select Tools > Delete Event Marks from the menu bar. The following window is displayed:



The event to be edited is displayed in a red box along with the information associated with it. Select the relevant event mark using the previous and next buttons and press delete to remove the event mark from the graph.

File Information

Experiments recorded in the OxyTrace+ software can be saved along with an information set giving details of the following:

- File name and path
- Date of experiment
- Start time of experiment
- Data acquisition rate used in the experiment
- OxyTrace+ version information
- Details of what traces were recorded (i.e. number of channels, auxiliary data
- A text description of the experiment

File information is viewed from the following window:

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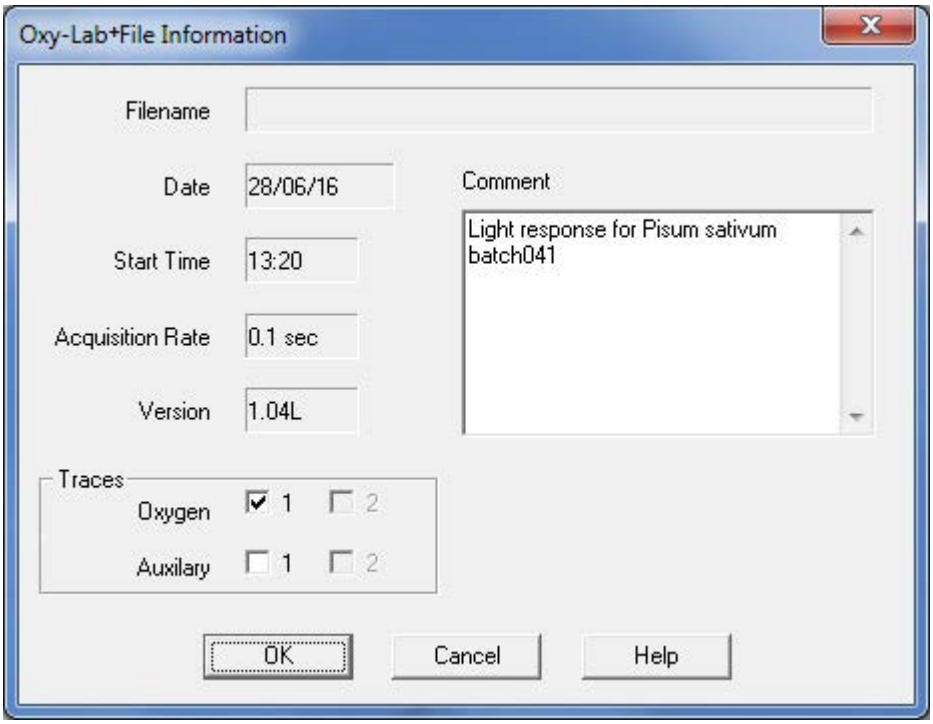

This window is accessed either by selecting View > File Information from the menu bar or by clicking the icon on the tool bar. A text description is added by typing the relevant comments into the Comments text box. The File Information dialogue is also initially displayed when opening previously saved measurement data.

Rate Measurements

Live Rate

During measurement, the oxygen signal is continuously monitored by OxyTrace+ software for the rate of change in oxygen tension. A Live Rate measurement is displayed in the Data Bar giving an indication of the rate at which oxygen is evolving or being consumed by the sample.
The Live Rate is calculated by performing a least squares regression over a user-defined duration of measurement. The duration is defined in the Setup Live Rate Display window which is accessed by selecting Rate > Setup Live Rate Display from the menu bar.

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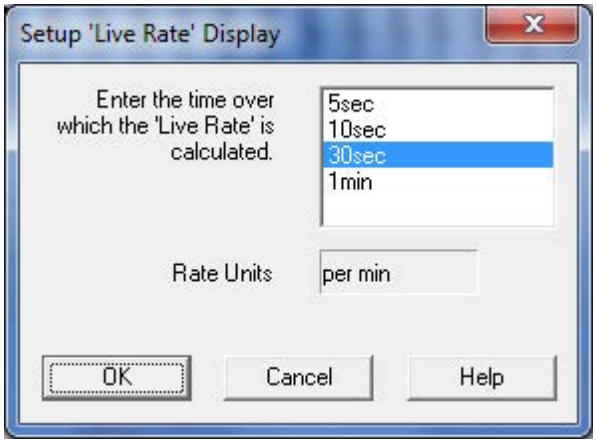

The rate is continuously re-calculated over the time period defined and displayed in the data bar in nmol/min for liquid-phase and µmol RTA/min for gas-phase measurements. The longer the time duration selected, the more stable the rate measurement will be. In addition, Live Rate data may also be plotted to the OxyTrace+ axes for graphical representation. Please refer to the Plotting Live Rate Data section for further information.

By selecting Tools > Graph Data plus Rate Data or the icon from the tool bar, both oxygen signal data and associated Live Rate data are plotted on a horizontally split screen with oxygen in the lower section and Live Rate data in the upper section as shown in the diagram below.

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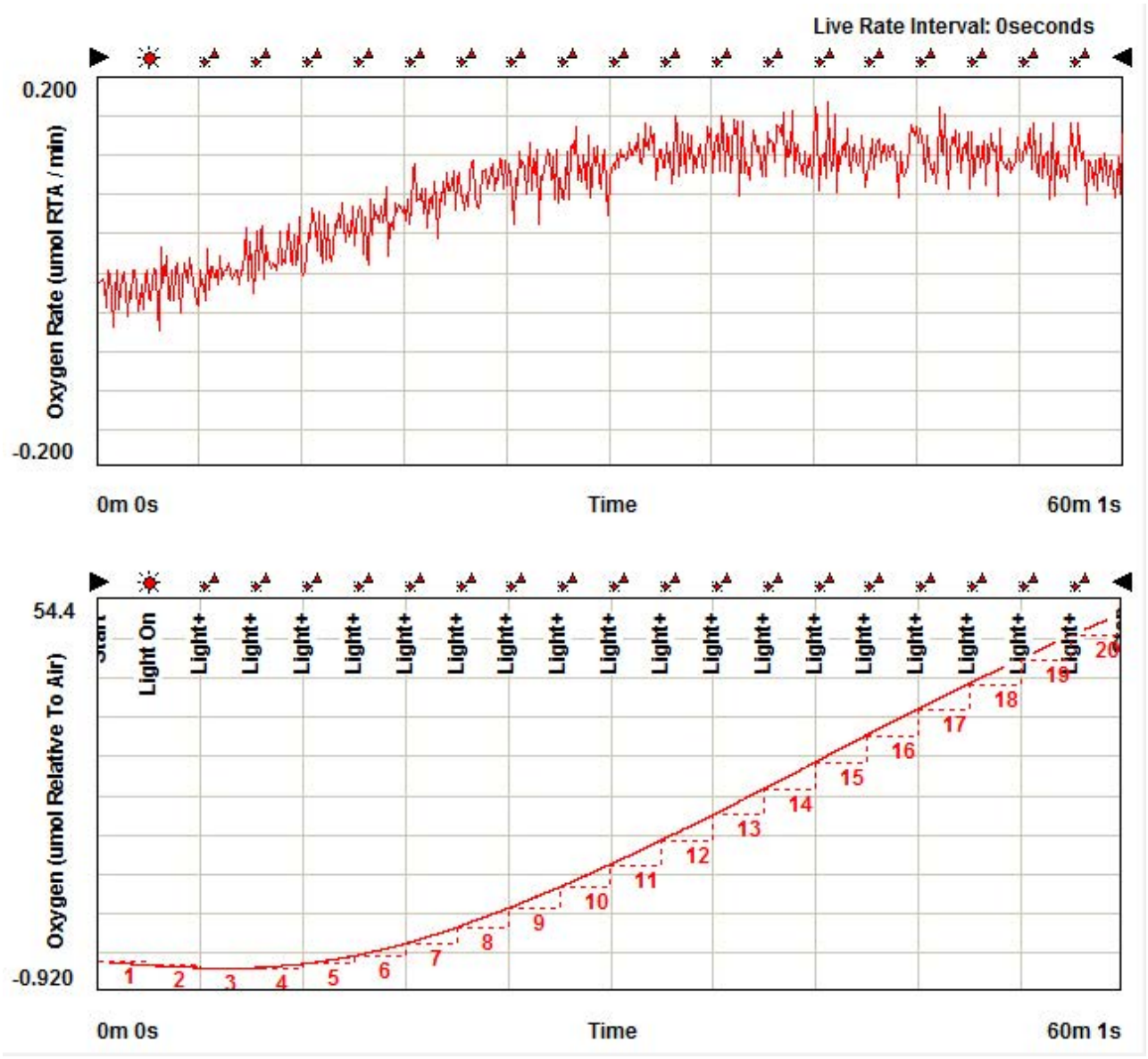

In this data display mode, additional axes scaling options are included in the Zoom XY axes settings dialogue allowing the Live Rate axes to be scaled as required.

Manual Rate Measurements

OxyTrace+ allows rate of change of oxygen tension to be measured on recorded data from the electrode disc using the Rate Measurement Function.

Rate measurements are made by either "clicking and dragging" a pair of Rate cursors to the required positions or by manually entering start and end times on the recorded oxygen signal in order to define the rate interval. The 2 cursors appears as vertical lines on the graph screen. The

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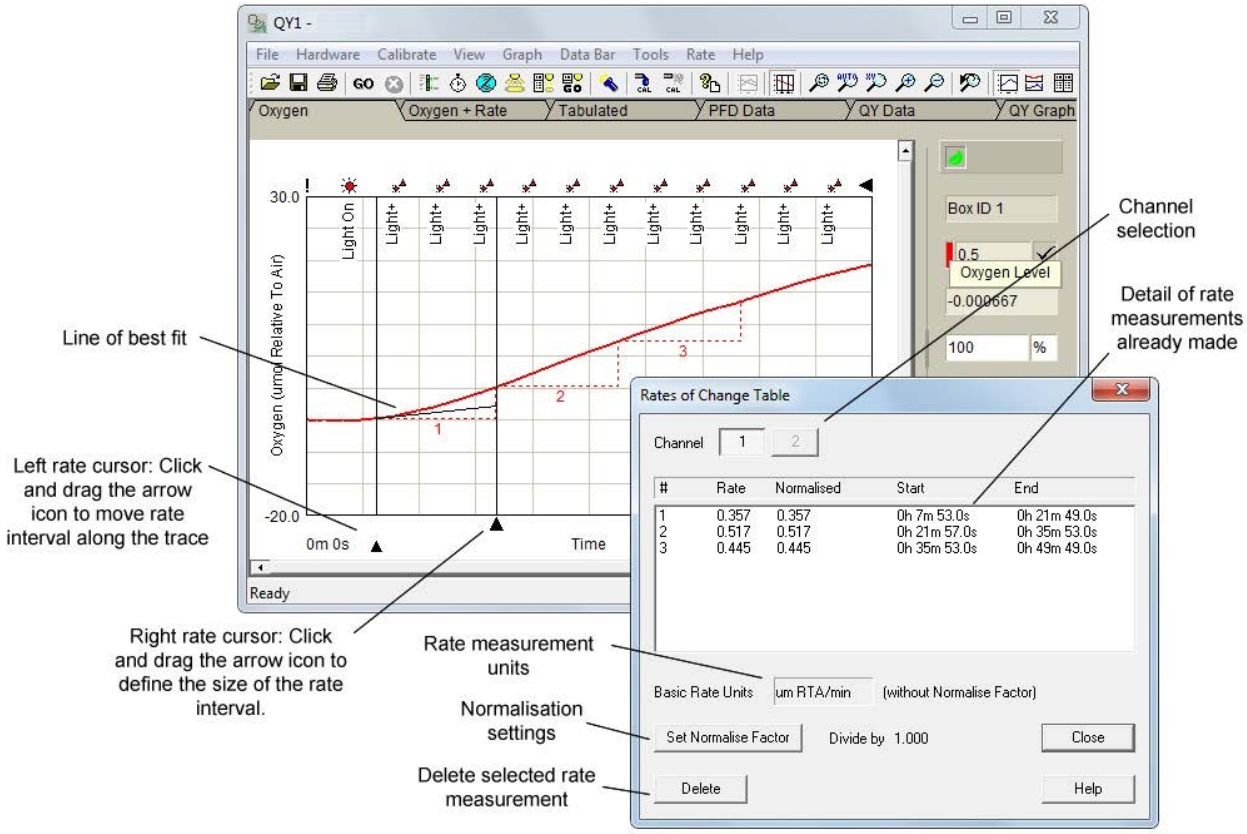
left rate cursor is used to move the rate interval along the trace whereas the right rate cursor is used to set the rate interval itself.



The rate cursors may be activated either by selecting Rate > Rate Cursors from the menu bar or by clicking the icon on the tool bar. The 2 vertical cursors will be displayed on the screen along with the Rate Table. To manually set the rate interval, select Rate > Enter Rate Cursor Times from the menu bar. A window is generated allowing start and end times to be set as shown in the diagram below.

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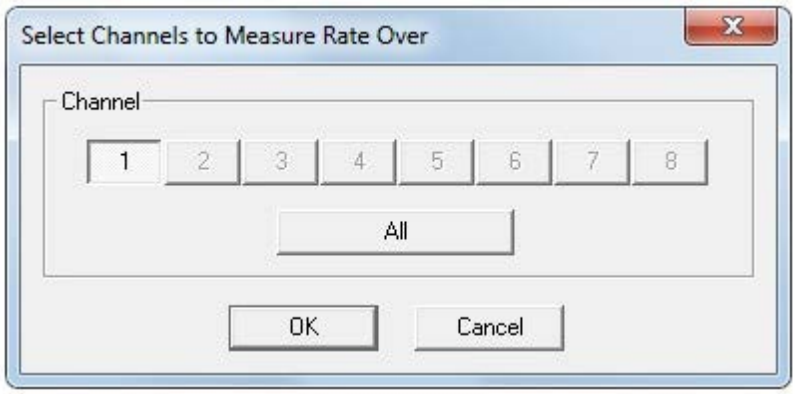
In a multi-channel system, rates may be measured either on all traces simultaneously or on individual channel traces. Once the rate cursors have been activated, select Rate > Select Channels from the menu bar. A window is displayed allowing either all traces or individual channels to be included in the rate measurement.



This option may also be selected from the Rate Table itself.

When the rate interval cursors are moved along the trace, OxyTrace+ software automatically draws a line of best fit between the 2 cursors. The line of best fit is calculated by least squares regression and can be displayed or hidden from the Rate > Setup Line of Best Fit option in the menu bar.

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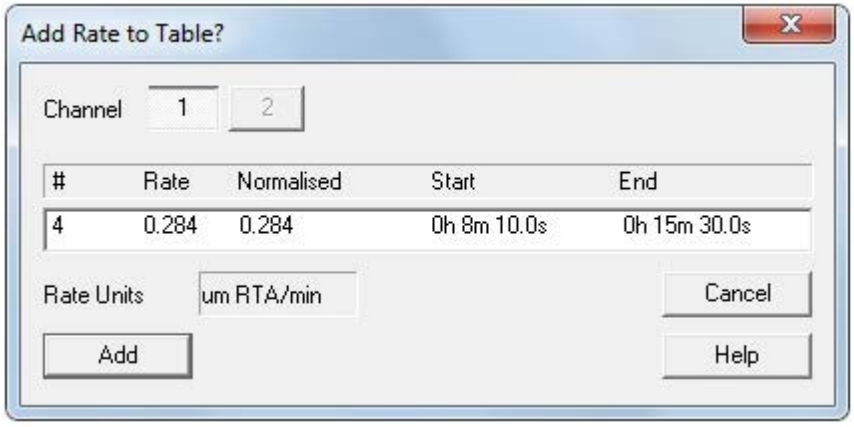
Line of Best Fit can be configured to calculate along individual channels or to be toggled between on and off (show/hide).

Once the desired rate interval and position has been defined, the rate can then be entered into the rate table by either selecting Rate > Add Rate to Table from the menu bar. Confirmation of the rate will be displayed before addition to the main rate table as shown in the image below:



Once the information displayed in the window above has been checked, it may be added to the main rate table simply by clicking the Add button.
Once added to the main rate table, rate data can be normalised if required. The normalise window allows a factor to be entered and also if the normalisation factor is to be multiplied or divided by.

In addition, the horizontal and vertical intersecting lines used to calculate the rate measurement are displayed on the graph once the rate has been added to the table. These graphics appear with labels which may be defined to represent the rate measurement number or the rate value. The labels can be configured to show rate measurement reference numbers to correspond with the measurements shown in the rate table or the actual rate value itself. The rate measurement labels are indicated on the trace by a series of horizontal and vertical lines above or below the trace showing how the slope has been calculated. These lines may be shown as coloured (to match the

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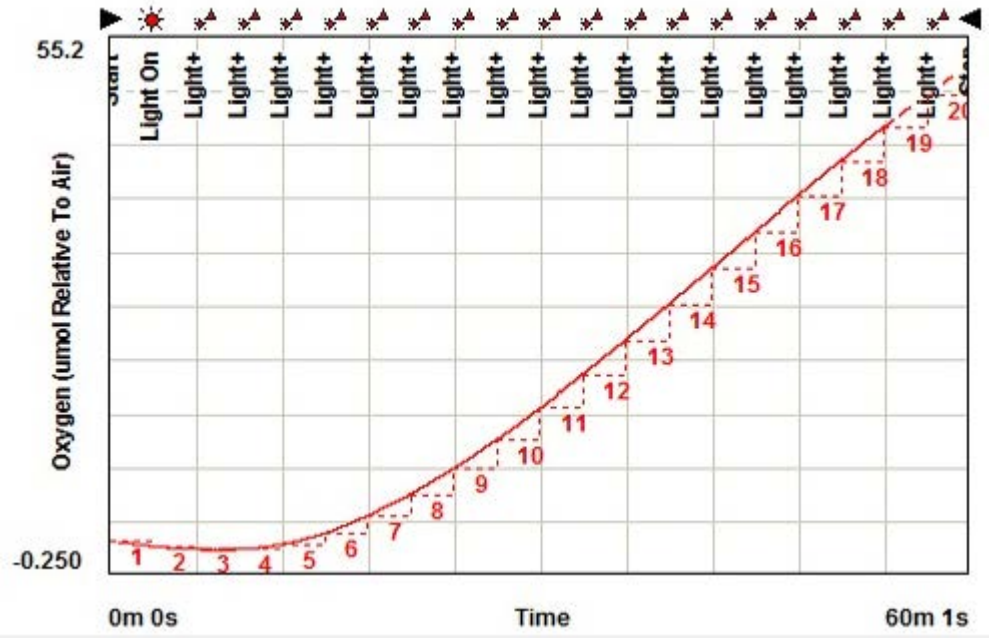
trace colour) or black. These options are accessed by selecting Tools > Options from the software menu bar.

The rates are displayed on the graph screen as shown in the diagram below:



Any measured rates are saved along with the file and are also included in the print out.

Tabulated Data

Once the experiment is complete, data can be viewed optionally in numerical format as the individual recorded data points. This data is shown either by selecting Tools > Tabulate Data from the menu bar or by clicking the icon from the tool bar. The data from all signals and channels is displayed as shown in the image below:

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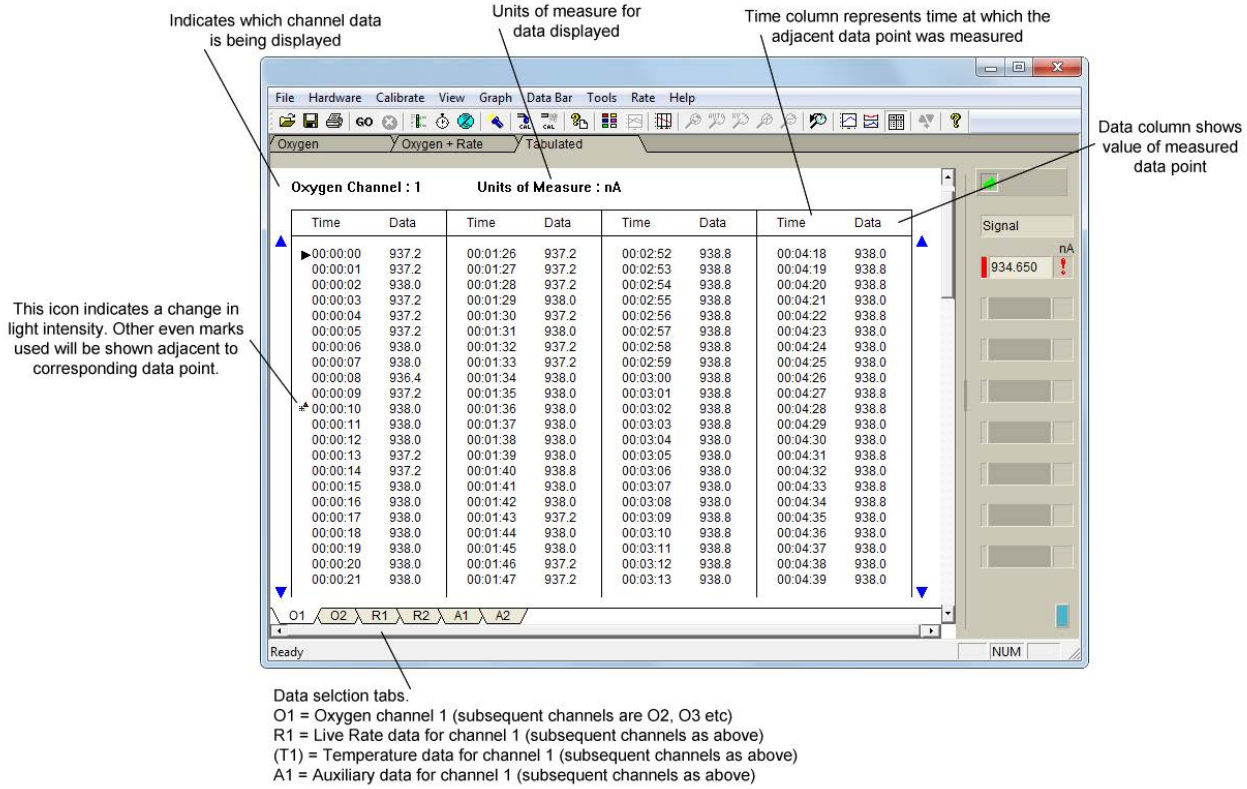

If the print function is selected whilst the tabulated data screen is displayed, the data will be printed in tabulated form.

Exporting Data to Other Software Packages

Data recorded in OxyTrace+ is saved as a *.CSV (Comma Separated Values) file. This file type may be opened and reviewed directly in Microsoft Excel so that data may be subjected to further, more advanced statistical analysis.

ALL DATA SAVED IN OXYTRACE+ SOFTWARE IS SPECIFICALLY ENCRYPTED. IF ANY AMENDMENTS ARE MADE TO DATA FILES OUTSIDE OF OXYTRACE+ SOFTWARE, NOTIFICATION OF EXTERNAL CHANGES IS DISPLAYED WHEN THE FILE IS RELOADED INTO THE SOFTWARE. THIS NOTIFICATION IS ALSO DISPLAYED ON PRINTED EXPERIMENTS.

Printing Data

OxyTrace+ allows a comprehensive print out of completed experiments. Information including data acquisition rates, gain and back off settings and calibration information are included in the print out along with a printed graph, event marker and rate measurement details. If the data has

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been modified outside of the OxyTrace+ software, a warning message will be printed at the top of each page as notification.

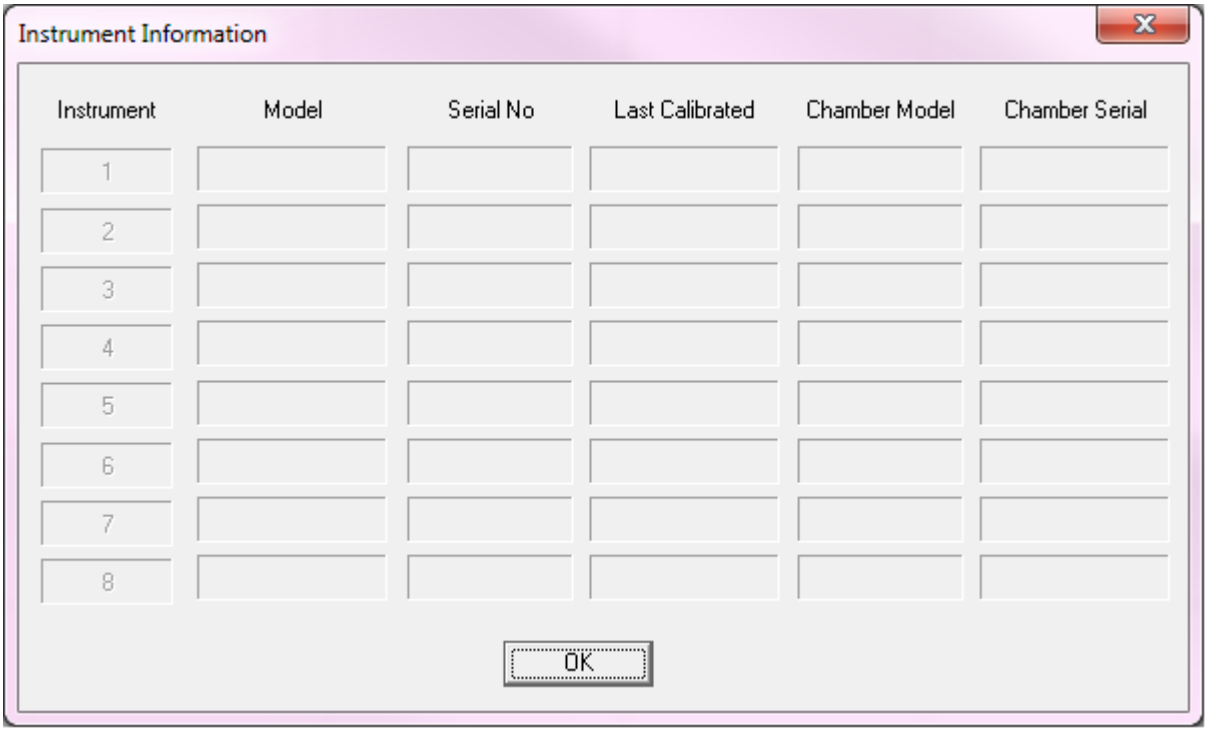
Alternatively, if only a single page printout showing the graph only may be printed by selecting the Simple Print option in the Options menu

If tabulated, numeric data is required, change the view to the tabulated data screen and select File > Print from the menu bar or click the icon on the tool bar.

Viewing Previously Saved Data

Previously saved files may be viewed at any time using the File > Open command from the menu bar or the icon from the toolbar. Alternatively, OxyTrace+ can be run in View Mode which allows the software to function as a stand alone operation when a control unit is not detected however, some operational functions will be disabled.

To operate OxyTrace+ in view mode when a control unit is not present, simply run the software as normal. The software initially scans for connected control units and when none are found, the following window is displayed.



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Pressing OK opens OxyTrace+ allowing data files from Oxylab+ , Oxygraph+ or Oxytherm+ control units (to be loaded and analysed using the integral data analysis functions. Obsolete commands and functions are disabled during view mode. These functions include:

- Recording controls
- Stirrer controls
- Gain / Back off settings

Normal function of the software may be re-established by reconnecting a powered control unit to the PC via USB cable and pressing the icon from the toolbar or by selecting Hardware > Scan for Instruments from the menu bar. This function looks for control units currently connected to the PC. Once found, the serial number, type and calibration status are displayed.

1.5.3.6 Recording an Auxiliary Signal

Connecting the Auxiliary Device

Oxygraph+, Oxytherm+ and Oxylab+ control units are able to record the signal from an optional auxiliary input. This signal could be from a range of different devices provided the device output is analogue between 0 - 4V. Example devices are:

- Thermometer
- pH electrode
- TPP+ electrode (or other ion selective electrode)
- Output from fluorimeter
- PAR

Enabling the Auxiliary Device

OxyTrace+ software is configured to accept an auxiliary input device from a window accessed by selecting Calibrate > Calibrate Auxiliary/pH/Other ISE from the menu bar. The following dialogue is generated.

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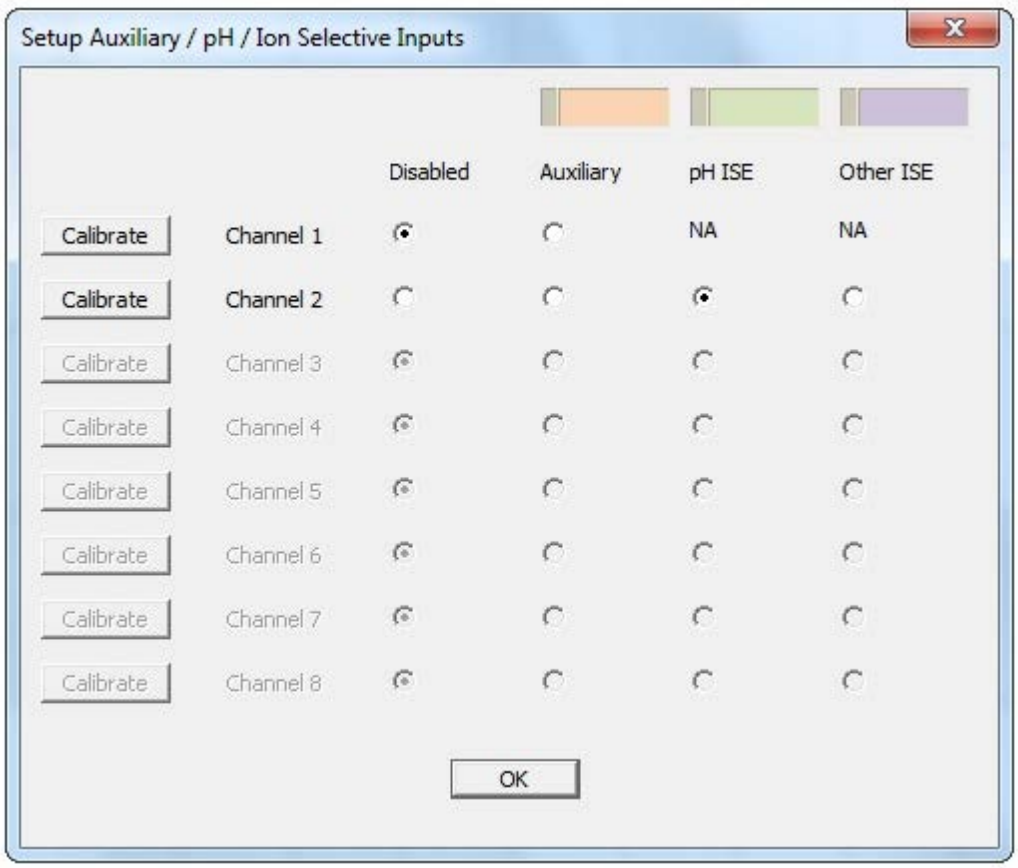

Auxiliary inputs for each active channel are enabled by checking the appropriate check box in the window shown in the diagram above. An auxiliary axis is added on the right side of the graph screen and the signal itself is plotted in the graph area as a real time chart recorder emulation. The signal is displayed in numerical format in the Data Bar (if enabled) along with the corresponding trace colour. All individual data points can be reviewed in tabulated format once the measurement has completed.

Calibrating the Auxilary Device

Auxiliary Channel Calibration

OxyTrace+ provides a 2-point calibration routine for any connected auxiliary device allowing the auxiliary signal to be recorded in actual calibrated units. The calibration routine takes mV readings at a lower and upper calibration points and calculates a calibration factor and offset accordingly.

To calibrate the auxiliary device, select Calibrate > Calibrate Auxiliary/pH ISE/other ISE from the menu bar.

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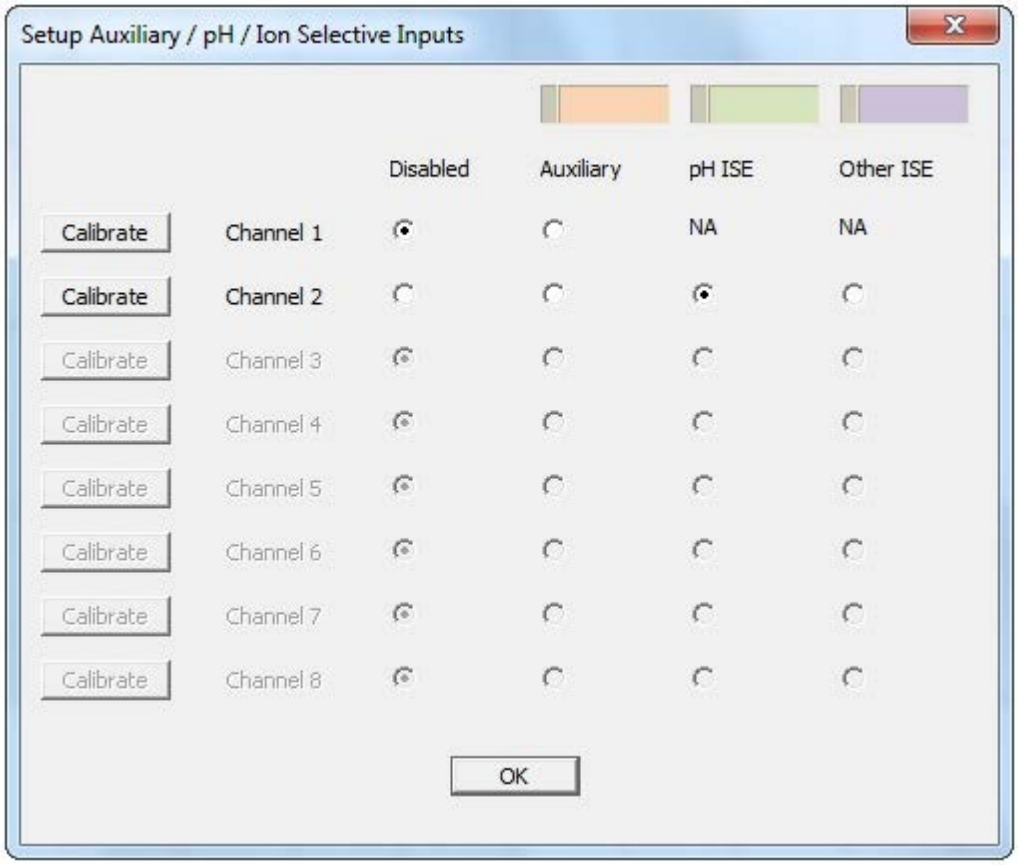
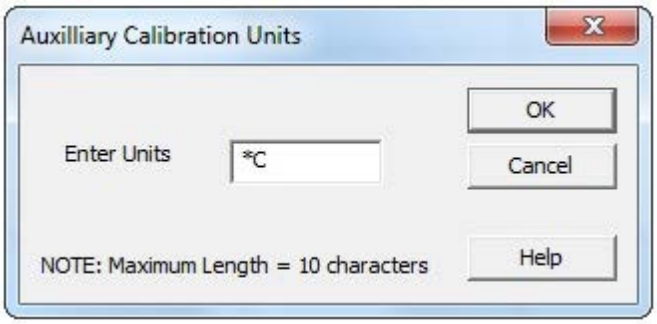


This displays all channels and any auxiliary devices that may be connected, from here you can enable which auxiliary device you want to use. Please refer to Auxiliary calibration, pH calibration and ISE calibration for more details on the calibration routines.

Auxiliary Device Calibration

Once the auxiliary channel has been enabled clicking the calibrate button will start the calibration routine.
The first stage of calibration is to enter the units required. This will be dependent on the type of auxiliary device you are using but in the example below, a temperature probe has been used



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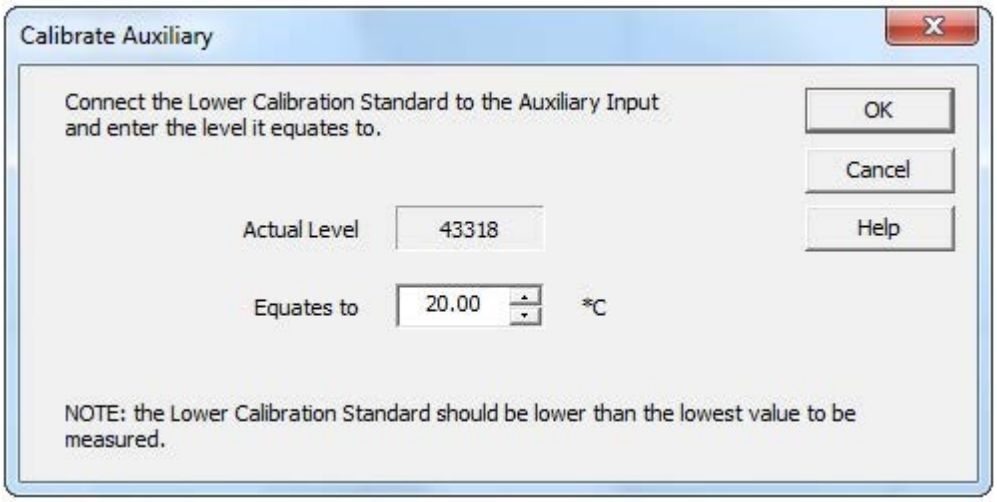
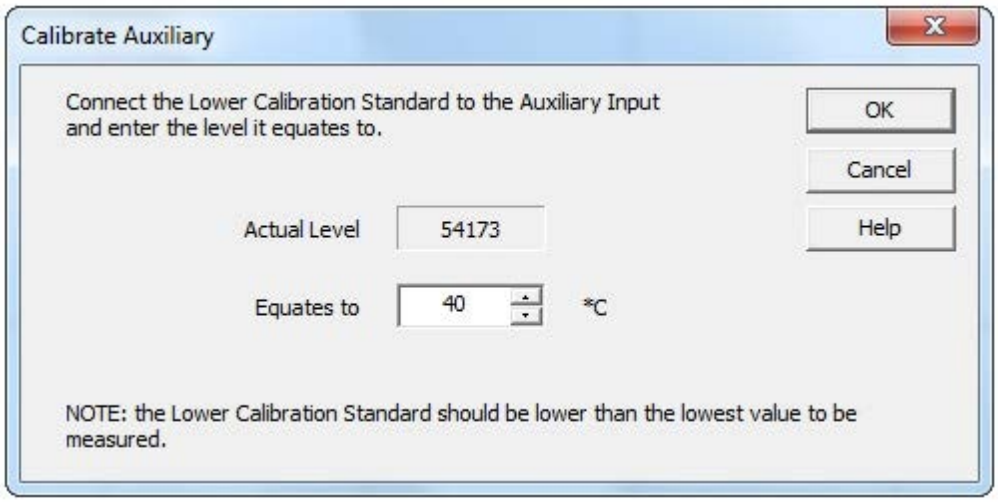
Once the required calibrated units have been entered, click OK to continue.

The next step is to assign the low calibration point. In this example, the temperature probe was situated in the chamber reaction vessel with a water bath connected to the chamber at a constant 20°C. This value should be entered into the "Equates to" spin box. Once the actual signal level has stabilized, click OK to continue.



The next step is to assign the upper calibration point. In this example,he temperature probe was situated in the chamber reaction vessel with a water bath connected to the chamber at a constant 40°C. As with the previous step, enter the calibration standard value into the "Equates to" spin box and once the actual signal level stabilizes, click OK to continue.



OxyTrace+ calculates the calibration factor and offset according to the values entered and associated auxiliary device signal levels. The calibration details are displayed in a Results dialogue at the end of the process which may be saved or discarded.

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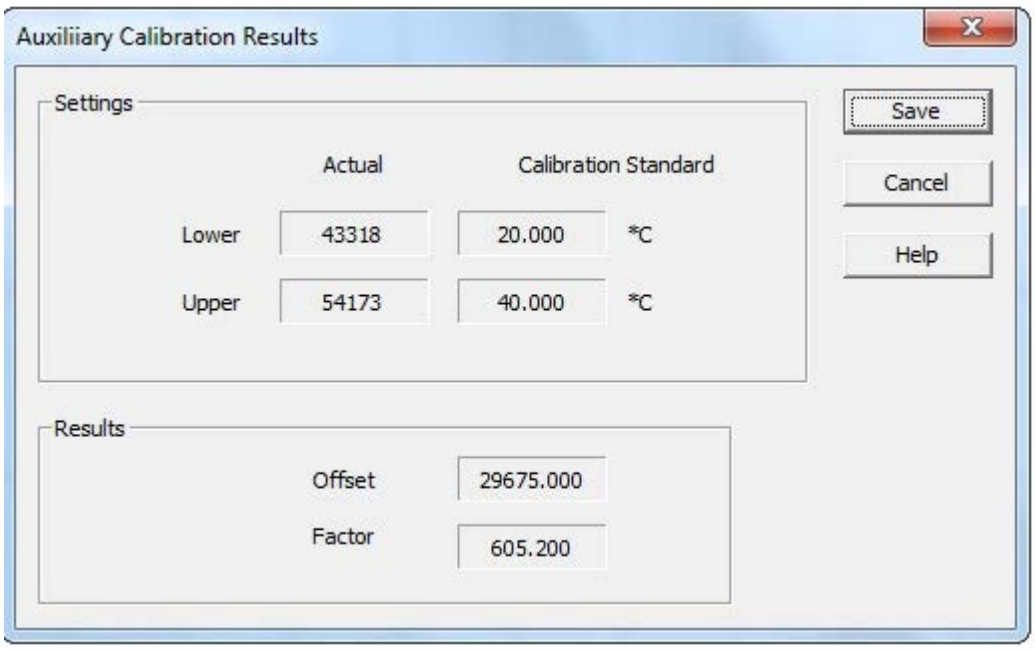

In multi-channel systems, if more than 1 auxiliary channel has been enabled, OxyTrace+ will proceed to the beginning of the auxiliary calibration routine to allow calibration of the next auxiliary channel in the system once the Save button is clicked.

pH Device Calibration

Once the pH channel has been enabled clicking the calibrate button will start the calibration routine. The first stage of calibration is to place the pH electrode into the first known pH solution. You will be prompted to do this before the software analyses the data from the electrode. Ideally the pH of this solution should be around the same as the lowest value you expect to measure.

Once the electrode has been placed into the first solution and the software has analysed the signal from it, enter the pH and the temperature of the solution into the dialogue boxes.

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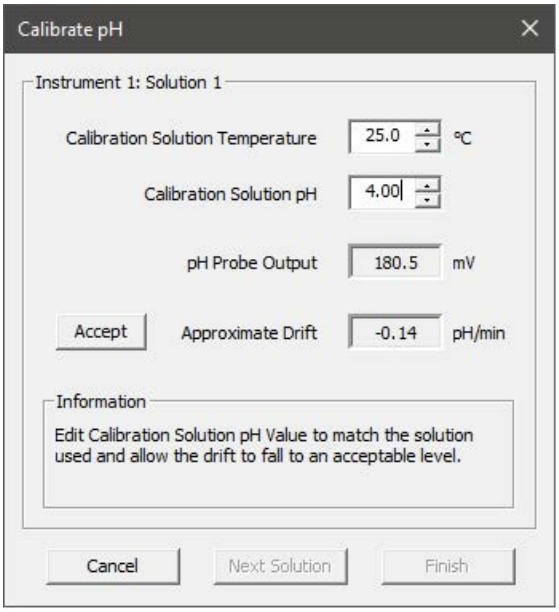

The software will wait for the signal to stabilise before making the "Next Solution" button available. However, you will be able to press the "Accept" button to the left of the "Approximate Drift" field if you consider the level of drift to be acceptable. **This will affect the accuracy of your calibration**.

After pressing "Next Solution" you will be asked if your next solution is available and then to to place the probe into that solution.

This should be of a pH in between your highest and lowest pH you expect to measure.
The software will analyse the signal again in the exact same manner as solution 1. Again, make sure the "Calibration Solution pH" field reflects the pH of the solution you are now using

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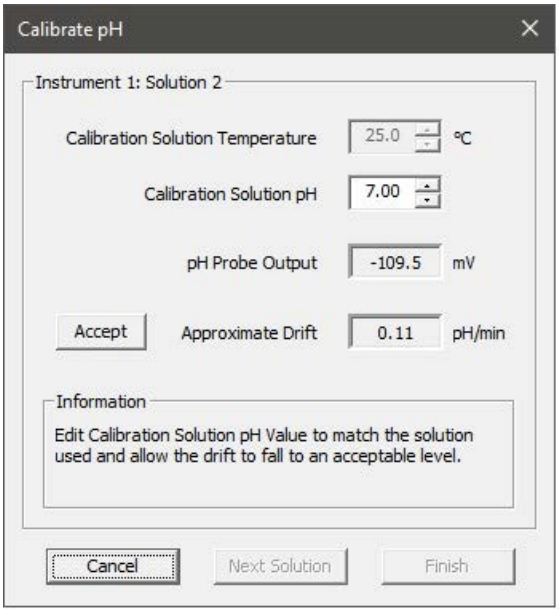
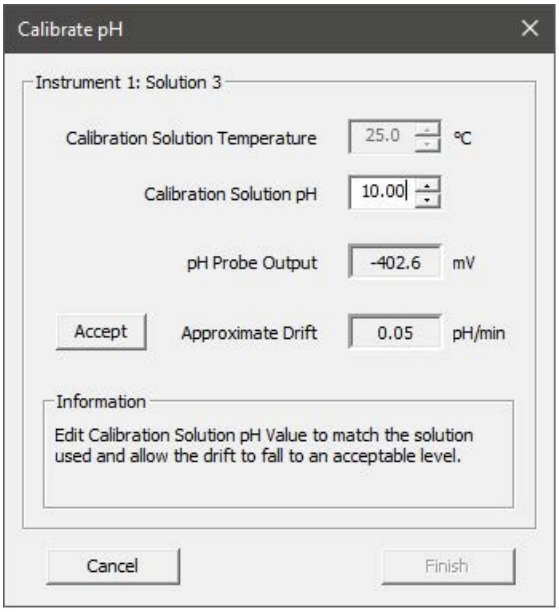


After pressing "Next Solution" you will be asked if your next solution is available and then to to place the probe into that solution.

This should have a pH around the same as the highest value you expect to measure.
The software will analyse the signal again in the same manner as solution 1 and 2. Again, make sure the "Calibration Solution pH" field reflects the pH of the solution you are now using



Press "Finish"

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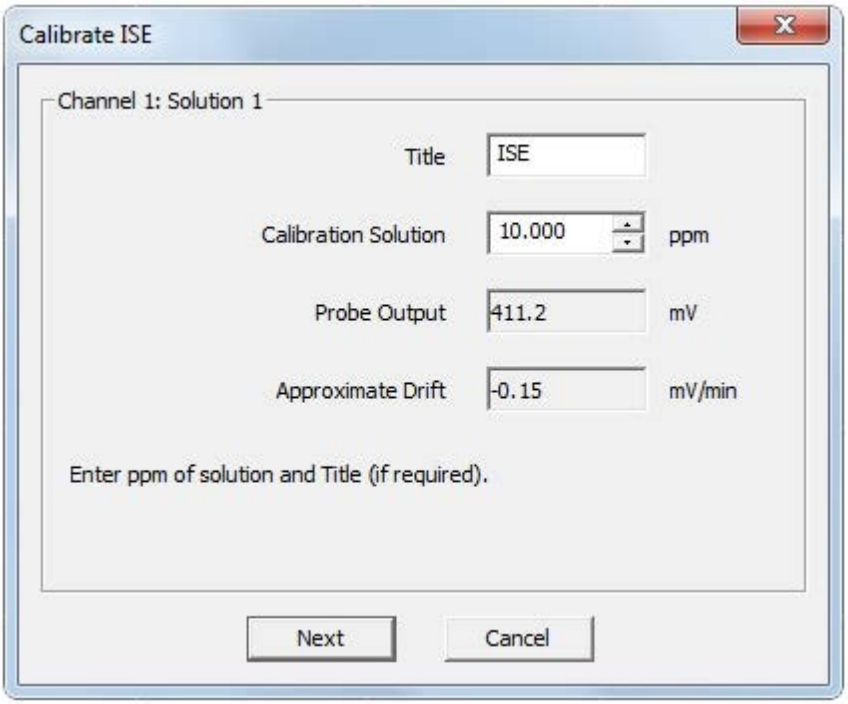
Once the calibration routine has finished the details will be saved and shown in the final dialogue box. They can also be found in the auxiliary calibration details window in the calibrate menu.

Ion Selective Electrode (ISE) Calibration

Once the ISE channel has been enabled clicking the calibrate button will start the calibration routine.

Before clicking the calibrate button make sure the ISE is in your first calibration solution as the software will analyse the signal from the ISE straight away.



On the first dialogue window you will be able to enter a title for the calibration, most commonly used is the type of ISE in use.

Once the signal has been analysed from the ISE, enter the known concentration of the solution into the calibration solution box. After this has been done press next and then place the ISE into the final calibration solution

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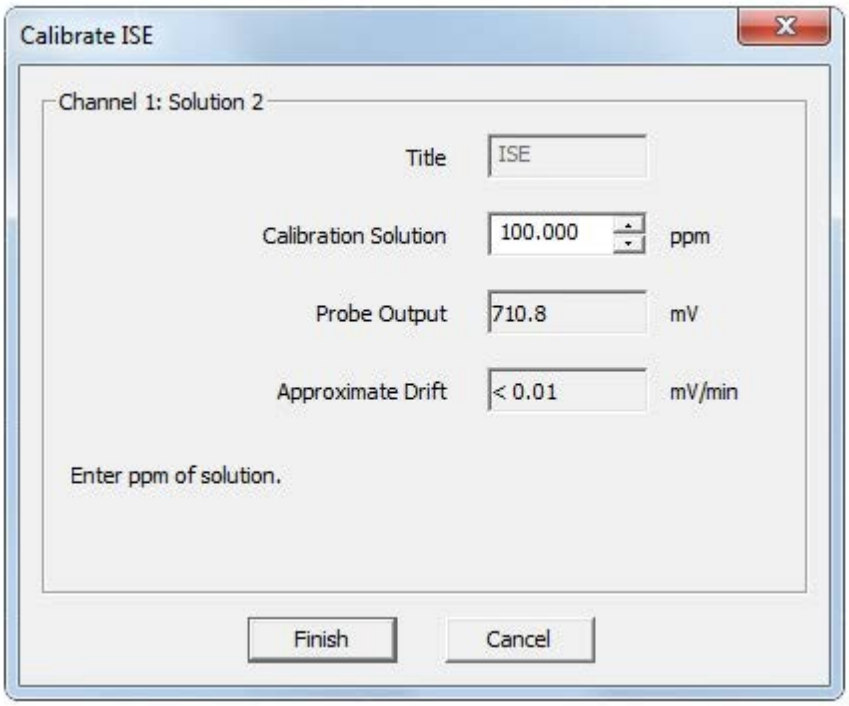

The software will analyse the signal from the ISE again, once the output has stabilised and the drift is stable enter the concentration of the calibration solution. Press the finish button to finalise the calibration.

Once the calibration routine has finished the details will be saved and shown in the final dialogue box. They can also be found in the auxiliary calibration details window found in the calibrate menu.

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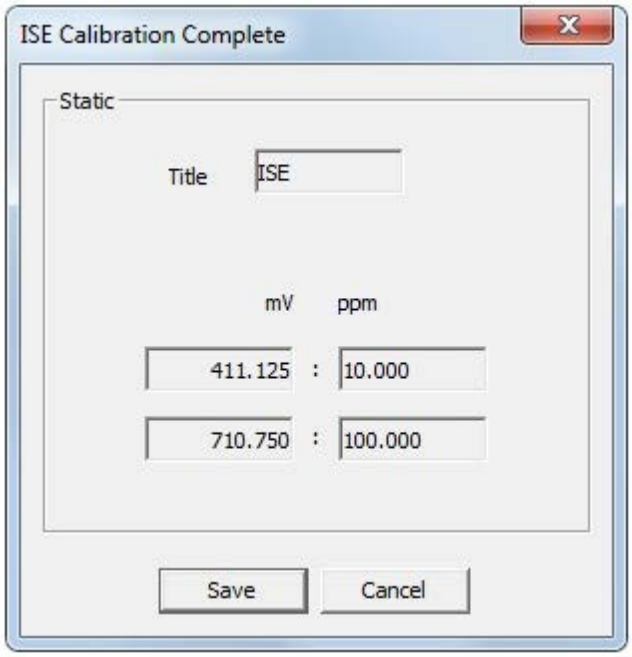
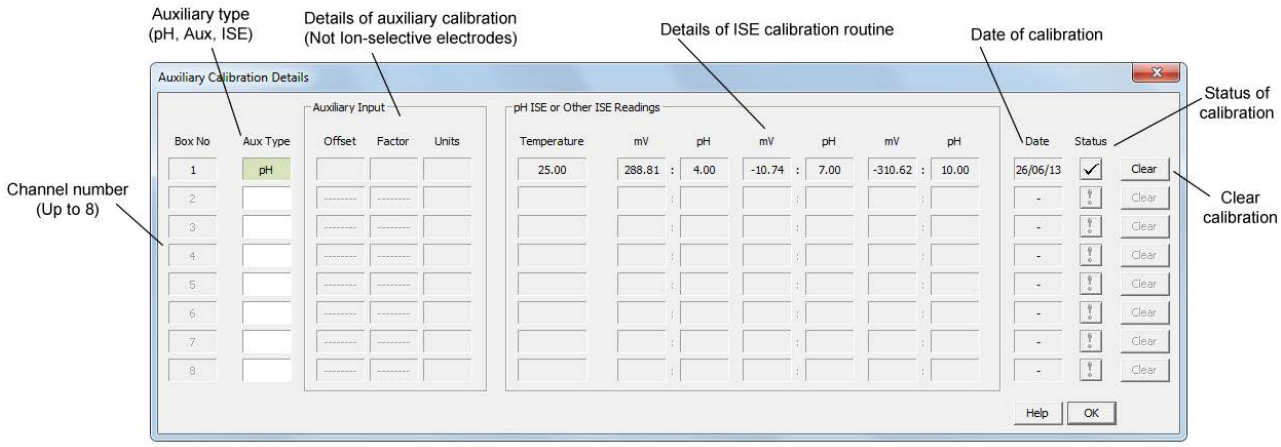


Auxiliary Device Calibration Details

To review calibration details for connected auxiliary devices, select Calibrate > Aux Calibration Details from the menu bar. Calibration details for each channel are displayed in the dialogue as shown below.



Set Calibration Warning

If the apparatus is intended for long measurement assays lasting more than a day or if the electrode is to be left prepared and in situ (please note that this should only be performed in liquid-phase electrode chambers as electrodes in gas-phase "dry out"), a calibration warning interval may be set so that after a given number of days post calibration, the user is prompted that

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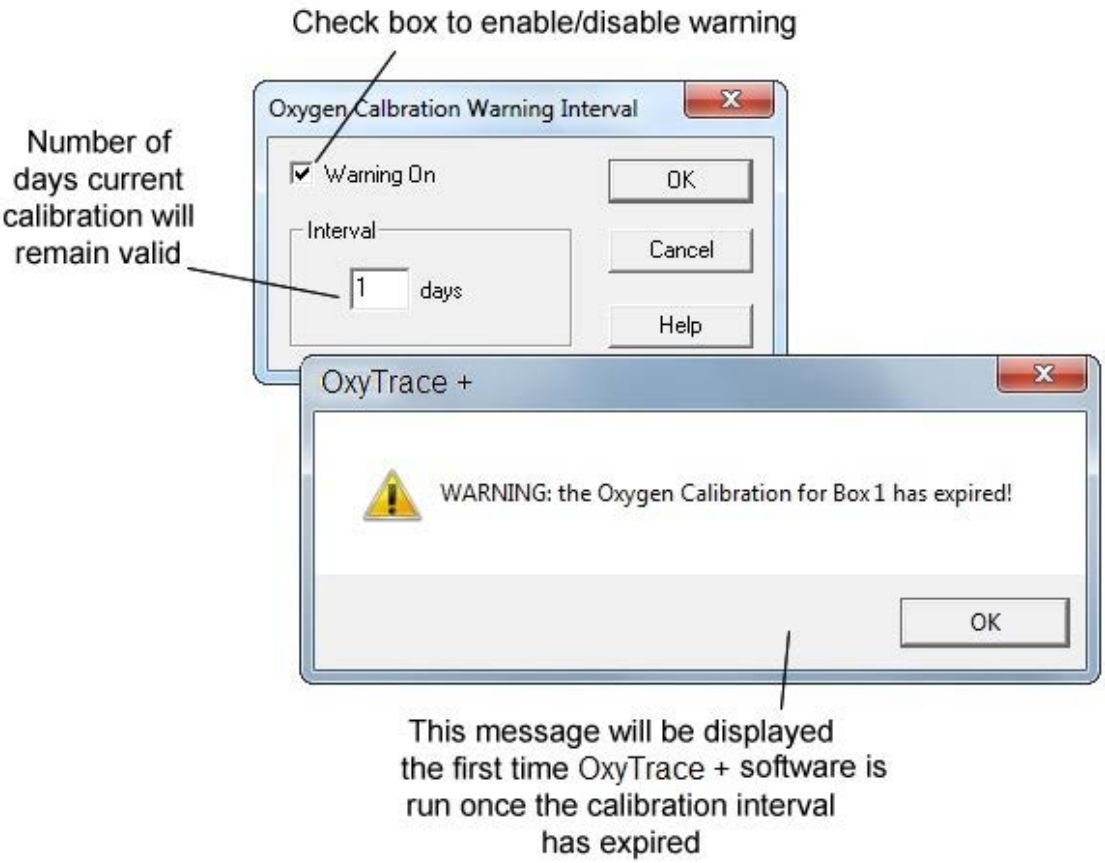
the calibration has expired when the software is initially run. A calibration warning can also be set in the same manner for any auxiliary devices connected to the instrument.



IT IS IMPORTANT TO REMEMBER THAT OVER TIME, A POLARISED DISC BUILDS UP A DEPOSIT OF BLACK SILVER OXIDE DUE TO THE NATURE OF THE ELECTROCHEMICAL REACTIONS TAKING PLACE. A DISC THAT IS LEFT MEASURING FOR MORE THAN A DAY MAY BEHAVE DIFFERENTLY AT THE END OF THE ASSAY THAN AT THE BEGINNING. PLEASE REFER TO THE SECTION LEAVING THE ELECTRODE DISC POLARISED OVERNIGHT.

1.5.3.7 Multi Channel Systems

Introduction to Multi-channel Systems

Both Oxygraph+ and Oxytherm+ control units are capable of operating either as stand alone systems or in complex multi-channel systems consisting of up to 8 control units. These control units may consist of a mixture of both types of unit linked via USB and operated by a single PC.

Oxylab+ control units may also be linked together but only up to a maximum of 2 channels.

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Each of the control units in the multi-channel system must be calibrated independently to allow for differences in electrode performance.

WHEN RUNNING A MULTI-CHANNEL SYSTEM, ALL UNITS MUST BE CALIBRATED. OXYTRACE+ SOFTWARE WILL NOT ALLOW A RECORDING FROM A SYSTEM WITH A MIX OF CALIBRATED AND UNCALIBRATED CONTROL UNITS PRESENT.

Setting Up a Multi Channel System

What Additional Hardware is Required?

It is perfectly possible to link 2 or more (depending on system type) control units together in a multi-channel setup without purchasing any additional hardware. However, there are options available which will cut the cost of multi-channel systems.

Oxylab+ based systems such as Chlorolab2+, Chlorolab 3+ and Leaflab 2+ may only be extended by one control unit to a dual channel system. Oxygraph+ and Oxytherm+ systems may be extended up to a maximum of 8 channels.

Oxygraph+ control units draw very little power. Consequently, it is possible to run up to 8 individual control units from a single power supply with the aid of the OXY/PDU Power Distribution Unit.

In addition, it would not be necessary to purchase a full set of spares and accessories with every control unit. Hansatech Instruments recommend the purchase of 1 spares and accessories kit for every 4 control units.

The most cost effective method of purchasing, for example, a system comprising 4 control units would include the following items:

- 1 x Oxygraph+ System complete

Consists of control unit, DW1/AD electrode chamber, S1 electrode disc, S1/SMB connection cable, USB Serial Cable, A2 membrane applicator, A3 top plate key and alignment jig, S2/P magnetic followers, S3 spare reaction vessels, S4 reel of membrane, S7A spare O-rings, S16 electrode cleaning kit, 12V power supply, mains cable and OxyTrace+ software.

- 3 x Additional channel Oxygraph+ control units

Including 3 x Oygraph+ control units, 3 x 12V power cable, 3 x S1/SMB electrode connection cables and USB serial cables.

- 3 x DW1/AD electrode chamber

Including 3 x S1 electrode disc.

- 1 x OXY/PDU power distribution unit.

For further information, please contact Hansatech Instruments.

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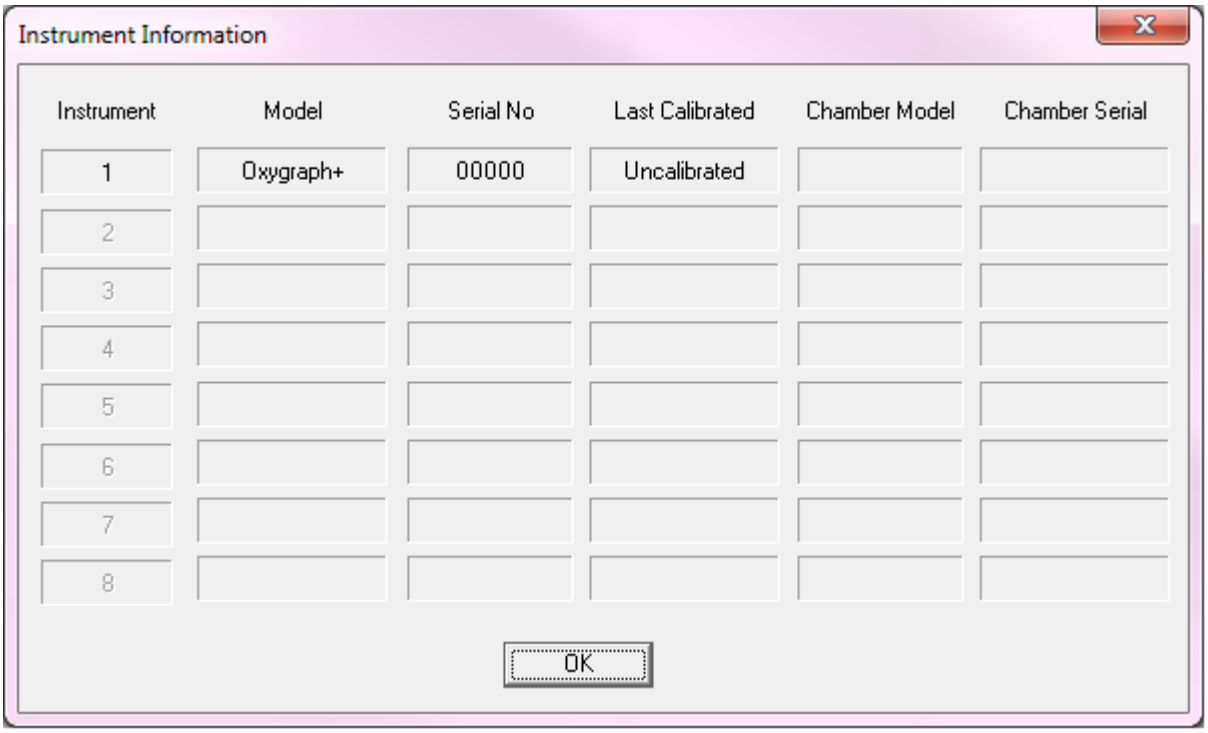
Oxytherm+ and Oxylab+ control units draw more power due to the Peltier electrode chamber and light source control respectively. Therefore, it is not possible to run more than 1 control unit per power supply.

Linking Control Units Together

Multiple control units are simply linked to the PC via standard USB serial cables. A USB hub may be required depending on the number of channels you wish to run and the amount of available ports supplied with your PC. Upon opening OxyTrace+, the software will automatically detect all units connected to the PC via USB cables. You will see the dialog demonstrated below with information for each of the connected units.



If you wish to add more units whilst the software is running simply set up the unit with both power supply and USB connection to the PC and select the icon on the toolbar to initiate a scan for the added USB control units. The dialogue above will then open again including information regarding the newly added unit.

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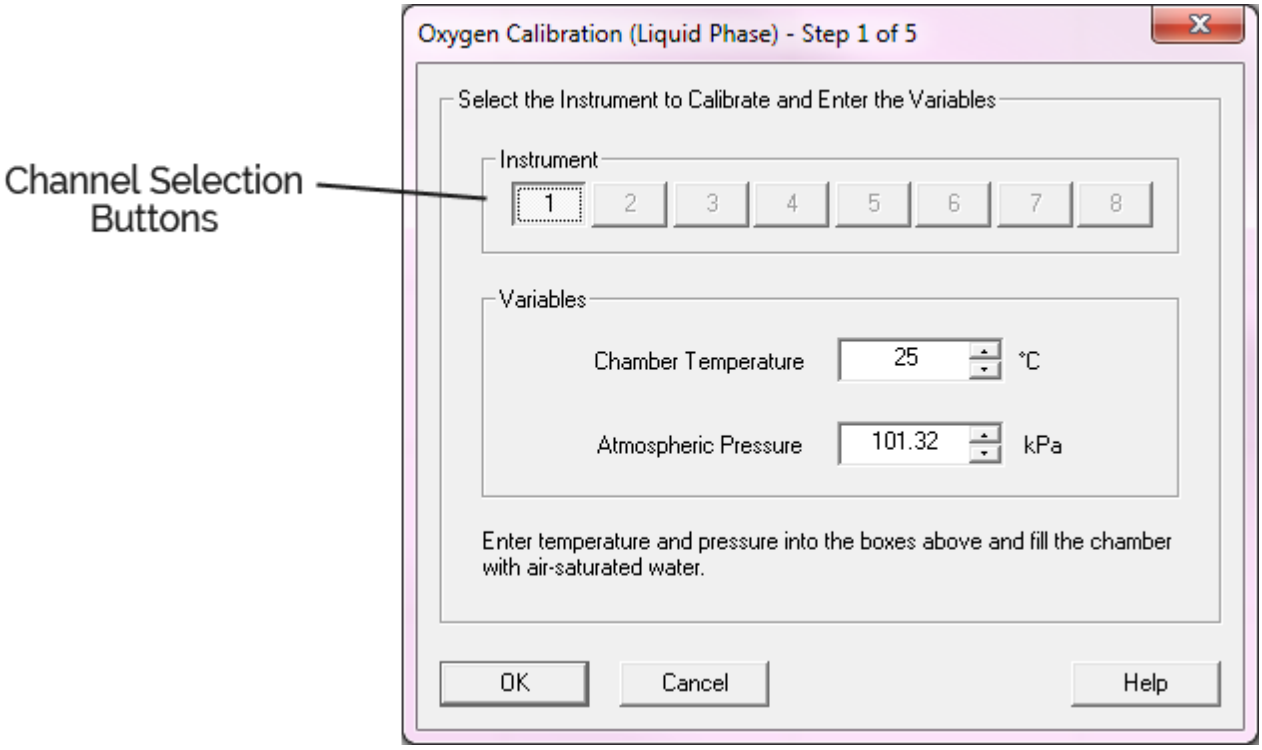
Using a Multi Channel System

Multi-channel Calibration & Configuration

All control units in a multi-channel system must be treated as individual control units and may not be simultaneously calibrated or configured. Any functions which are channel specific have the ability to select the required channel number for which actions are to be performed. For example, the diagram below shows the liquid phase calibration dialogue. Note the channel selection buttons allowing for individual control units to be selected and calibrated separately.



Multi-channel Stirring

The magnetic stirrer control dialogue allows magnetic stirrers in individual control units to be operated and configured separately or universally under a single control.

Multi Channel

If using a multi-channel system the following dialogue is generated:

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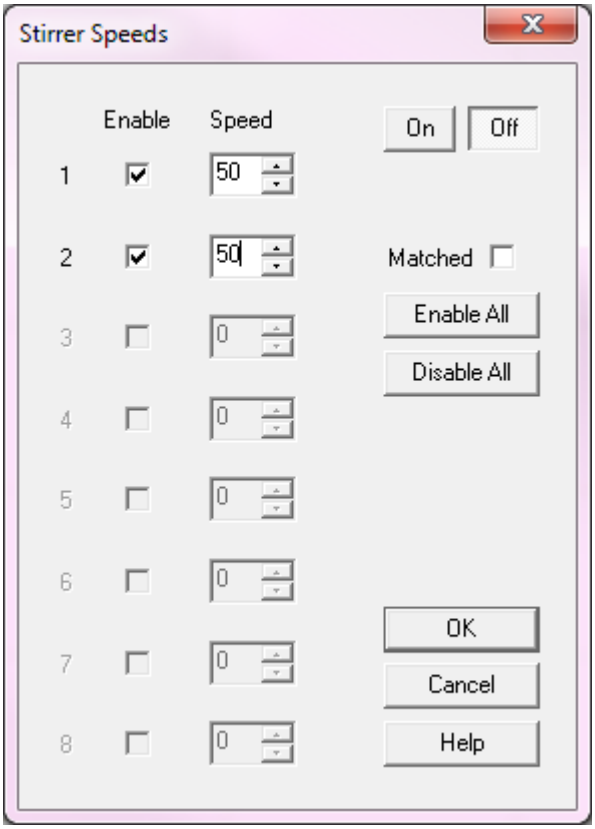

The 'Enable' check boxes allow the required stirrers to be selected individually. Alternatively 'Enable All' or 'Disable all' can be used to quickly select which stirrers are used.

The 'Matched' functions allows all stirrers to be quickly set to the same value. Pressing this will grey out the 'Speed' section for all other channels except for the first channel, this box is then used as the master speed. Any speed entered into this box will be updated for all available channels.

Once the stirrer speeds have been configured, the 'On' or 'Off' buttons will toggle the stirrers on and off. Stirrers can then also quickly be turned off/on bu pressing the icon from the toolbar.

Multi-channel Data Display

Each control unit is capable of recording and plotting 2 separate traces on the OxyTrace+ graph screen. In addition to the oxygen signal recorded from the electrode disc, an auxiliary signal of 0 - 4V may also be plotted (please click here for full details on recording auxiliary signals). This means that in a multi-channel system, there is the potential of plotting up to 16 traces per screen (2 signals from each of the maximum of 8 control units). For this reason, all traces may be customised in order to differentiate between signals according to colour and data point style from the window shown below.

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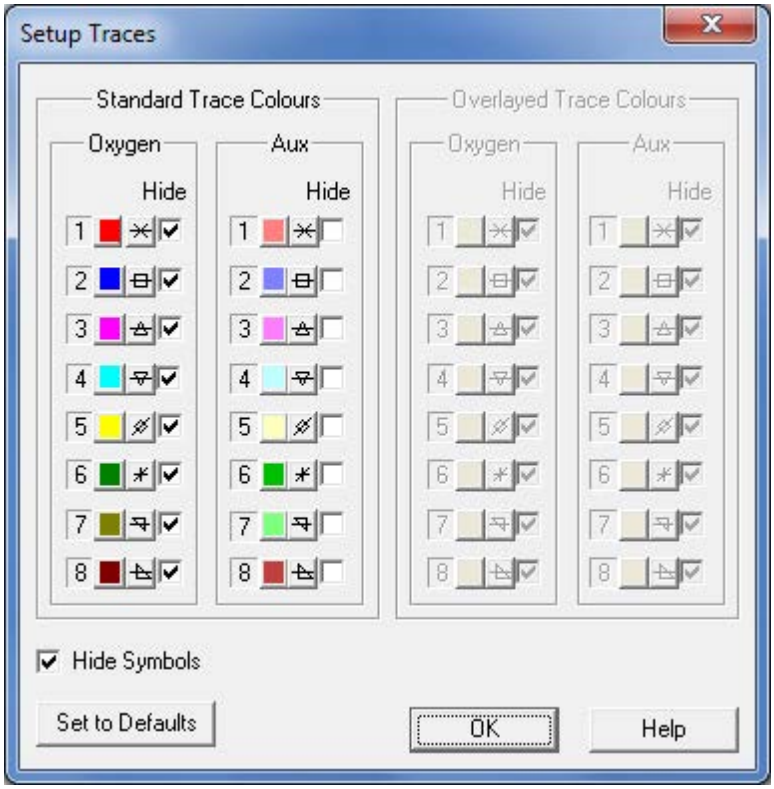

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1.5.4 LH36/2R Light Source

1.5.4.1 The LH36/2R Light Source

The LH36/2R consists of an array of 36 red LED's and is designed for use with the DW3 liquid-phase electrode chamber and LD1/2 and LD2/3 gas-phase electrode chambers. The LH36/2R requires a connection with the Oxylab electrode control unit which is responsible for the regulation of light intensity according to a user defined PFD (Photon Flux Density) table. The LH36/2R has an integral cooling fan which automatically switches on to cool the housing when the light intensity reaches a certain point.

The LH36/2R has a maximum intensity of 750 µmol m^{-2} s^{-1} in the gas-phase chambers and 900 µmol m-2 s-1 in DW3.

1.5.4.2 Light Source Calibration

Before any light response measurements are made, the light source requires calibration so that the Oxylab+ electrode control unit can set the light intensities at calibrated units of µmolsm-2s-1. Regardless of the light source in use, the calibration routine is identical. The light source is set at 10 increasing intensities between minimum and maximum output and a calibration factor is derived from the measured values at each step.

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For Chlorolab 2+ and 3+ electrode systems, the QTP1 PAR/Temperature probe may be connected to the rear of the Oxylab+ control unit allowing an automatic light calibration routine to be used. Please refer to the Mounting QTP1 in the Electrode Chamber section for details on how to setup the electrode chamber, light source and QTP1 probe for calibration.

For the Leaflab 2+ electrode system, the QSRED cannot be directly interfaced with the Oxylab+ control unit so the manual light calibration routine should be used.

To setup the QSRED, LD2/3 electrode chamber and LH36/2R light source ready for calibration, please follow the steps below referring to the diagram below:

- Place the LH36/2R on the bench, fan side down.
- Remove the upper section of the LD2/3 electrode chamber and place this on to the LH36/2R light source with the upper window interfacing with the light source.
- Place the large area sensor of the QSRED onto the lower window of the LD2/3 upper section. This should be facing upwards.
- Follow the instructions below for manual calibration of the light source using QSRED.

Both Automatic and Manual light calibration routines can be initiated from directly within the PFD Setup Table or from the Calibrate menu.

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Before selecting the calibration type, ensure that the PAR sensor is positioned optimally for the light source in use.

Automatic Calibration

The Oxylab control unit will automatically step through the 10 light settings and record the measured PAR value for each step. At the end of the 10 steps, O2view will calculate the calibration factor.

IN ORDER TO CALIBRATE TWO LED1 LIGHT SOURCES IT IS NOT POSSIBLE TO USE THE AUTOMATIC CALIBRATION ROUTINE. TO CALIBRATE TWO LED1 LIGHT SOURCES A MANUAL CALIBRATION ROUTINE MUST BE USED; AT EACH LIGHT STEP THE QTP1 MUST BE TURNED TO FACE EACH OF THE LIGHT SOURCES IN TURN AND THE INTENSITY ADDED BEFORE ENTERING THE VALUES IN THE SOFTWARE.

Manual Calibration

Oxylab will set the light source intensity at the lowest level and prompt for a PAR value from an external PAR sensor. When this value has been entered, the light source will be set to the next intensity step. This process repeats until 10 light intensity steps between minimum and maximum intensity have been recorded. At the end of the 10 steps, OxyTrace+ will calculate the calibration factor.

Viewing Light Calibration Details

Light source calibration details may be viewed by selecting Calibrate > Light Calibration Details from the menu bar. The dialogue that is generated indicates calibration status (calibrated, calibration expired or uncalibrated), maximum light intensity available and date of calibration.

Set Light Calibration Warning Interval

This function allows the light source calibration to be assigned a validity period in weeks. Once the validity period expires, the light source calibration status changes to calibration expired and a warning message will be generated by OxyTrace+ prompting a light source re-calibration recommendation.

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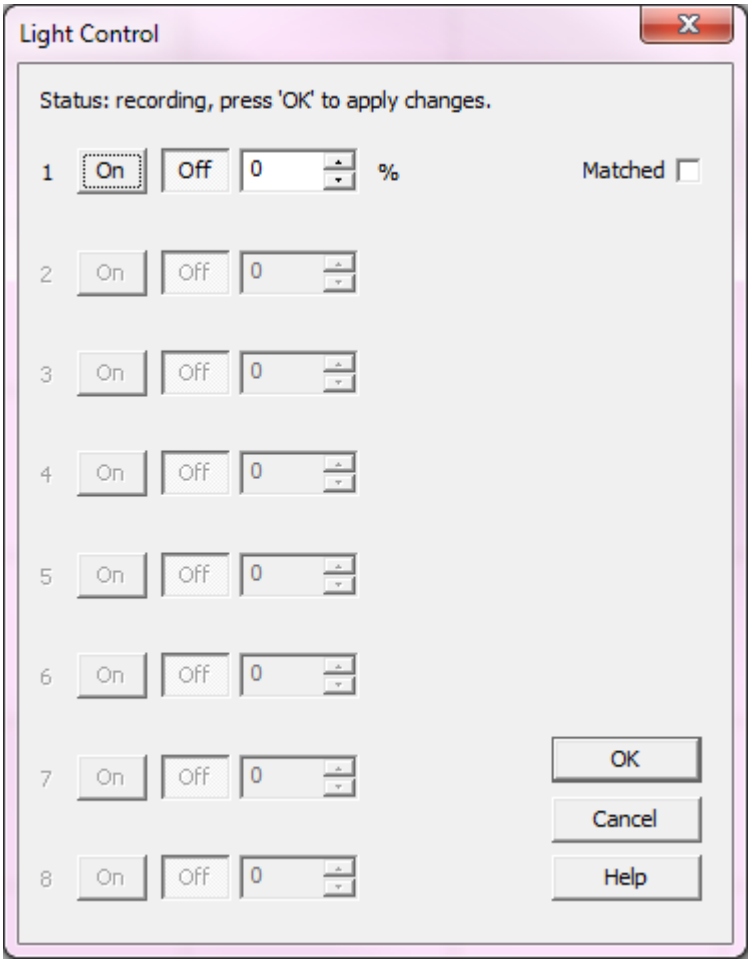
1.5.4.3 Manual Control of Light Source

The light source, when connected to the Oxylab+ control unit, may be controlled either automatically via the PFD table function or manually. To open the Manual Light Source Control dialogue, select Hardware > Actinic Light Control from the menu bar or the icon from the tool bar.

The Light Control dialogue allows light sources on either 1 or both Oxylab+ electrode control unit channels to be set to a given level of intensity of between 0-100% of the available range of intensity when light source is uncalibrated or in calibrated units of µmols m^{-2} s^{-1}. Please refer to the Light Source Calibration section for further information on how to calibrate the light source.



In a Multi-channel system, by checking the 'Matched' option, all connected systems are updated with the same light intensity. Channel 1 becomes the master box and any value entered here will be applied to all fields.

Channels can be operated separately by using the 'On' and 'Off' buttons next to each channel.

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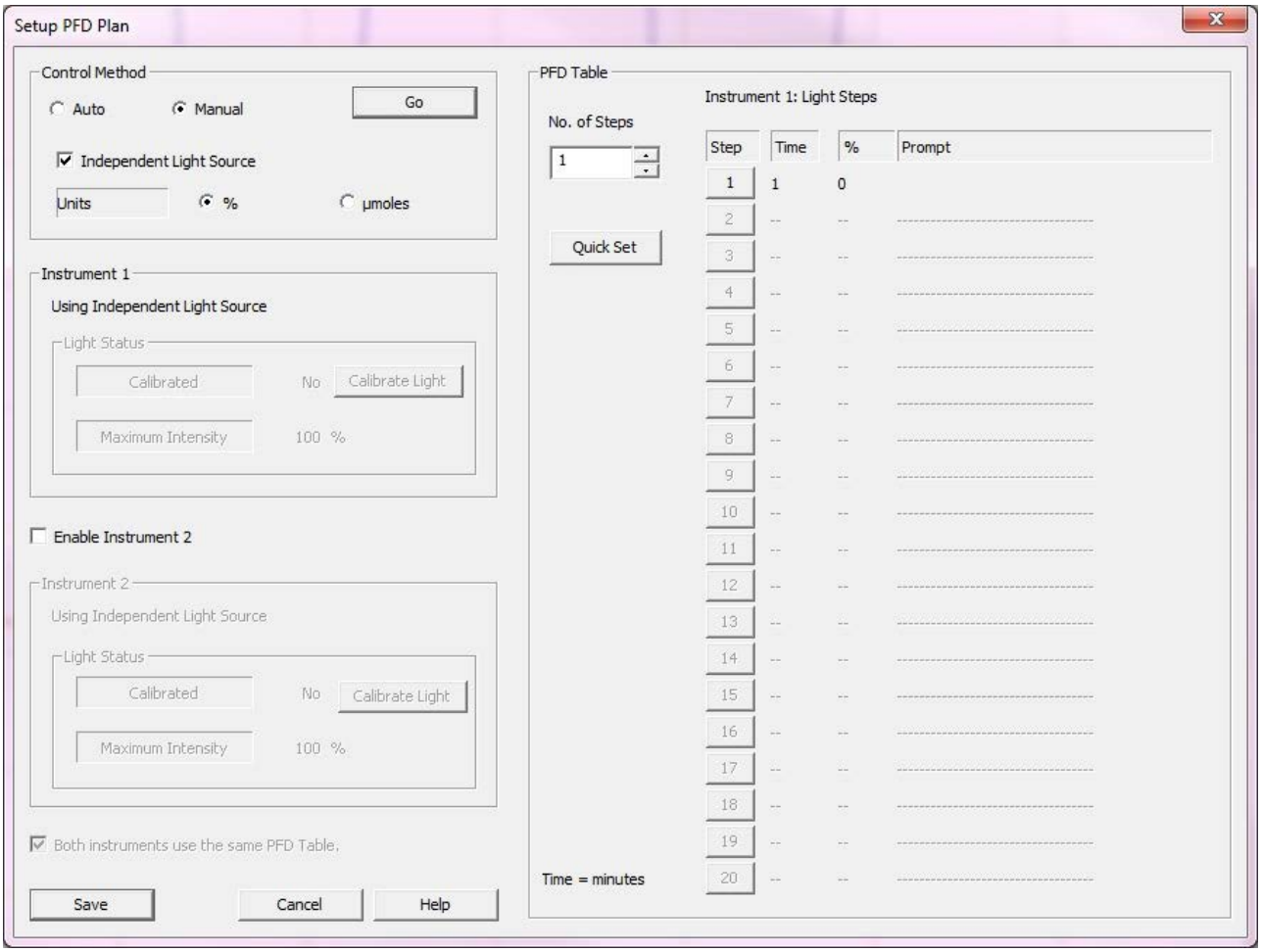
When a recording is taking place the 'OK' button will apply the changes.

Once the light sources have been configured they may also be switched on and off by clicking the icon in the Data bar.

1.5.4.4 Configuration of PFD table for Automatic Light Source Control

The PFD table allows complex light response experiments to be defined. Up to 20 individual light steps at user-definable intensity and duration may be setup with changes in light intensity activated automatically or with user intervention as required.
The PFD table is opened by selecting Hardware > Setup NEW PFD Table from the menu bar or by clicking the icon on the tool bar.



The PFD table is arranged into different areas of functionality as follows:

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- Control Method

This area of the PFD table allows the functionality of OxyTrace+ to be altered for convenience. In most cases when the either of the LED light sources are connected to the Oxylab+ control unit, the PFD table should be set to Auto. This allows OxyTrace+ to fully automate the intensity change procedure from start to finish.

Manual may be selected in situations where, for example, and addition to the sample vessel is required prior to the intensity change. In this mode, OxyTrace+ pauses the signal recording and displays a prompt on screen (defined in the Prompt section of each light step - see below). Recording will not proceed until the OK button is clicked. This functionality is also useful when light sources other than the LED light sources are used. The LS2 white light source, for example, cannot be interfaced directly to the Oxylab+ control unit and relies on manual changes in combinations of neutral density filters to alter the intensity i.e. intensity changes cannot be automated. The pause in recording allows the combinations of neutral density to be changed before recording proceeds.

- Light Status

This area of the PFD table indicates the calibration status of the connected light source. When using the LED light sources, the light calibration routine is initiated from this area. When the light source is calibrated, the maximum intensity available for use for the connected light source is also displayed here. Please refer to the Light Source Calibration section for further information on how to calibrate the LED light sources.

- PFD Table

The number of steps required can be selected using the input box. By clicking on any one of the numbered buttons, each step can be configured Individually. Alternatively, the 'Quick Set' button can then be selected which will produce the following dialogue:

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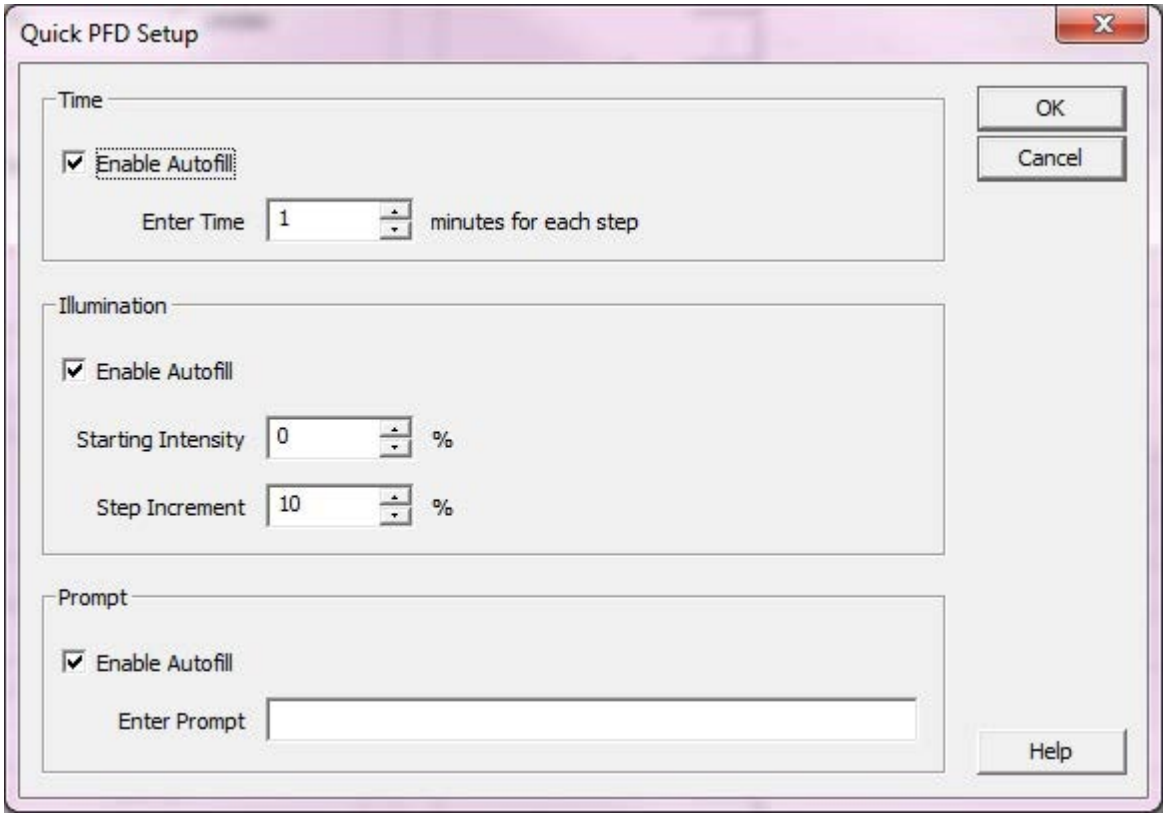

Here the time, light intensity increase and prompts can be selected ans will apply to each step.

1.6 QSRED Large Area Quantum Sensor

1.6.1 Introduction to QSRED

QSRED is a large area quantum sensor sensitive to light in the waveband approximately 550nm to 750nm (red) waveband. QSRED has been specifically supplied for the purpose of calibrating the LH36/26 in Leaflab 2 systems. The diagram below highlights the main features:

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- A: Sensor unit
- B: Control unit
- C: Analogue output
- D: Set zero potentiometer
- E: Range switch
- F: Power on/off

The measuring unit has three ranges, 0- 2000, 0- 200.0 and 0-20.00. The decimal point on the display will change with the range switch automatically.
At all times the most sensitive range possible should be selected, the unit will not be damaged by accidental over-ranging.

The instrument may require zero adjustment from time to time. This is easily accomplished by covering the top of the sensor completely to exclude all light. The scale should show '-000' with the zero sign showing intermittently. To achieve this, set the instrument to the most sensitive range, and using a small screwdriver adjust the potentiometer located just inside the small hole adjacent to the sensor plug and socket.

This adjustment will not affect full scale calibration. Extreme conditions of temperature or humidity may cause a zero drift.

The diagram below shows the quantum response of the QSRED quantum sensor.

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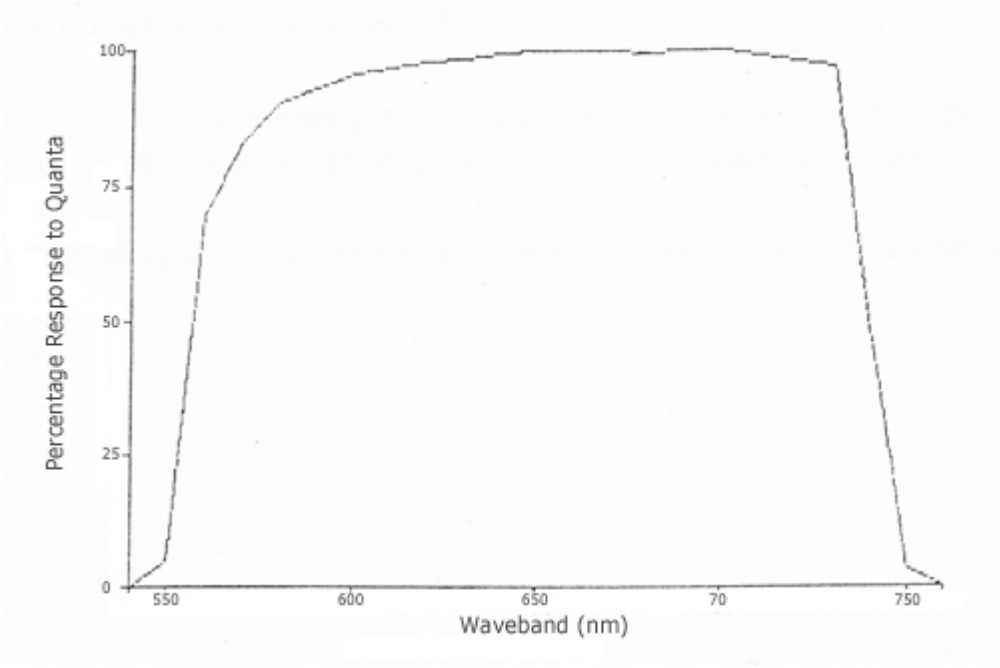

1.7 Leaflab 2 + System Troubleshooting

1.7.1 Communication Problems between Control Unit and the PC

If no control unit is found when opening OxyTrace+ the following dialogue will be displayed.

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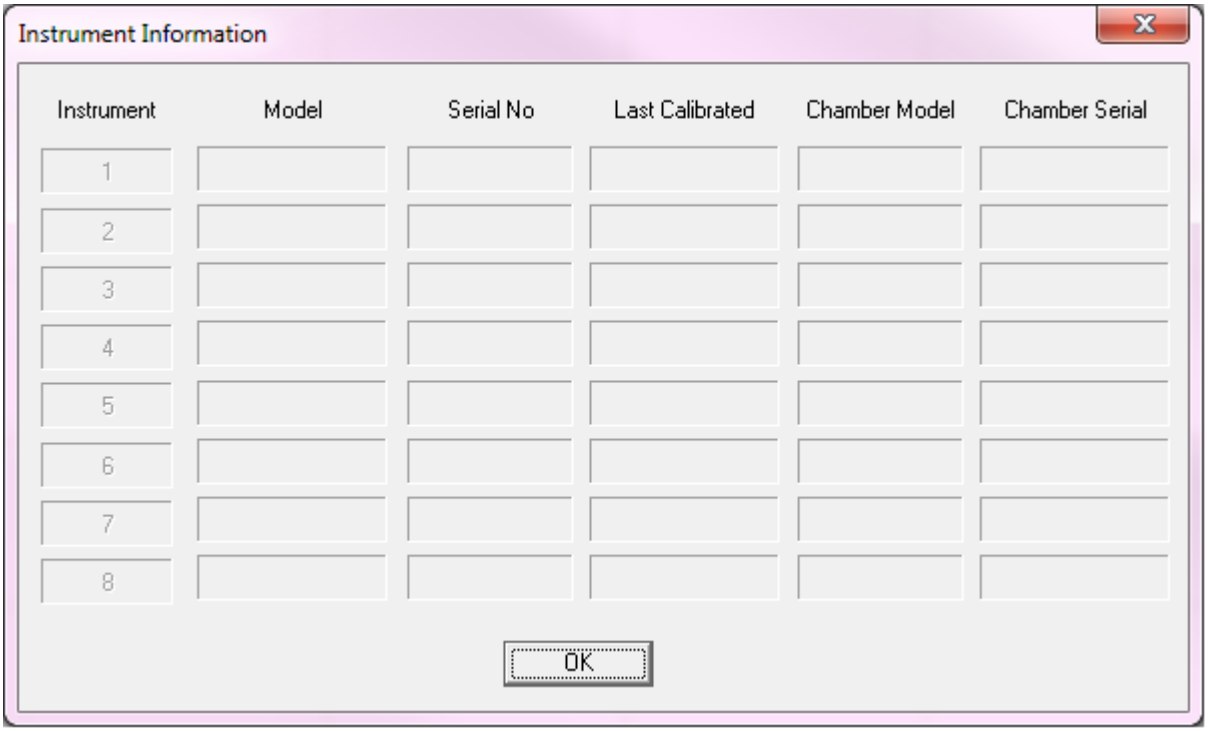

If the above dialogue is displayed then please follow the steps below:

- Ensure that the control unit is connected to a power supply and the ⏻ symbol on the front panel of the unit is flashing intermittently
- Check the USB connection from the control unit to the PC making sure that if a USB hub is being used that the latest drivers have been installed
- If using a USB hub try connecting the control unit directly to a USB port on the PC and restart the software

If you are still experiencing problems with communications, please contact Hansatech Instruments for further assistance.

1.7.2 Box Test

1.7.2.1 Control Unit Diagnostic Tools

In the event of problems with the electrode system such as drifting or unstable signal, system over or under scale or lack of response to changes in oxygen tension, it is necessary to isolate the area of the system that is causing the fault. The electrode system essentially consists of 3 main areas where faults could occur:

- Control unit
- Electrode disc

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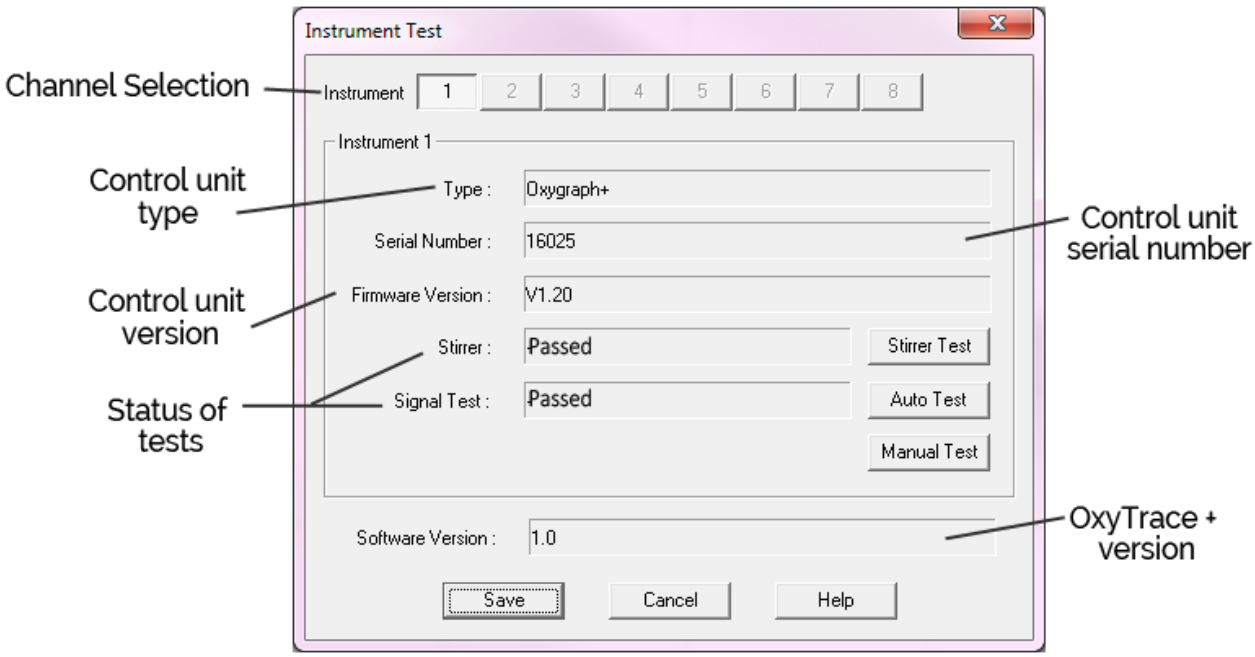
- Connection cable

By checking these areas individually on a step by step basis, faults may be relatively easily diagnosed.

OxyTrace+ features an Instrument Test function which allows the control unit to be subjected to a series of diagnostic tests with controlled parameters in order to ensure that the electronics of the control unit are functioning to within specification. This feature can be accessed by selecting Hardware > Instrument Test from the menu bar.



2 types of test are launched from this window:

- Stirrer Test
- Signal Test

If any of the Test Status boxes read Failed, please contact Hansatech Instruments fur further assistance.

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1.7.2.2 Stirrer Test

The stirrer test diagnostic allows the function of the magnetic stirrer to be tested. The stirrer test diagnostic has 2 modes of function:

- Automatic Test mode
- Manual Test mode

When selecting Automatic Test, OxyTrace+ will set the stirrer at 3 different speeds; Off, Slow & Fast. At each speed, OxyTrace+ requires user input to confirm that the stirrer is either off or rotating at the appropriate speed.

In Manual Test mode, clicking either of the 3 buttons at the bottom of the Test Stirrer dialogue allows the functionality of the stirrer to be verified.

1.7.2.3 Signal Test

The Signal Test diagnostics allow the signal processing electronics within the electrode control unit to be tested. In situations where signal drift, excessive noise or unexpected signal levels are recorded, the Signal Test diagnostic will be able to indicate if there is a genuine problem with the control unit itself.

THE SIGNAL TEST REQUIRES BOTH THE ELECTRODE DISC AND THE ELECTRODE CONNECTION CABLE TO BE DISCONNECTED FROM THE REAR OF THE CONTROL UNIT BEFORE THE TEST IS INITIATED.

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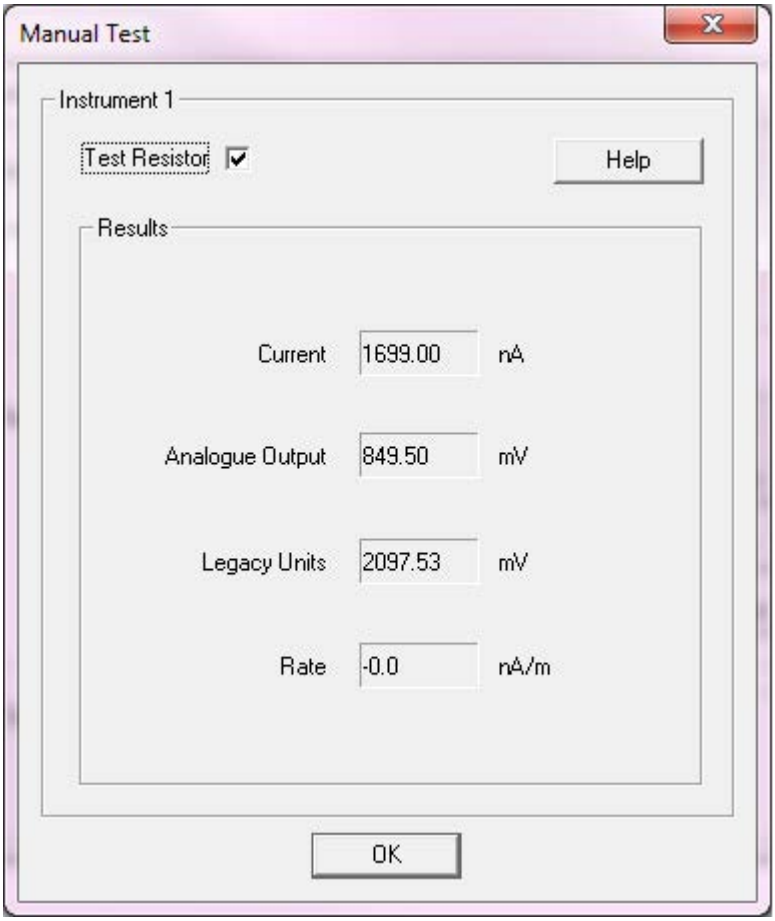
The signal test diagnostic uses a test resistor to replicate an ideal electrode disc response at air line which is approximately 2049mV. During the signal test, OxyTrace+ will set the control unit to different levels of gain and back off and ensure that the signal level responds accordingly.

Manual Test

The diagram below shows the manual test dialogue which is shown when manual test is selected from the box test options. The dialog displays the signal from the box in Current (nA) and the legacy units (mV). The Analogue output is the signal level being output via the phono connector on the rear of the control unit.



Firstly, ensure that the "Test Resistor" box is checked (see above). The Instrument test diagnostic uses a test resistor to replicate an ideal electrode disc response at airline.
After leaving the box for a couple of minutes to stabilise the following can be observed:

Current: This should also be a stable signal at around 1700nA.

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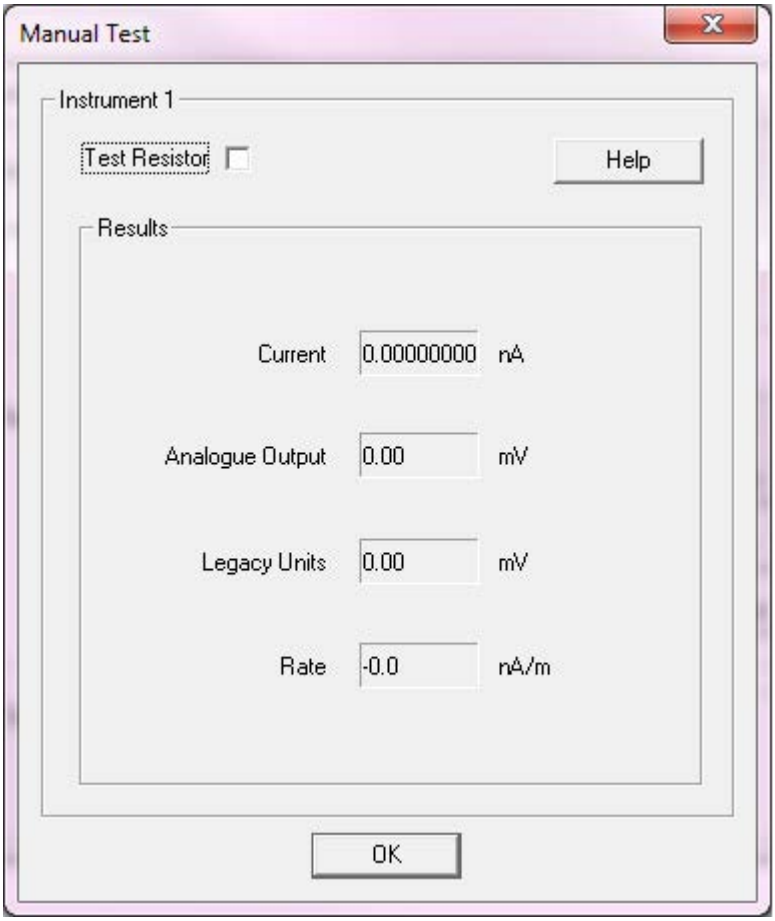
Analogue Output: The signal level being output via the phono connector on the rear of the control unit.
Legacy Units: This signal level should be stable and between 2000-2100mV.

If this information is present and correct now un-check the "Test Resistor" box and leave the unit for a couple of minutes to stabalise you will then be presented similar signal levels to those below:



All signal levels should now be stable with figures close to 0.

If the above is true and your signal levels are both stable and within the ranges stated then you can be assured that your equipment is fully operational and drift free.

If this is not the case then please contact Hansatech Instruments Ltd for assistance.

Auto Test

When Auto Test is selected, OxyTrace+ will conduct a signal test using a set of predefined gain and back off conditions and automatically record the results based on those settings. If the results

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fall within the manufacturers guidelines, the signal test will pass. If the test fails, contact Hansatech Instruments for further advice in this situation.

1.7.3 Electrode Disc Diagnostic Tools

Possible Causes of Signal Drift.

- Inadequate electrode preparation. If the membrane and space paper have not been applied properly, i.e. the membrane layer of the electrode dome is particularly uneven, this could result in an unstable signal as the barrier of electrolyte trapped between the membrane and the cathode is inconsistent. In areas where the electrolyte layer is thicker, oxygen would take longer to diffuse through to the platinum cathode than in areas where the electrolyte layer is normal. Please refer to the Electrode Preparation section for details on correct preparation of the electrode.
- Improper electrode mounting. When the electrode is mounted into the base of the electrode chamber, if the base ring is screwed on too tightly, the membrane will become stretched over the top of the dome. This will result in a higher oxygen diffusion potential through the membrane to he cathode due to the pores in the PTFE becoming stretched/enlarged. As the membrane begins to relax, the speed at which oxygen can diffuse through the membrane to the platinum cathode decreases resulting in a downwards signal drift.
- Temperature control. If the chamber is not correctly temperature controlled or the sample of air saturated water has not been pre-equilibrated to the correct assay temperature, a signal drift will occur.
- Disc condition. If the disc has not been cleaned, there may be deposits on the silver anode which are by-products of the electrochemical reactions taking place on the electrode disc when polarised. In the first instance, a brown deposit of silver chloride may be present. This deposit may, in some cases, be desirable as it is an electrical conductor and may improve the sensitivity of the electrode disc. However, in time, black deposits of silver oxide may build up. This deposit is an electrical insulator and will therefore decrease the surface area of silver available for electrochemistry to occur. The response time of the disc will be greatly affected in worst cases. Please refer to the Electrode Maintenance section for full details on the cleaning procedures for electrode discs.
- Damaged electrode disc. If the disc has not been maintained correctly or there is a genuine fault with the disc, a drifting signal may indicate a problem more permanent than those described above.

If the signal still drifts, please contact Hansatech Instruments for further assistance.

Electrode Signal Increases During Zero Oxygen Stage of Liquid-Phase Calibration.

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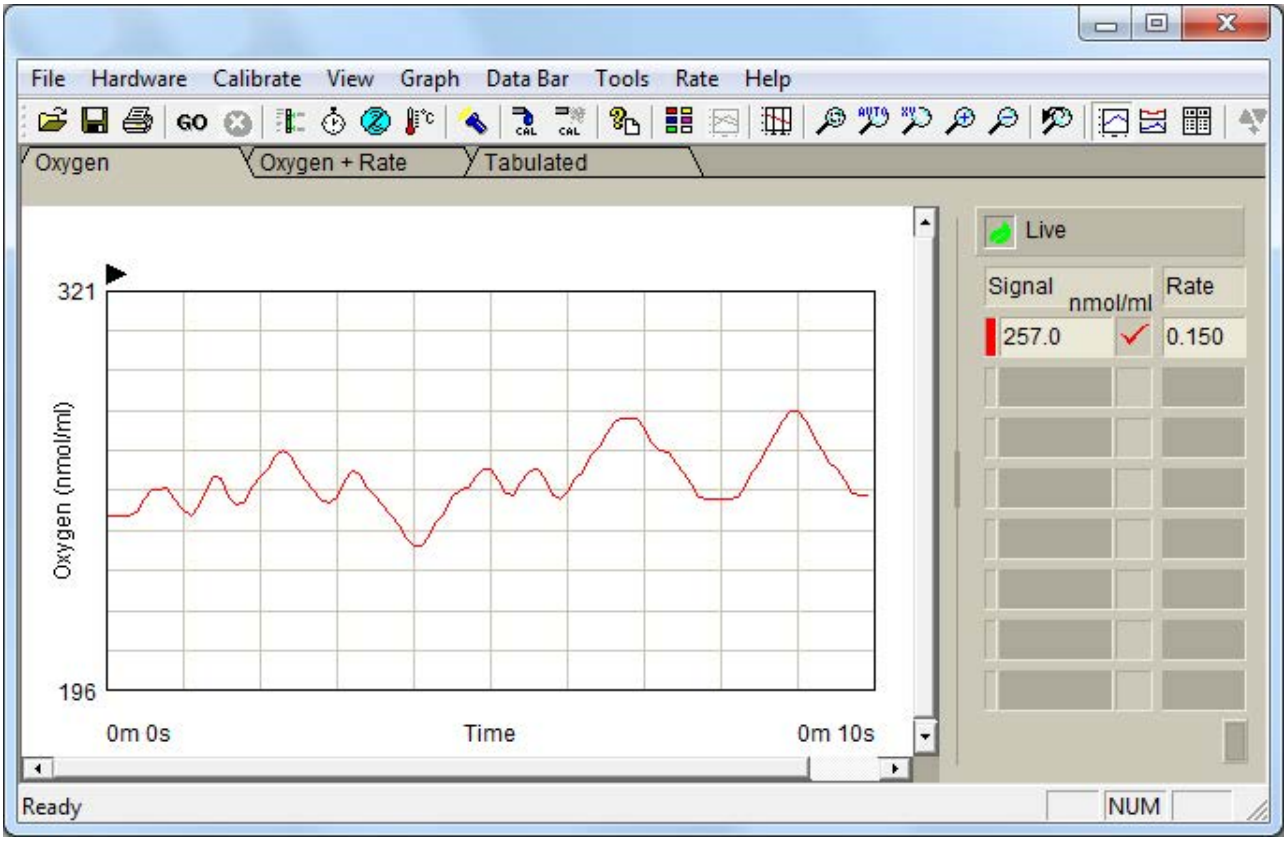
There have been some issues where, during the zero oxygen stage of liquid-phase calibration, the electrode signal decreases correctly following the addition of sodium dithionite. However, instead of reaching a lower plateau, the signal begins to increase above the original airline level. In these cases, it is more than likely that the sodium dithionite is past it's effective shelf-life and therefore ineffective.

Either use a fresh batch of sodium dithionite or repeat the calibration using nitrogen gas to achieve the zero oxygen signal. If the problem still exists, please contact Hansatech Instruments fur further assistance.

Possible Causes of Excessive Signal Noise.

Noisy signals can be caused by the same issues described in the section above. It is necessary to check the electrode disc according to the guidelines listed above. If the signal is still noisy, please contact Hansatech Instruments for further assistance.



1.7.4 Testing the Electrode Connection Cable for Breaks or Electrical Shorts

If the connection cable has an intermittent short circuit, this will be seen on the trace as a sudden rise to a maximum signal followed by a plateau. Checking the mV signal from the electrode disc in

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the Instrument Summary window (Hardware > Instrument Summary) should show that the signal is over scaled at 4095 mV when plateaued. If the signal remains at his level, and a problem with the electrode disc and control unit have been ruled out, it may be a dead short in the connection cable. If the signal only remains at this level temporarily and manipulating the cable restores the signal, the problem may be an intermittent short caused by a cable break.

The connection cable may also be "buzzed out" using a multimeter.

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